



**HAL**  
open science

## **IL-6 supports the generation of human long-lived plasma cells in combination with either APRIL or stromal cell-soluble factors.**

Michel Jourdan, Mailys Cren, N. Robert, Karine Bolloré, Thierry Fest, Christophe Duperray, Fabien Guilloton, Dirk Hose, Karin Tarte, Bernard Klein

### ► To cite this version:

Michel Jourdan, Mailys Cren, N. Robert, Karine Bolloré, Thierry Fest, et al.. IL-6 supports the generation of human long-lived plasma cells in combination with either APRIL or stromal cell-soluble factors.. *Leukemia*, 2014, 28 (8), pp.1647-56. 10.1038/leu.2014.61 . hal-00974531

**HAL Id: hal-00974531**

**<https://univ-rennes.hal.science/hal-00974531>**

Submitted on 23 Jan 2015

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 **IL-6 SUPPORTS THE GENERATION OF HUMAN LONG-LIVED PLASMA CELLS**  
2 **IN COMBINATION WITH EITHER APRIL OR STROMAL CELL SOLUBLE**  
3 **FACTORS**

4  
5 **M Jourdan<sup>1</sup>, M Cren<sup>1</sup>, N Robert<sup>2</sup>, K Bollore<sup>2</sup>, T Fest<sup>3,4</sup>, C Duperray<sup>1,2</sup>, F**  
6 **Guilloton<sup>4</sup>, D Hose<sup>5,6</sup>, K Tarte<sup>3,4</sup> and B Klein<sup>1,2,7</sup>**

7  
8  
9 1 INSERM, U1040, Montpellier, France;

10 2 Centre Hospitalier Universitaire Montpellier, Institute of Research in Biotherapy,  
11 Montpellier, France;

12 3 Pôle Cellules et Tissus, Centre Hospitalier Universitaire, Rennes, France;

13 4 INSERM, U917, Rennes, France;

14 5 Medizinische Klinik und Poliklinik V, Universitätsklinikum Heidelberg, Heidelberg,  
15 Germany;

16 6 Nationales Centrum für Tumorerkrankungen, Heidelberg, Germany;

17 7 Université MONTPELLIER1, UFR Médecine, Montpellier, France;

18  
19 Running title: Obtaining human long-lived plasma cells *in vitro*

20  
21  
22 The authors have no conflict of interest to declare.

23  
24  
25 Corresponding Author:  
26 Bernard Klein  
27 INSERM U1040, Institute for Research in Biotherapy  
28 CHU Montpellier, Hospital St Eloi  
29 Av Augustin Fliche  
30 34295 Montpellier -FRANCE  
31 [bernard.klein@inserm.fr](mailto:bernard.klein@inserm.fr)  
32 tel +33(0) 4 67 33 04 55  
33 fax +33(0) 4 67 33 04 59

34 **ABSTRACT**

35 The recent understanding of plasma cell (PC) biology has been obtained from murine  
36 models mainly. The current concept is that plasmablasts home to the BM and further  
37 differentiate into long-lived PCs (LLPCs). These LLPCs survive for months in contact  
38 with a complex niche comprising stromal cells (SCs) and hematopoietic cells both  
39 producing recruitment and survival factors. Using a multi-step culture system, we  
40 show here the possibility to differentiate human memory B cells into LLPCs surviving  
41 for at least 4 months *in vitro* and producing immunoglobulins continuously. A  
42 remarkable feature is that IL-6 is mandatory to generate LLPCs *in vitro* together with  
43 either APRIL or soluble factors produced by SCs, unrelated to APRIL/BAFF, SDF-1,  
44 or IGF-1. These LLPCs are out of the cell cycle, express highly PC transcription  
45 factors and surface markers.

46 This model shows a remarkable robustness of human LLPCs, which can survive and  
47 produce highly immunoglobulins for months *in vitro* without contact with niche cells,  
48 providing the presence of a minimal cocktail of growth factors and nutrients. This  
49 model should be useful to understand further normal PC biology and its deregulation  
50 in premalignant or malignant PC disorders.

51

52 **Key words:** Plasma cell; B cell; differentiation; microarray

53 **INTRODUCTION**

54 Mature memory plasma cells (PCs), termed long lived plasma cells (LLPCs), are  
55 located in the bone marrow (BM) or mucosa and may survive for years, insuring long-  
56 term immune memory.<sup>1</sup> LLPCs are rare cells (0.25 % of BM cells)<sup>2,3</sup> making their  
57 study difficult in humans. Recent knowledge about their generation and maintenance  
58 was obtained with murine models mainly. After selection of antigen (Ag) specific B  
59 cells in the germinal center, involving mutations in immunoglobulin (Ig) variable  
60 genes and isotype switching, centrocytes differentiate into memory B cells (MBCs) or  
61 plasmablasts (PBs). These PBs migrate to medullary cords, exit into the lymph  
62 through a sphingosine phosphate gradient and get to the peripheral blood.<sup>4</sup>  
63 Circulating PBs have to find a specific niche in the BM or mucosa that will provide  
64 them with the factors to survive and fully differentiate. The restricted number of  
65 suitable PC niches is supposed to be the main limiting factor explaining LLPC rarity.<sup>4</sup>  
66 In mice, Tokoyoda *et al.* have reported the PC niche to be a VCAM1<sup>+</sup> SDF-1<sup>+</sup> stromal  
67 cell that could be shared by hematopoietic progenitors and pre-pro B cells.<sup>5</sup> Several  
68 hematopoietic cell subsets have been described to be involve in PC retention,  
69 maturation, and maintenance in the BM, including macrophages, eosinophils, and  
70 megakaryocytes.<sup>6</sup> At least 3 growth factors and chemokines, produced by the  
71 exquisite PC niche, are recognized to control PC survival: SDF-1, APRIL/BAFF, and  
72 IL-6.<sup>4</sup> SDF-1, produced by SCs is essential to recruit mouse PBs into the BM.<sup>5,7</sup>  
73 BAFF and APRIL are produced by hematopoietic cells and are critical to support PC  
74 survival.<sup>6</sup> BAFF binds to 3 receptors, BAFF receptor, BCMA and TACI, whereas  
75 APRIL binds to BCMA and TACI *in vivo*.<sup>8</sup> Although both APRIL and BAFF can  
76 support murine LLPC survival,<sup>9</sup> APRIL appears to be more efficient than BAFF to  
77 promote LLPC survival, at least in mice. First, APRIL binds to BCMA with a higher

78 affinity than BAFF,<sup>8</sup> and BCMA is highly expressed in PCs.<sup>10</sup> Secondly, APRIL and  
79 its receptor TACI bind to heparan sulfate chains, in particular to the proteoglycan  
80 syndecan-1,<sup>11,12</sup> which is a hallmark of mature PCs.<sup>13</sup> Thirdly, the long-term survival  
81 of transferred LLPCs is impaired in APRIL<sup>-/-</sup> mice, but unaffected in BAFF<sup>-/-</sup> ones.<sup>14</sup>  
82 IL-6, produced by SCs or dendritic cells, is essential for the generation of PBs, and  
83 its role in supporting LLPC survival is controversial in mice. The generation of Ag-  
84 specific PCs is dramatically decreased in IL-6<sup>-/-</sup> mice<sup>15</sup> and the ability of BM SCs to  
85 support the survival of purified BM LLPCs *in vitro* is inhibited by an anti-IL-6  
86 monoclonal antibody (mAb) or lost using IL-6<sup>-/-</sup> mice SCs.<sup>16,17</sup> Similarly, the survival of  
87 LLPCs in the culture of intestine biopsies is impaired by an anti-IL-6 mAb or an  
88 APRIL/BAFF inhibitor.<sup>18</sup> But the persistence of LLPCs transferred into IL-6<sup>-/-</sup> mice is  
89 unaffected.<sup>14</sup>

90 In humans, the factors that promote the differentiation of PBs into LLPCs are poorly  
91 identified *in vivo*, due to the rarity of bone marrow PCs (BMPCs) and the ethical  
92 difficulty to harvest the BM. The majority of the studies dealing with PC survival and  
93 growth were done with malignant PCs.<sup>19,20</sup> Using *in vitro* models of differentiation of B  
94 cells into PCs, we and others have shown that *in vitro* generated PCs are early PCs  
95 with a phenotype close to that of circulating PBs and PCs in healthy individuals.<sup>10,21,22</sup>  
96 These *in vitro* generated PCs expressed highly CD38 and CD31 and lack B cell  
97 antigens (CD20, CD22, CD24) except CD19. They express CD138 but at lower level  
98 than BMPCs, they express CD62L unlike BMPCs, and fail to express CD9, VCAM1,  
99 and CCR2.<sup>10</sup> Recent studies have proposed a role for osteoclasts<sup>23</sup> or SCs to support  
100 human PC survival *in vitro*.<sup>24</sup> In this last study, blood B cells could be differentiated  
101 into Ig-secreting LLPCs in contact with soluble factors produced by a mouse SC line.  
102 However, the molecular mechanisms of this supportive activity remain unknown.<sup>24</sup>

103 In the current study, we show that IL-6 is mandatory for the *in vitro* survival of LLPCs  
104 in combination with either APRIL or BAFF or APRIL/BAFF-unrelated SC soluble  
105 factors. These LLPCs are non-cell cycling, survive for months *in vitro*, while  
106 producing Igs continuously.

## 107 **MATERIALS AND METHODS**

### 108 **Reagents**

109 Human recombinant IL-2, TACI-Fc, and APRIL were purchased from R&D Systems  
110 (Minneapolis, MN), IFN- $\alpha$  (IntronA) from Merck Canada Inc. (Kirckland, Canada), IL-6  
111 and IL-15 from AbCys SA (Paris, France), BAFF, SDF-1 $\alpha$ , IGF-1, and IL-10, from  
112 Peprotech (Rocky Hill, NJ, USA), the B-E8 anti-IL-6 mouse mAb from Diaclone  
113 (Besançon, France), the SDF-1 inhibitor AMD3100 from Sigma (Sigma-Aldrich, St  
114 Louis, MO), the IGF-1R inhibitor from Novartis Pharma (Basel, Switzerland), and  
115 IKK16, a selective inhibitor targeting both IKK1 and IKK2, from R&D Systems.

### 116 **Cell samples**

117 Peripheral blood cells from healthy volunteers were purchased from the French  
118 Blood Center (Toulouse, France) and CD19<sup>+</sup>CD27<sup>+</sup> MBCs purified ( $\geq$  95% purity) as  
119 described.<sup>10</sup> When indicated, D10 early PCs (CD20<sup>-</sup>CD138<sup>+</sup>) were FACS-sorted  
120 using FITC-conjugated anti-CD20 mAb and PE- or APC-conjugated anti-CD138  
121 mAb. The purity of FACS-sorted cell populations was  $\geq$  95% as assayed by  
122 cytometry. Resto-6 stromal cells were used as a source of SCs. These SCs were  
123 obtained from a 15%/25% Percoll interface of dissociated human tonsil cells as  
124 previously described.<sup>25</sup> Plastic-adherent cells were selected and expanded in RPMI  
125 1640 culture medium and 10% fetal calf serum (FCS) yielding to the Resto-6 SCs  
126 after 8 passages. Resto-6 SCs express usual mesenchymal stromal cell markers  
127 (CD90, CD73, and CD105) and can acquire properties of fibroblastic reticular cells  
128 (FRC) including expression of high levels of adhesion molecules and  
129 gp38/podoplanin, production of a dense meshwork of transglutaminase, and  
130 production of inflammatory and lymphoid chemokines upon stimulation by TNF $\alpha$  and  
131 Lymphotoxin- $\alpha$ 1 $\beta$ 2.<sup>25</sup> Resto-6 SCs support efficiently the growth and survival of

132 normal B and T cells and of malignant lymphoma B cells, in particular after FRC-  
133 commitment. They were used between passages 8 and 15.

#### 134 **Cell cultures**

135 PCs were generated through a four-step culture. All cultures were performed in  
136 Iscove's modified Dulbecco medium (IMDM, Invitrogen) and 10% FCS. In step 1,  
137 purified peripheral blood MBCs ( $1.5 \times 10^5/\text{ml}$ ) were activated for 4 days by CpG  
138 oligodeoxynucleotide and CD40 ligand (sCD40L) - 10  $\mu\text{g}/\text{ml}$  of phosphorothioate  
139 CpG oligodeoxynucleotide 2006 (Sigma), 50 ng/ml histidine tagged sCD40L, and  
140 anti-poly-histidine mAb (5  $\mu\text{g}/\text{ml}$ ), (R&D Systems) - with IL-2 (20 U/ml), IL-10 (50  
141 ng/ml) and IL-15 (10 ng/ml) in 6 well culture plates. In step 2, PBs were generated by  
142 removing CpG oligonucleotides and sCD40L and changing the cytokine cocktail (IL-  
143 2, 20 U/ml, IL-6, 50 ng/ml, IL-10, 50 ng/ml and IL-15, 10 ng/ml). In step 3, PBs were  
144 differentiated into early PCs adding IL-6 (50 ng/ml), IL-15 (10 ng/ml) and IFN- $\alpha$ 500  
145 U/ml) for 3 days. In step 4, early PCs were differentiated into LLPCs using either  
146 coculture with SCs, transwell culture, or a cytokine cocktail, and the cultures  
147 maintained for months. Confluent monolayers of SCs were generated in 6-, 24-, or  
148 48-well flat-bottom culture plates and PCs were then added onto the monolayers  
149 together with various cytokines. These cocultures of PCs and SCs could be  
150 maintained for months with the same SC monolayer, adding fresh culture medium  
151 and growth factors once by week. Cocultures of PCs and SCs without cell contact  
152 were done seeding SCs in the lower chamber of 6-well transwells and PCs in the  
153 upper compartment, both compartments being separated by a 0.4  $\mu\text{m}$ -polycarbonate  
154 membrane (Corning, New-York, NY). SC-conditioned medium (SC-CM) was obtained  
155 by culturing confluent monolayers of SCs for 5 days with culture medium. The culture  
156 supernatant was 0.2  $\mu\text{M}$  filtered and frozen and 50% of SC-CM was added to PC

157 cultures, and renewed every week. Finally, LLPCs were also obtained adding IL-6  
158 (10 ng/ml) and either APRIL (200 ng/ml) or BAFF (200 ng/ml).

### 159 **Assay for cell viability and cell growth**

160 Cell concentration and viability were assessed using trypan blue dye exclusion test.  
161 The number of metabolic active cells was also determined using intracellular ATP  
162 quantitation with a Cell Titer Glo Luminescent Assay (Promega Corporation,  
163 Madison, WI).

### 164 **Cell cycle analysis, immunophenotypic analysis, and cytology**

165 The cell cycle was assessed using DAPI staining (Sigma-Aldrich) and cells in the S  
166 phase using incubation with bromodeoxyuridine (BrdU) for 1 hour and labelling with  
167 an anti-BrdU antibody (APC BrdU flow kit, BD Biosciences) according to  
168 manufacturer's instructions. Cells were stained with a combination of 4 to 7 mAbs  
169 conjugated to different fluorochromes. The Cytotfix/Cytoperm kit (BD Biosciences)  
170 was used for intracellular staining of IgM, IgA, IgG or Ki67 antigen.<sup>10</sup> Flow cytometry  
171 analysis was performed with a FACS Aria cytometer using FACSDiva 6.1 (Becton  
172 Dickinson, San Jose, CA) and with a Cyan ADP cytometer driven by the Summit  
173 software (Beckman Coulter). Kaluza software (Beckman Coulter) was used for data  
174 analysis. The fluorescence intensity of the cell populations was quantified using the  
175 stain index (SI) formula: [mean fluorescence intensity (MFI) obtained from a given  
176 mAb minus MFI obtained with a control mAb]/[2 times the standard deviation of the  
177 MFI obtained with the same control mAb].<sup>10</sup> Cytospin smears of cell-sorted CD20<sup>-</sup>  
178 CD138<sup>+</sup> D30 PCs were stained with May-Grünwald-Giemsa.

### 179 **Analysis of Ig secretion**

180 **ELISA.** Flow cytometry sorted PCs were cultured at 10<sup>6</sup> cells/ml for 24 hours and  
181 culture supernatants harvested. IgM, IgA, or IgG concentrations were assessed by

182 ELISA using human IgM, IgA, and IgG ELISA kits from Bethyl Laboratories  
183 (Montgomery, TX), according to the manufacturer's recommendations.

184 **ELISPOT.** The number of IgM, IgA, or IgG secreting PCs was evaluated with the  
185 ELISPOT assay,<sup>26</sup> seeding 500 PCs by well in ELISPOT plates and culturing cells  
186 for 18 hours. The number and size of IgM, IgA, and IgG elispots were assessed  
187 using the Biosys Bioreader 5000 apparatus (Biosys, Miami, FL).

#### 188 **Microarray hybridization and bioinformatics analysis**

189 RNA was extracted and hybridized to human genome U133 Plus 2.0 GeneChip  
190 microarrays, according to the manufacturer's instructions (Affymetrix, Santa Clara,  
191 CA). Gene expression data are deposited in the ArrayExpress public database  
192 (<http://www.ebi.ac.uk/microarray-as/ae/>). The accession numbers are E-MEXP-3034  
193 for prePBs, E-MEXP-2360 for PBs and BMPCs, E-MEXP-3945 for day 10 (D10) early  
194 PCs and D30 PCs, and E-MTAB-2118 for Resto-6 SCs. Gene expression data were  
195 analyzed with our bioinformatics platforms (RAGE, <http://rage.montp.inserm.fr/>)<sup>27</sup> and  
196 Amazonia (<http://amazonia.transcriptome.eu/>).<sup>28</sup> Genes differentially expressed  
197 between cell populations were determined with the SAM statistical microarray  
198 analysis software.<sup>29</sup> The clustering was performed and visualized with the Cluster  
199 and TreeView softwares.<sup>30</sup> Gene annotation and networks were generated with the  
200 Reactome Functional Interaction Cytoscape plugin (<http://www.cytoscape.org/>).

#### 201 **Statistical analysis**

202 Statistical comparisons were made with the non-parametric Mann-Whitney test,  
203 unpaired or paired Student's *t*-test using SPSS software. *P*-values  $\leq .05$  were  
204 considered as significant.

205 **RESULTS**

206 **IL-6 in combination with APRIL or BAFF or APRIL/BAFF-unrelated stromal-cell**  
207 **soluble factors supports the generation and survival of PCs *in vitro***

208 Starting from MBCs, early PCs can be generated within 10 days using a 3-step  
209 culture. These early PCs could survive poorly in presence of IL-6 (Figure 1A). Adding  
210 human tonsil SCs could promote PC survival with 28% surviving CD138 PCs at day  
211 30 (Figure 1A). In the presence of SCs, the CD138 fluorescence staining index at  
212 day 14 was increased 3 fold ( $P = .006$ ) compared to day 14 PCs generated with IL-6  
213 alone (Figure 1B and 1C). It progressively increased 2 fold from day 14 to day 30 ( $P$   
214  $= .032$ ) and then was stable up to day 60 (Figures 1B and 1C). Coculturing early PCs  
215 separated from SCs in transwell culture plates promoted PC survival at the same  
216 extent as coculture of PCs with SCs at day 14, 17, 24 and 30, showing a contact  
217 between PCs and SCs is not mandatory (Figure 2A). In agreement, coculture  
218 supernatant of PCs with SCs (PC/SC-CM) or of SCs alone (SC-CM) also promoted  
219 PC survival at day 30 (Figure 2A). Various growth recombinant factors - APRIL,  
220 BAFF, SDF-1, IGF-1 - known to sustain malignant PC survival<sup>19</sup> did not induce PC  
221 survival when used alone (Figure 2B). IL-6 alone supported PC survival but at a 76%  
222 lower level than that induced by the SC-CM ( $P = .003$ ). Adding APRIL together with  
223 IL-6 increased significantly 6.5 fold ( $P = .02$ ) the IL-6 PC survival activity (Figure 2B).  
224 Adding BAFF also increased the PC survival induced by IL-6 ( $P = .02$ ) and SDF-1  
225 and/or IGF-1 did not increase IL-6 activity (Figure 2B). The PC survival induced by  
226 the SC-CM was fully abrogated by an anti-IL-6 mAb showing the critical role of IL-6  
227 produced by SCs, but was unaffected by an APRIL/BAFF inhibitor, TACI-Fc (Figure  
228 2C). The TACI-Fc APRIL/BAFF inhibitor fully blocked the ability of APRIL to increase  
229 2.2 fold the PC generation induced by the SC-CM (Figure 2C). It also inhibited the

230 APRIL-induced growth of the XG1 myeloma cell line (supplementary Figure 1).<sup>31</sup> As  
231 APRIL/BAFF activate NF- $\kappa$ B pathways,<sup>32</sup> we investigated whether these pathways  
232 could be involved in PC generation by SCs. An inhibitor of both canonical and  
233 alternative NF- $\kappa$ B pathways (IKK16) did not affect the generation of PCs supported  
234 by SCs, whereas it impeded the additive effect of APRIL in PC generation promoted  
235 by SCs (Figure 3). Thus, SCs produce communication signals, which cooperate with  
236 IL-6 to promote PC survival and are not inhibited by a NF- $\kappa$ B inhibitor. This is in  
237 agreement with the low expression of NF- $\kappa$ B-induced genes<sup>33</sup> in PCs generated with  
238 SCs *in vitro* contrarily to BMPCs (supplementary Figure 2). These communication  
239 signals are not SDF-1 or IGF-1 since the SC-CM activity was unaffected by their  
240 specific inhibitors (Figure 2C). Adding these 3 inhibitors together (APRIL/BAFF, SDF-  
241 1 and IGF-1 inhibitors) yielded to an apparent decrease in SC-CM induced PC  
242 generation, which did not reach statistical significance (Figure 2C).  
243 Of note, PCs could survive up to 120 days in these culture conditions adding fresh IL-  
244 6, APRIL and culture medium weekly and cultures were stopped after 120 days for  
245 convenience but not due to a decline in PC survival. These PCs were thus termed  
246 LLPCs. These results indicate that IL-6 is mandatory to induce the long-term survival  
247 of LLPCs in combination with either APRIL or BAFF or with APRIL/BAFF-unrelated  
248 soluble factors produced by SCs.

#### 249 **Characterization of D30 PCs**

250 D30 PCs had a more mature PC phenotype than D10 early PCs with a progressive  
251 7.4-fold increase in CD138 density (Figure 1C,  $P \leq .001$ ), a 2.2-fold increase in CD54  
252 staining index, a 1.5-fold increase in CD9<sup>+</sup> PCs, and a decrease in HLA-DR, CD45,  
253 CD62L, CCR10 expressing PCs and staining indexes ( $P \leq .05$ , Figure 4 and  
254 supplementary Figure 3). D30 PCs did not proliferate (0.04% cells incorporating

255 BrdU), whereas a low fraction of D10 early PCs was in the S phase of the cell cycle  
256 (4% BrdU+) (Figure 5A and 5B). D30 PCs displayed the cytology of mature PCs and  
257 produced cytoplasmic kappa or lambda Ig light chains (supplementary Figure 4). Day  
258 60 PCs comprise IgG PCs only, with a quick disappearance of IgM PCs (no more  
259 detectable at day 30) and a progressive one of IgA PCs as assayed by FACS, ELISA  
260 and ELISPOT assays (Figures 6 A-F). Of note, the rate of IgG production was similar  
261 between day 30 and day 60 PCs (Figure 6F). The number of IgG and IgM producing  
262 cells and the Ig isotype produced were not significantly different starting from the  
263 same number of LLPCs generated with either SC coculture, IL-6+APRIL, SC-CM or  
264 APRIL+SC-CM (Figure 6G). There was a trend in increasing IgA secreting cells in the  
265 two experiments adding APRIL compared to the culture groups without APRIL ( $P <$   
266  $.1$ ). Combining data of these two experiments show APRIL increased 3.4 fold the  
267 number of IgA secreting cells (47 vs. 14 IgA secreting cells/500 PCs,  $P < .001$ , Figure  
268 6G).

#### 269 **B and PC transcription factors**

270 D30 PCs had a higher expression of genes coding for IRF4 and BLIMP1 PC  
271 transcription factors than D10 early PCs and failed to express *PAX5* ( $P \leq .02$ , Figure  
272 7). D30 PCs expressed 3-fold more the spliced form of *XBP1* mRNA than D10 early  
273 PCs and 2-fold more the unspliced mRNA form than early PCs, resulting in an  
274 increased *XBP1s/XBP1u* mRNA ratio ( $P \leq .009$ , Figure 7). Of note, *BCL6* gene  
275 expression in D30 PCs was low compared to that in B lymphocytes but significantly  
276 higher than that in early PCs ( $P = .04$ , Figure 7).

#### 277 **Gene expression profile (GEP) of D30 PCs**

278 *In vitro* generated D4 prePBs, D7 PBs, D10 early PCs, D30 PCs, and BMPCs  
279 purified from healthy individuals were profiled using Affymetrix U133 plus 2.0

280 microarrays. D30 PCs were generated using cocultures with SCs. The 5 populations  
281 are classified into 2 major clusters, a PC cluster comprising D10 early PCs, D30 PCs,  
282 and BMPCs and a PB cluster comprising prePBs and PBs (Figure 8). To look for  
283 genes indicator of LLPCs, we ran a SAM supervised analysis comparing D10 early  
284 PCs to D30PCs+BMPCs starting from the 5000 genes with the highest variance. 160  
285 probe sets (141 unique genes) were overexpressed in D30PCs+BMPCs compared to  
286 D10 early PCs (Wilcoxon statistic, fold change  $\geq 2$ , FDR  $\leq 1\%$ ) and 490 (427 unique  
287 genes) in D10 early PCs versus D30PCs+BMPCs (supplementary Table 1). Genes  
288 coding for translation, focal adhesion, IL-6 signalling, and integrin signalling pathways  
289 were enriched in D30PC+BMPC genes (supplementary Table 2), and genes coding  
290 for DNA replication and mitosis in D10 early PCs (supplementary Table 3). The gene  
291 expression profiles of LLPCs harvested from the spleen of patients with primary  
292 immune thrombocytopenia treated with rituximab anti-CD20 mAb was recently  
293 documented and compared to that of PBs harvested from the spleen of untreated  
294 patients.<sup>34</sup> Similarly to the current *in vitro* generated LLPCs, LLPCs from these  
295 patients overexpress genes coding for PC transcription factors (*JUN*, *FOS*, *EGR1*),  
296 negative regulators of the cell cycle (*KLF4*, *KLF2*, *PPP1R15A*) and cell  
297 surface/cytokine receptors (*CD9*, *SDC1*, *FCRL5*). Conversely, similarly to *in vitro*-  
298 generated D10 early PCs, patients' PBs overexpress genes coding for positive cell  
299 cycle regulators (*CCND2*, *BUB1B*, *BUB1*, *TIMELESS*, *CENPF*, *MAD2L1*, *BIRC5*,  
300 *ZWINT*, *MKI67*, *MCM4*, *CCNB2*) and surface/cytokine receptors (*ITGB1*, *TNFSF10*)  
301 (supplementary Table 1). Comparing gene expression between BMPCs and D30  
302 PCs, 198 unique genes were overexpressed in BMPCs and 555 in D30 PCs (SAM  
303 supervised analysis, Wilcoxon statistic, fold change  $\geq 2$ , FDR  $\leq 1\%$ , supplementary  
304 Table 4). Genes coding for protein metabolism, translation, antigen processing and

305 presentation, and CXCR4 signalling were enriched in BMPC genes (supplementary  
306 Table 5). Genes coding for glypican pathway, TGF $\beta$  receptor and Smad2/3  
307 signalling, protein export and proteasome were enriched in D30 PC genes  
308 (supplementary Table 6). GEP were done using D30 PCs generated with SCs and  
309 IL-6. It could be of interest to investigate further whether adding APRIL together with  
310 IL-6 or SC-CM could change the gene expression profiling, making it closer to that of  
311 BMPCs.

## 312 **DISCUSSION**

313 This study shows i) the feasibility to generate human mature PCs *in vitro*, ii) the long-  
314 term survival of these PCs does not require a contact with niche cells, but only cell  
315 communication factors, in particular IL-6 and APRIL.

316 These PCs are called long-lived PCs because they are non-cycling PCs surviving  
317 and producing Igs for months *in vitro* as their counterpart *in vivo*.<sup>1,35,36</sup> In addition,  
318 they have a phenotype similar to that of LLPCs *in vivo*: high expression of CD138,  
319 increased expression of CD9, weak expression of CD62L, CD45 and HLA-DR  
320 compared to PBs and early PCs.<sup>22,37</sup> These *in vitro* generated LLPCs expressed  
321 *IRF4* and *PRDM1* genes coding for PC transcription factors at a higher level than  
322 early PCs. Murine LLPCs also highly express Blimp1 compared to early PCs in the  
323 BM.<sup>38</sup> XBP1 is a master regulator of unfold protein response critical to protect PCs  
324 from stress induced by high Ig production.<sup>39</sup> *XBP1* mRNA has to be spliced to  
325 encode for an active protein and, in agreement, we found that LLPCs had an  
326 increased ratio of spliced to unspliced *XBP1* mRNAs compared to early PCs. LLPCs  
327 expressed weakly but significantly *BCL6* gene compared to early PCs. This deserves  
328 further study since Bcl6 is also inducible in malignant PCs in response to SC-derived  
329 factors, conferring a survival advantage on them.<sup>2</sup>

330 A second major finding is that the generation and survival of human LLPCs do not  
331 require a contact with niche cells *in vitro*, but can be obtained with 2 recombinant  
332 growth factors only, IL-6 and APRIL. APRIL can be replaced by BAFF, which  
333 activates the same receptors. In addition, APRIL can be replaced by APRIL/BAFF-  
334 unrelated soluble factors produced by SCs. Of note, an inhibitor of both the canonical  
335 and alternative NF- $\kappa$ B pathways did not affect the generation of LLPCs supported by  
336 SCs, whereas it abrogated the additive effect of APRIL in getting LLPCs with SCs. In

337 addition, a set of genes, whose expression is induced by NF- $\kappa$ B pathway activation in  
338 malignant PCs,<sup>33</sup> is poorly expressed in LLPCs generated with SCs *in vitro* compared  
339 to BMPCs as previously mentioned by Cocco *et al.*<sup>24</sup> Thus the activation of the NF-  
340  $\kappa$ B pathway is not mandatory to generate LLPCs *in vitro*, but could enhance it. The  
341 current data are in line with recent findings showing that a combination of APRIL,  
342 BAFF, IGF-1, SDF-1 and VEGF can support modestly the *in vitro* 14-day survival of  
343 human PCs harvested from the BM, whereas STAT3 activating cytokines, in  
344 particular IL-6, are critical.<sup>40</sup>

345 Using more complex culture conditions, including IL-6, IL-21, IFN- $\alpha$  and SC-CM, a  
346 recent study has shown human LLPCs can be generated *in vitro* also.<sup>24</sup> That the *in*  
347 *vitro* long-term survival of human PCs can be supported by 2 growth factors (IL-6 and  
348 APRIL) only is quite surprising regarding the current view of the complexity of the PC  
349 niche, comprising SCs and various hematopoietic cells (eosinophils, dendritic cells,  
350 megakaryocytes, neutrophils, basophils).<sup>6,14,41</sup> SCs are thought to serve as docking  
351 cells bringing close together PCs and hematopoietic cells,<sup>14</sup> but also producing  
352 soluble factors promoting PC survival, in particular IL-6 and galectin.<sup>42,43</sup> The ability  
353 of hematopoietic cells to sustain PC survival is due mainly by their ability to produce  
354 APRIL.<sup>9,11</sup>

355 The fact that soluble growth factors can replace niche cells for the generation and  
356 survival of LLPCs *in vitro* suggests it could be the case *in vivo* and questions about  
357 the regulation of LLPC count *in vivo*. It is generally assumed that the tiny PC count in  
358 the BM is regulated by the rarity of BM niche cells<sup>5</sup>, new PCs being in competition  
359 with old ones for the availability of niche cells.<sup>44</sup> The current finding suggests the PC  
360 niche is mainly a liquid niche comprising a life-sustaining mixture and concentration  
361 of chemokines and growth factors, which is likely the case close to the docking SCs

362 attracting both PCs and hematopoietic cells.<sup>14</sup> When entering the BM, if a PC cannot  
363 migrate close to a SC, it will not encounter the life-sustaining concentrations of  
364 soluble factors and die. But in case of deregulated production of these cell  
365 communication signals such as in inflammatory conditions, one can expect many  
366 PCs may survive *in vivo*. This may explain the accumulation of LLPCs in the spleen  
367 of patients with primary immune thrombocytopenia treated with Rituximab anti-CD20  
368 mAb, in association with a 2-fold increase in BAFF concentration in the spleen  
369 compared to Rituximab untreated patients.<sup>34</sup> Besides genomic abnormalities, this  
370 could also explain the progressive accumulation of premalignant PCs and then  
371 malignant PCs in patients with malignant PC disorders who display increased plasma  
372 concentrations of IL-6,<sup>45</sup> APRIL or BAFF.<sup>46</sup>

373 The current finding of a mandatory role of IL-6 to promote the survival of human  
374 LLPCs *in vitro* questions the role of IL-6 for LLPC maintenance. In mice, whereas IL-  
375 6 produced by SCs is mandatory to get the survival of BMPCs *in vitro*,<sup>16,17</sup> the  
376 survival of transferred LLPCs is not impaired in *IL-6*<sup>-/-</sup> mice unlike *APRIL*<sup>-/-</sup> ones.<sup>14</sup>  
377 This is likely due to bias in the murine or human models used. In particular, whereas  
378 LLPCs can not be transferred in *APRIL*<sup>-/-</sup> mice<sup>14</sup>, Ag-specific PCs can be generated in  
379 *APRIL*<sup>-/-</sup> mice using repeated Ag boosts indicating additional factors can replace  
380 APRIL.<sup>14</sup> In *IL-6*<sup>-/-</sup> mice, a role of the other cytokines able to trigger gp130 IL-6  
381 transducer chain and/or STAT3 activation and to supplement for a deficit in IL-6  
382 induced signalling to support PC survival has not been evaluated. This is the case in  
383 humans since a recent study has emphasized that inhibition of STAT3 activation by  
384 small compounds can fully block the *in vitro* 14-day survival of human PCs harvested  
385 from the peripheral blood or BM of healthy individuals. STAT3 activation in these PCs  
386 could be driven by either IL-6, IL-10 or IL-21.<sup>40</sup>

387 These *in vitro* models to get human PCs likely introduce some bias, in particular by  
388 the method to activate B cells (through BCR, CD40, or TLR), the combination of  
389 cytokines used to generate prePBs, PBs, early PCs and then LLPCs, the origin of  
390 stromal cells and the culture conditions influencing PC metabolism (nutrients,  
391 glucose, O<sub>2</sub> concentration).<sup>10,21,24,26</sup> In the current model, we used a stromal cell line  
392 obtained from tonsils because it grow easily until confluence and at confluence, can  
393 survive for several months without proliferating but providing a continuous SC  
394 support. In initial experiments, similar data were obtained in term of phenotype and  
395 long-term survival with BM SCs. But it is of major interest to investigate further  
396 whether the use of SCs from different tissue origins could change the phenotype and  
397 gene expression profiling of LLPCs, in particular their proximity with BMPCs. For  
398 example, a progressive loss of IgA secreting PCs occurred in cultures with tonsil SCs  
399 or SC-CM likely due to the lack of a critical survival factor for IgA PC survival *in vitro*.  
400 Adding APRIL can revert this loss, increasing 3.4 fold the survival of IgA secreting  
401 PCs and this could eventually also occur with BM SCs. The ease of the current  
402 model to get PCs *in vitro* will make possible further identification of these possible  
403 biases.

404 All prePBs and PBs generated in this model express CD19 whereas Chaidos *et al.*<sup>47</sup>  
405 reported recently the existence of CD19<sup>-</sup>CD38<sup>+</sup>CD138<sup>-</sup> plasmablasts (called Pre-  
406 PCs) in healthy individuals, together with the known CD19<sup>+</sup>CD38<sup>+</sup>CD138<sup>-</sup> PBs and  
407 CD19<sup>+</sup>CD38<sup>+</sup>CD138<sup>+</sup> PCs.<sup>22</sup> The malignant counterpart of these Pre-PCs are found  
408 in patients with MM. Running a supervised analysis of gene expression profiling of  
409 malignant Pre-PCs and PCs, Chaidos *et al.* found enrichment of genes coding for  
410 epigenetic pathways.<sup>47</sup> The *in vitro* model we used likely failed to generate these  
411 CD19<sup>-</sup>CD38<sup>+</sup>CD138<sup>-</sup> since all prePBs generated at day 4, PBs at day 7 and PCs at

412 day 10 express CD19.<sup>10,26</sup> In addition, the epigenetic genes differentially expressed  
413 between malignant Pre-PCs and PCs could not classify the current *in vitro* generated  
414 PBs, early PCs and LLPCs (data not shown).  
415 Besides its interest for understanding the fine pathways controlling PC generation  
416 and survival in humans, the current model should be promising to study the  
417 mechanisms involved in malignant PC disorders and controlling the activity of drugs  
418 used to treat patients with these disorders. As several genes whose expression or  
419 abnormalities are associated with disease activity have been identified,<sup>48-55</sup> their  
420 modulation throughout the different stages of PC generation (prePBs, PBs, early  
421 PCs, LLPCs) could help to understand better their function. Of note, it is now feasible  
422 to force or repress the expression of a given gene in these PCs using measles  
423 envelop pseudotyped lentiviral delivery.<sup>56</sup> The same holds true for drugs used to treat  
424 patients with MM, in particular to identify if these drugs could target a specific PB or  
425 PC stage and the underlying mechanism.

426 **ACKNOWLEDGEMENTS**

427 This work was supported by grants from ARC (SL220110603450, Paris France), from  
428 ANR ([2012-109/087437](https://doi.org/10.1007/978-2-287-08743-7)), and the European Community (FP7-OVERMYR). We thank  
429 Mrs Pantesco from the Microarray Core Facility of IRB  
430 (<http://irb.montp.inserm.fr/en/index.php?page=Plateau&IdEquipe=6>), Dr Duperray  
431 from the cytometry platform of IRB  
432 (<http://irb.montp.inserm.fr/en/index.php?page=Plateau&IdEquipe=3>, Montpellier Rio  
433 Imaging) and Dr Reme from the IRB Bioinformatics platform  
434 (<http://irb.montp.inserm.fr/en/index.php?page=Plateau&IdEquipe=18>).

435

436 **AUTHOR CONTRIBUTIONS**

437 MJ designed research, performed the experiments and wrote the paper.  
438 KT, TF and FG, provided stromal cells and corrected the paper.  
439 MC, NR, and KB performed the experiments.  
440 CD provided assistance for cytometry experiments.  
441 DH provided data of gene expression profiling of BMPCs.  
442 BK is the senior investigator who designed research and wrote the paper.

443

444 **CONFLICT OF INTEREST**

445 The authors have no conflict of interest to declare.

446

447 Supplementary information is available at Leukemia's website.

448 **REFERENCES**

- 449 1 Radbruch A, Muehlinghaus G, Luger EO, Inamine A, Smith KGC, Dörner T, *et*  
450 *al.* Competence and competition: the challenge of becoming a long-lived  
451 plasma cell. *Nat Rev Immunol* 2006; **6**: 741–750.
- 452 2 Hideshima T, Mitsiades C, Ikeda H, Chauhan D, Raje N, Gorgun G, *et al.* A  
453 proto-oncogene BCL6 is up-regulated in the bone marrow microenvironment in  
454 multiple myeloma cells. *Blood* 2010; **115**: 3772–3775.
- 455 3 Terstappen L, Steen J, Seger-Nolten MJ, Loken MR. Identification and  
456 characterization of plasma cells in normal bone marrow by high resolution flow  
457 cytometry. *Blood* 1990; **9**: 1739–1747.
- 458 4 Tangye SG. Staying alive: regulation of plasma cell survival. *Trends Immunol*  
459 2011; **32**: 595–602.
- 460 5 Tokoyoda K, Egawa T, Sugiyama T, Choi BI, Nagasawa T. Cellular niches  
461 controlling B lymphocyte behavior within bone marrow during development.  
462 *Immunity* 2004; **20**: 707–718.
- 463 6 Chu VT, Berek C. The establishment of the plasma cell survival niche in the  
464 bone marrow. *Immunol Rev* 2013; **251**: 177–188.
- 465 7 Hargreaves DC, Hyman PL, Lu TT, Ngo VN, Bidgol A, Suzuki G, *et al.* A  
466 coordinated change in chemokine responsiveness guides plasma cell  
467 movements. *J Exp Med* 2001; **194**: 45–56.
- 468 8 Vincent FB, Saulep-Easton D, Figgett WA, Fairfax KA, Mackay F. The  
469 BAFF/APRIL system: Emerging functions beyond B cell biology and

- 470 autoimmunity. *Cytokine Growth Factor Rev* 2013; **24**: 203–215.
- 471 9 Benson MJ, Dillon SR, Castigli E, Geha RS, Xu S, Lam K-P, *et al.* Cutting  
472 edge: the dependence of plasma cells and independence of memory B cells on  
473 BAFF and APRIL. *J Immunol* 2008; **180**: 3655–3659.
- 474 10 Jourdan M, Caraux A, De Vos J, Fiol G, Larroque M, Cognot C, *et al.* An in  
475 vitro model of differentiation of memory B cells into plasmablasts and plasma  
476 cells including detailed phenotypic and molecular characterization. *Blood* 2009;  
477 **114**: 5173–5181.
- 478 11 Huard B, McKee T, Bosshard C, Durual S, Matthes T, Myit S, *et al.* APRIL  
479 secreted by neutrophils binds to heparan sulfate proteoglycans to create  
480 plasma cell niches in human mucosa. *J Clin Invest* 2008; **118**: 2887–2895.
- 481 12 Moreaux J, Sprynski A-C, Dillon SR, Mahtouk K, Jourdan M, Ythier A, *et al.*  
482 APRIL and TACI interact with syndecan-1 on the surface of multiple myeloma  
483 cells to form an essential survival loop. *Eur J Haematol* 2009; **83**: 119–129.
- 484 13 Wijdenes J, Vooijs WC, Clement C, Post J, Morard F, Vlta N, *et al.* A  
485 plasmocyte selective monoclonal antibody (B-B4) recognizes syndecan-1. *Br J*  
486 *Haematol* 1996; **94**: 318–323.
- 487 14 Belnoue E, Tougne C, Rochat AF, Lambert PH, Pinschewer DD, Siegrist CA.  
488 Homing and Adhesion Patterns Determine the Cellular Composition of the  
489 Bone Marrow Plasma Cell Niche. *J Immunol* 2012; **188**: 1283–1291.
- 490 15 Kopf M, Baumann H, Freer G, Freudenberg M, Lamers M, Kishimoto T, *et al.*  
491 Impaired immune and acute-phase responses in interleukin-6-deficient mice.

- 492 *Nature* 1994; **368**: 339–342.
- 493 16 Minges Wols HA, Underhill GH, Kansas GS, Witte PL. The role of bone  
494 marrow-derived stromal cells in the maintenance of plasma cell longevity. *J*  
495 *Immunol* 2002; **169**: 4213–4221.
- 496 17 Cassese G, Arce S, Hauser AE, Lehnert K, Moewes B, Mostarac M, *et al.*  
497 Plasma Cell Survival Is Mediated by Synergistic Effects of Cytokines and  
498 Adhesion-Dependent Signals. *J Immunol* 2003; **171**: 1684–1690.
- 499 18 Mesin L, Di Niro R, Thompson KM, Lundin KEA, Sollid LM. Long-lived plasma  
500 cells from human small intestine biopsies secrete immunoglobulins for many  
501 weeks in vitro. *J Immunol* 2011; **187**: 2867–2874.
- 502 19 Mahtouk K, Moreaux J, Hose D, Rème T, Meissner T, Jourdan M, *et al.* Growth  
503 factors in multiple myeloma: a comprehensive analysis of their expression in  
504 tumor cells and bone marrow environment using Affymetrix microarrays. *BMC*  
505 *Cancer* 2010; **10**: 198.
- 506 20 Anderson KC, Carrasco RD. Pathogenesis of myeloma. *Annu Rev Pathol*  
507 2011; **6**: 249–274.
- 508 21 Mei HE, Yoshida T, Sime W, Hiepe F, Thiele K, Manz RA, *et al.* Blood-borne  
509 human plasma cells in steady state are derived from mucosal immune  
510 responses. *Blood* 2009; **113**: 2461–2469.
- 511 22 Caraux A, Klein B, Paiva B, Bret C, Schmitz A, Fuhler GM, *et al.* Circulating  
512 human B and plasma cells. Age-associated changes in counts and detailed  
513 characterization of circulating normal CD138- and CD138+ plasma cells.

- 514 *Haematologica* 2010; **95**: 1016–1020.
- 515 23 Geffroy-Luseau A, Jégo G, Bataille R, Campion L, Pellat-Deceunynck C.  
516 Osteoclasts support the survival of human plasma cells in vitro. *Int Immunol*  
517 2008; **20**: 775–782.
- 518 24 Cocco M, Stephenson S, Care MA, Newton D, Barnes NA, Davison A, *et al.* In  
519 Vitro Generation of Long-lived Human Plasma Cells. *J Immunol* 2012; **189**:  
520 5773–5785.
- 521 25 Amé-Thomas P, Maby-El Hajjami H, Monvoisin C, Jean R, Monnier D, Caulet-  
522 Maugendre S, *et al.* Human mesenchymal stem cells isolated from bone  
523 marrow and lymphoid organs support tumor B-cell growth: role of stromal cells  
524 in follicular lymphoma pathogenesis. *Blood* 2007; **109**: 693–702.
- 525 26 Jourdan M, Caraux A, Caron G, Robert N, Fiol G, Reme T, *et al.*  
526 Characterization of a transitional preplasmablast population in the process of  
527 human B cell to plasma cell differentiation. *J Immunol* 2011; **187**: 3931–3941.
- 528 27 Reme T, Hose D, De Vos J, Vassal A, Poulain PO, Pantesco V, *et al.* A new  
529 method for class prediction based on signed-rank algorithms applied to  
530 Affymetrix microarray experiments. *BMC Bioinformatics* 2008; **9**: 16.
- 531 28 Le Carrour T, Assou S, Tondeur S, Lhermitte L, Lamb N, Reme T, *et al.*  
532 Amazonia!: An Online Resource to Google and Visualize Public Human whole  
533 Genome Expression Data. *Open Bioinformatics J* 2010; **4**: 5–10.
- 534 29 Tusher VG, Tibshirani R, Chu G. Significance analysis of microarrays applied  
535 to the ionizing radiation response. *Proc Natl Acad Sci USA* 2001; **98**: 5116–

- 536 5121.
- 537 30 Eisen MB, Spellman PT, Brown PO, Botstein D. Cluster analysis and display of  
538 genome-wide expression patterns. *Proc Natl Acad Sci USA* 1998; **95**: 14863–  
539 14868.
- 540 31 Sprynski AC, Hose D, Caillot L, Reme T, Shaughnessy JDJ, Barlogie B, *et al.*  
541 The role of IGF-1 as a major growth factor for myeloma cell lines and the  
542 prognostic relevance of the expression of its receptor. *Blood* 2009; **113**: 4614–  
543 4626.
- 544 32 Bossen C, Schneider P. BAFF, APRIL and their receptors: structure, function  
545 and signaling. *Semin Immunol* 2006; **18**: 263–275.
- 546 33 Annunziata CM, Davis RE, Demchenko Y, Bellamy W, Gabrea A, Zhan F, *et al.*  
547 *al.* Frequent engagement of the classical and alternative NF-kappaB pathways  
548 by diverse genetic abnormalities in multiple myeloma. *Cancer Cell* 2007; **12**:  
549 115–130.
- 550 34 Mahévas M, Patin P, Huetz F, Descatoire M, Cagnard N, Bole-Feysot C, *et al.* B  
551 cell depletion in immune thrombocytopenia reveals splenic long-lived plasma  
552 cells. *J Clin Invest* 2012; **123**: 432–442.
- 553 35 Slifka MK, Antia R, Whitmire JK, Ahmed R. Humoral immunity due to long-lived  
554 plasma cells. *Immunity* 1998; **8**: 363–372.
- 555 36 Manz RA, Hauser AE, Hiepe F, Radbruch A. MAINTENANCE OF SERUM  
556 ANTIBODY LEVELS. *Annu Rev Immunol* 2005; **23**: 367–386.
- 557 37 Medina F, Segundo C, Campos-Caro A, Gonzalez-Garcia I, Brieva JA. The

- 558 heterogeneity shown by human plasma cells from tonsil, blood, and bone  
559 marrow reveals graded stages of increasing maturity, but local profiles of  
560 adhesion molecule expression. *Blood*. 2002; **99**: 2154–2161.
- 561 38 Kallies A. Plasma Cell Ontogeny Defined by Quantitative Changes in Blimp-1  
562 Expression. *J Exp Med* 2004; **200**: 967–977.
- 563 39 Iwakoshi NN, Lee AH, Vallabhajosyula P, Otipoby KL, Rajewsky K, Glimcher  
564 LH. Plasma cell differentiation and the unfolded protein response intersect at  
565 the transcription factor XBP-1. *Nat Immunol* 2003; **4**: 321–329.
- 566 40 Rodríguez-Bayona B, Ramos-Amaya A, López-Blanco R, Campos-Caro A,  
567 Brieva JA. STAT-3 Activation by Differential Cytokines Is Critical for Human -  
568 Generated Plasma Cell Survival and Ig Secretion. *J Immunol* 2013; **191**: 4996–  
569 5004.
- 570 41 Winter O, Moser K, Mohr E, Zotos D, Kaminski H, Szyska M, *et al.*  
571 Megakaryocytes constitute a functional component of a plasma cell niche in the  
572 bone marrow. *Blood* 2010; **116**: 1867–1875.
- 573 42 Tsai C-M, Chiu Y-K, Hsu T-L, Lin I-Y, Hsieh S-L, Lin K-I. Galectin-1 promotes  
574 immunoglobulin production during plasma cell differentiation. *J Immunol* 2008;  
575 **181**: 4570–4579.
- 576 43 Anginot A, Espeli M, Chasson L, Mancini SJC, Schiff C. Galectin 1 Modulates  
577 Plasma Cell Homeostasis and Regulates the Humoral Immune Response. *J*  
578 *Immunol* 2013 ; e-pub ahead of print 24 April 2013;  
579 doi:10.4049/jimmunol.1201885

- 580 44 Odendahl M, Mei H, Hoyer BF, Jacobi AM, Hansen A, Muehlinghaus G, *et al.*  
581 Generation of migratory antigen-specific plasma blasts and mobilization of  
582 resident plasma cells in a secondary immune response. *Blood*. 2005; **105**:  
583 1614–1621.
- 584 45 Bataille R, Jourdan M, Zhang XG, Klein B. Serum levels of interleukin 6, a  
585 potent myeloma cell growth factor, as a reflect of disease severity in plasma  
586 cell dyscrasias. *J Clin Invest* 1989; **84**: 2008–2011.
- 587 46 Moreaux J, Legouffe E, Jourdan E, Quittet P, Reme T, Lugagne C, *et al.* BAFF  
588 and APRIL protect myeloma cells from apoptosis induced by interleukin 6  
589 deprivation and dexamethasone. *Blood* 2004; **103**: 3148–3157.
- 590 47 Chaidos A, Barnes CP, Cowan G, May PC, Melo V, Hatjiharissi E, *et al.*  
591 Clinical drug resistance linked to interconvertible phenotypic and functional  
592 states of tumor-propagating cells in multiple myeloma. *Blood* 2013; **121**: 318–  
593 328.
- 594 48 Walker BA, Leone PE, Chiecchio L, Dickens NJ, Jenner MW, Boyd KD, *et al.*  
595 A compendium of myeloma-associated chromosomal copy number  
596 abnormalities and their prognostic value. *Blood* 2010; **116**: e56–e65.
- 597 49 Morgan GJ, Walker BA, Davies FE. The genetic architecture of multiple  
598 myeloma. *Nat Rev Cancer* 2012; **12**: 335–348.
- 599 50 Kassambara A, Hose D, Moreaux J, Walker BA, Protopopov A, Reme T, *et al.*  
600 Genes with a spike expression are clustered in chromosome (sub)bands and  
601 spike (sub)bands have a powerful prognostic value in patients with multiple  
602 myeloma. *Haematologica* 2012; **97**: 622–630.

- 603 51 Kassambara A, Hose D, Moreaux J, Rème T, Torrent J, Rossi JF, *et al.*  
604 Identification of pluripotent and adult stem cell genes unrelated to cell cycle  
605 and associated with poor prognosis in multiple myeloma. *PLoS ONE* 2012; **7**:  
606 e42161.
- 607 52 Kassambara A, Schoenhals M, Moreaux J, Veyrune J-L, Rème T, Goldschmidt  
608 H, *et al.* Inhibition of DEPDC1A, a Bad Prognostic Marker in Multiple Myeloma,  
609 Delays Growth and Induces Mature Plasma Cell Markers in Malignant Plasma  
610 Cells. *PLoS ONE* 2013; **8**: e62752.
- 611 53 Schoenhals M, Kassambara A, Veyrune JL, Moreaux J, Goldschmidt H, Hose  
612 D, *et al.* Kruppel-like factor 4 blocks tumor cell proliferation and promotes drug  
613 resistance in multiple myeloma. *Haematologica* 2013; e-pub ahead of print 12  
614 April 2013; doi:10.3324/haematol.2012.066944.
- 615 54 Moreaux J, Klein B, Bataille R, Descamps G, Maiga S, Hose D, *et al.* A high-  
616 risk signature for patients with multiple myeloma established from the  
617 molecular classification of human myeloma cell lines. *Haematologica* 2011; **96**:  
618 574–582.
- 619 55 Hose D, Moreaux J, Meissner T, Seckinger A, Goldschmidt H, Benner A, *et al.*  
620 Induction of angiogenesis by normal and malignant plasma cells. *Blood* 2009;  
621 **114**: 128–143.
- 622 56 Schoenhals M, Frecha C, Bruyer A, Caraux A, Veyrune JL, Jourdan M, *et al.*  
623 Efficient transduction of healthy and malignant plasma cells by lentiviral vectors  
624 pseudotyped with measles virus glycoproteins. *Leukemia* 2012; **26**: 1663–  
625 1670.

626 **FIGURE LEGENDS**

627 **Figure 1. Generation and survival of mature CD138<sup>bright</sup> PCs *in vitro*.**

628 (A) PCs (CD38<sup>+</sup>CD138<sup>+</sup> cells) were generated with or without stromal cells (SCs) in  
629 the presence of IL-6 (10 ng/ml) and cell count and viability assayed at day (D) 10, 14,  
630 24, 30, and 60 using trypan blue dye exclusion. Data are the mean concentration of  
631 viable cells  $\pm$  SD determined in six separate experiments. \*The mean value is  
632 significantly higher than that in culture without SCs at the same culture day using a  
633 paired *t*-test ( $P \leq .05$ ). (B) Expression of CD138. The histograms show FACS  
634 labelling with an anti-CD138 (black) or an isotype-matched control mAb (white) of  
635 one experiment representative of four. The percentages of CD138 positive cells and  
636 CD138 staining index (SI) are indicated in the panels. (C) CD138 fluorescence  
637 staining index. Data are the mean CD138 staining index  $\pm$  SD of four separate  
638 experiments. Statistical analysis was done using a paired *t*-test. ns = not significant.

639 **Figure 2. IL-6 in combination with APRIL or BAFF or SC-CM supports the**  
640 **generation and survival of PCs *in vitro*.**

641 (A) D10 early PCs were FACS sorted and cultured from D10 to D30 either in contact  
642 with SCs (culture with SCs), or with SCs placed in a lower chamber of a transwell  
643 culture plate (SCs+PCs in transwells), or with supernatant of PC and SC coculture  
644 (PC/SC-CM), or culture supernatant of SCs (SC-CM). Results are the mean of viable  
645 cell counts  $\pm$  SD of 3-paired experiments. ns: the mean count of PCs at a given  
646 culture day is not significantly different from that in the SC group at the same culture  
647 day using a paired *t*-test (B) FACS-sorted early PCs were cultured from D10 to D30  
648 with culture medium and 10% FCS (Co) or in the presence of recombinant cytokines  
649 and/or the SC-CM. The concentrations of cytokines used were 10 ng/ml for IL-6 and  
650 IGF-1 and 200 ng/ml for APRIL, BAFF, and SDF-1. Counts of metabolic active cells

651 were assayed quantifying intracellular ATP amount with a Cell Titer Glo Luminescent  
652 Assay. Results are the mean  $\pm$  SD of the luminescent signals expressed as the  
653 percentage of that in the group cultured with the SC-CM, determined in 3- to 7-paired  
654 experiments. \*The mean value is significantly different from that in the SC-CM group  
655 using a paired *t*-test. \*\*The mean value is significantly different from that in the IL-6  
656 group using a paired *t*-test ( $P \leq .05$ ). (C) Effect of various inhibitors of cytokines on  
657 the generation of plasma cells induced by the SC-CM. The anti-IL-6 mAb was used  
658 at a concentration of 10  $\mu$ g/ml, the TACI-Fc at 10  $\mu$ g/ml, the AMD3100 (SDF-1  
659 inhibitor) at 10  $\mu$ M and the IGF-1R inhibitor (NVP-AEW541) at 1  $\mu$ M. Metabolic active  
660 cells were assayed quantifying intracellular ATP amount with a Cell Titer Glo  
661 Luminescent Assay. Results are the mean  $\pm$  SD of the luminescent signals  
662 expressed as the percentage of that in the group cultured with the SC-CM,  
663 determined in 3- to 10-paired experiments. \*The mean value is significantly different  
664 from that in the SC-CM group using a paired *t*-test.

665 **Figure 3. A NF- $\kappa$ B inhibitor does not affect the generation of PCs by SCs but**  
666 **inhibited the additive effect of APRIL.**

667 PCs were generated with SCs only or with SCs and 200 ng/ml recombinant APRIL.  
668 The IKK16 NF- $\kappa$ B inhibitor (1 or 3  $\mu$ M) was added for 4 days in 2 culture groups and  
669 PCs counted at the end of the culture. For each experiment the PC count was  
670 expressed as the percentage of the PC count obtained with the control group of the  
671 same experiment (SCs or SCs + APRIL). Results are the mean percentages  $\pm$  SD of  
672 five separate experiments. \*The mean percentage is significantly decreased  
673 compared to that in the SCs + APRIL group using a paired *t*-test.

674 **Figure 4. D30 PCs have a more mature phenotype than D10 early PCs.**

675 PC phenotype was assessed by flow cytometry. Results are the mean percentage  $\pm$   
676 SD of positive cells and the mean staining index  $\pm$  SD determined in 4 to 6 separate  
677 experiments. \*The mean value is significantly different from that in D10 PCs using a  
678 paired *t*-test ( $P \leq .05$ ).

679 **Figure 5. D30 PCs do not cell cycle.**

680 The cell cycle was assessed using DAPI staining and quantification of cells in the S  
681 phase using bromodeoxyuridine (BrdU) incorporation and labelling with an anti-BrdU  
682 antibody. (A) Dot plots show a representative experiment out of three. The  
683 percentages of cells in the G0/G1, S, and G2/M phases are indicated. (B) Results are  
684 the mean percentage  $\pm$  SD of cells in the S phase of three separate experiments.

685 **Figure 6. D30 and D60 PCs are functional PCs that produce Igs continuously.**

686 FACS-sorted D10 early PCs were cultured with SCs (A-F) or growth factors as  
687 indicated (G). (A-C) Cytoplasmic (cy) Igs (IgG, IgA, and IgM) (A), cy-light chains (B),  
688 and surface (s) Igs (C) were assessed by flow cytometry. Results are the mean  
689 percentage  $\pm$  SD of positive cells from 5 separate experiments. (D) IgG, IgA, and IgM  
690 production was assessed by ELISA. Results are the mean  $\pm$  SD of Ig production in  
691 pg per cell and per day determined in 3 separate experiments. (E) The number of  
692 IgG-, IgA-, and IgM-secreting cells was assessed by ELISPOT. Results are the mean  
693 Ig-secreting cell number  $\pm$  SD from 4 separate experiments. (F) ELISPOTs from a  
694 representative experiment are shown. (G) FACS sorted D10 early PCs were cultured  
695 with SCs or with a combination of cytokines and/or SC-CM as indicated. The number  
696 of Ig-secreting cells was determined by ELISPOT at D30 of culture. Results are the  
697 mean Ig-secreting cell number  $\pm$  SD from 3 separate experiments. \*The mean value  
698 is significantly different from that in D10 PCs using a paired *t*-test.

699 **Figure 7. Gene expression of transcription factors involved in PC**  
700 **differentiation.**

701 Naive B cells (BCs), D10 early PCs, and D30 PCs were FACS sorted. Gene  
702 expression of *IRF4*, *PRDM1*, *XBP1u*, *XBP1s*, *PAX5*, and *BCL6* assayed by real-time  
703 RT-PCR. The mRNA level in the different cell populations was compared assigning  
704 the arbitrary value 1 to gene expression in BCs. Data are the mean value  $\pm$  SD of  
705 gene expression determined in 5 separate experiments. The ratio *XBP1s/XBP1u* in  
706 D10 early PCs and D30 PCs is shown. \*The mean value is significantly different from  
707 that in D10 PCs using a paired *t*-test ( $P \leq .05$ ).

708 **Figure 8. Unsupervised clustering of gene expression profile of purified D4**  
709 **prePBs, D7 PBs, D10 early PCs, D30 PCs, and BMPCs.**

710 D4 prePBs, D7 PBs, D10 early PCs, D30 PCs and BMPCs were profiled using  
711 Affymetrix U133 plus 2.0 microarray and an unsupervised hierarchical clustering was  
712 run with the 5000 probe sets with the highest variance (log transform, center genes  
713 and arrays, uncentered correlation and average linkage). The 5 populations are  
714 classified into 2 major clusters, a PC cluster comprising D10 early PCs, D30 PCs and  
715 BMPCs ( $r = 0.11$ ) and a plasmablast cluster comprising prePBs and PBs ( $r = 0.20$ ).  
716 The horizontal lines represent the normalized and centered expression of each of the  
717 5000 genes in the samples and are depicted according to the color scale shown at  
718 the bottom (-1.5 to 1.5 on a log base 2 scale).

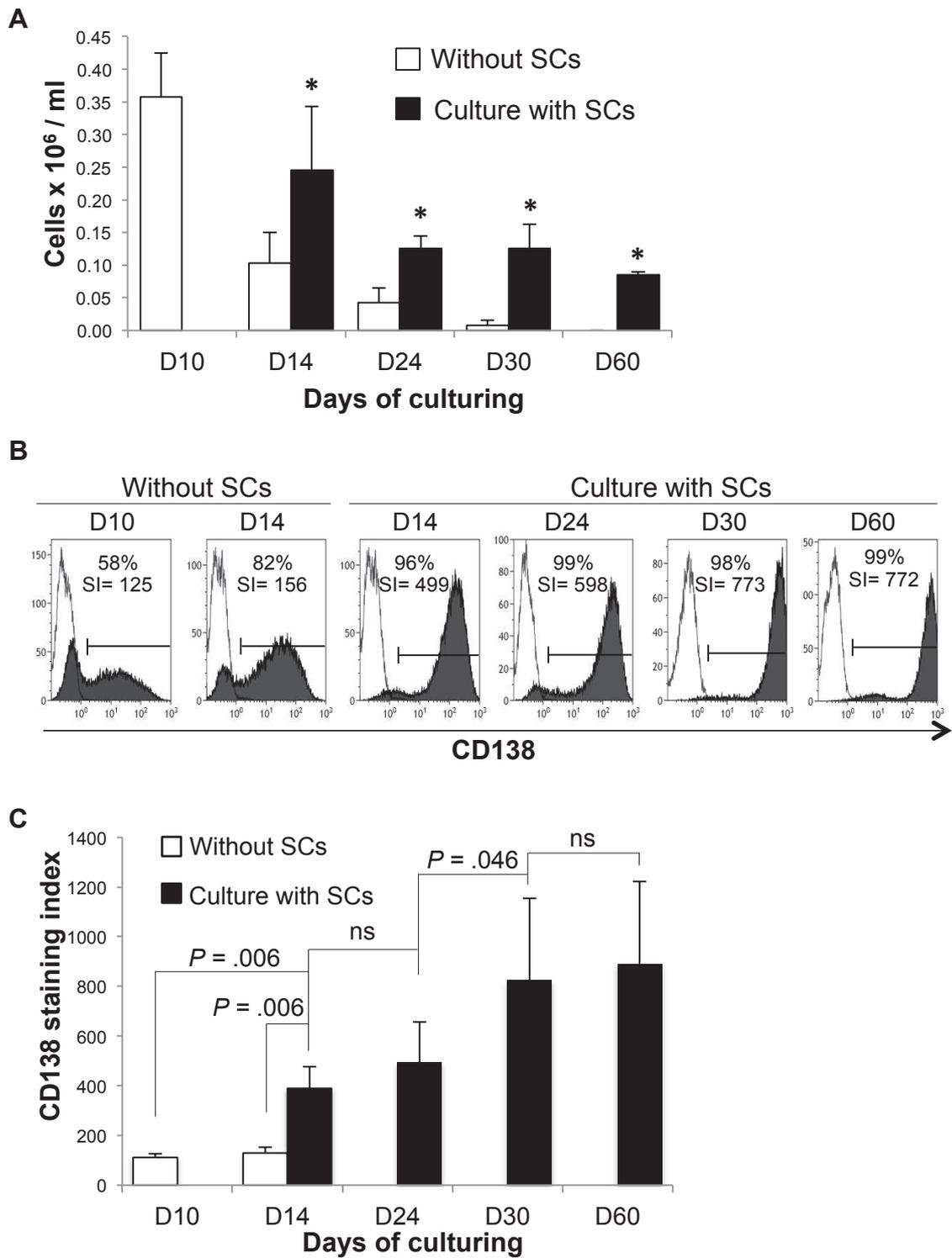


Figure 1

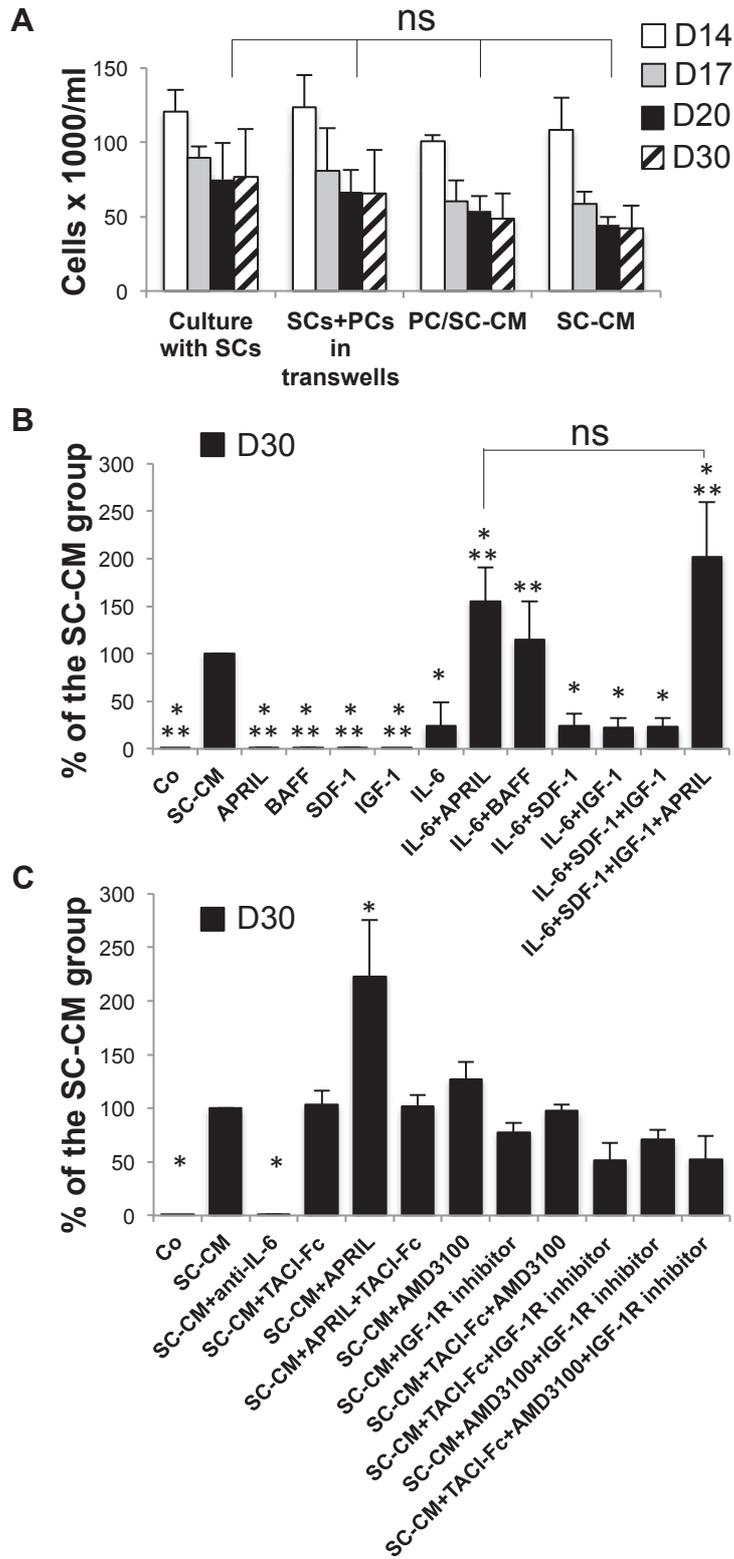


Figure 2

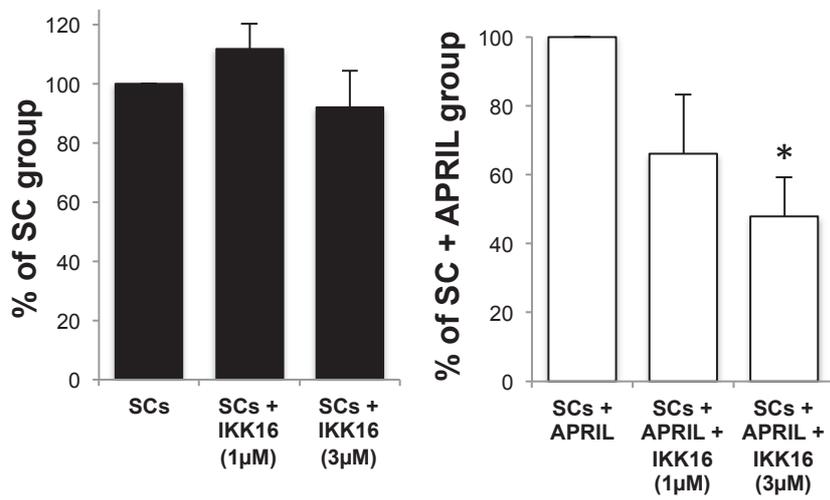


Figure 3

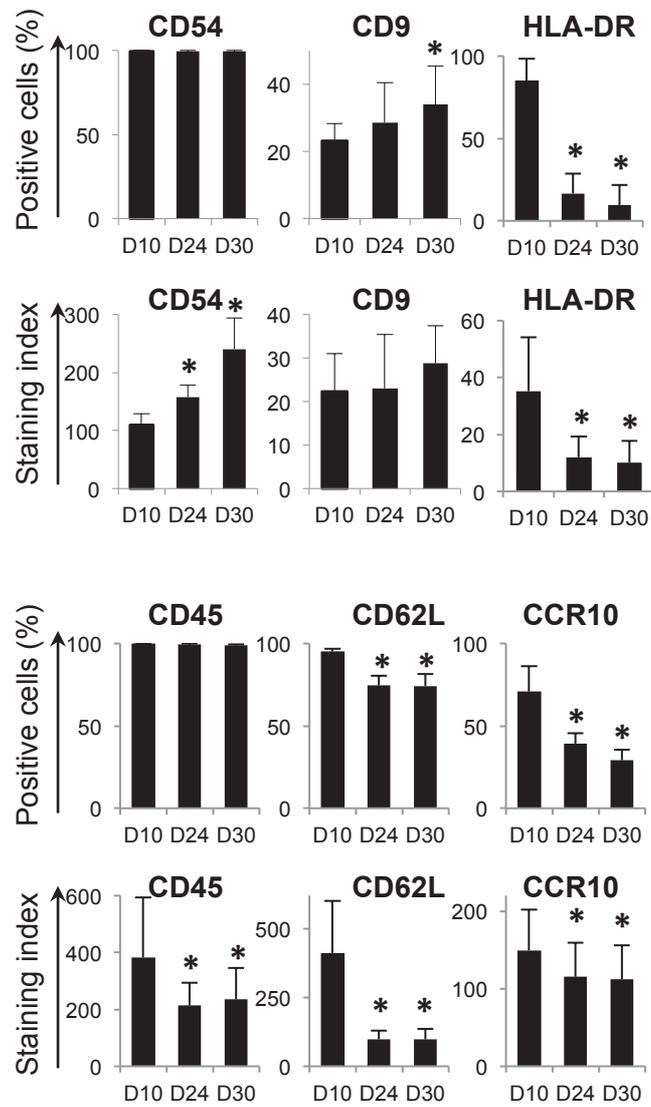


Figure 4

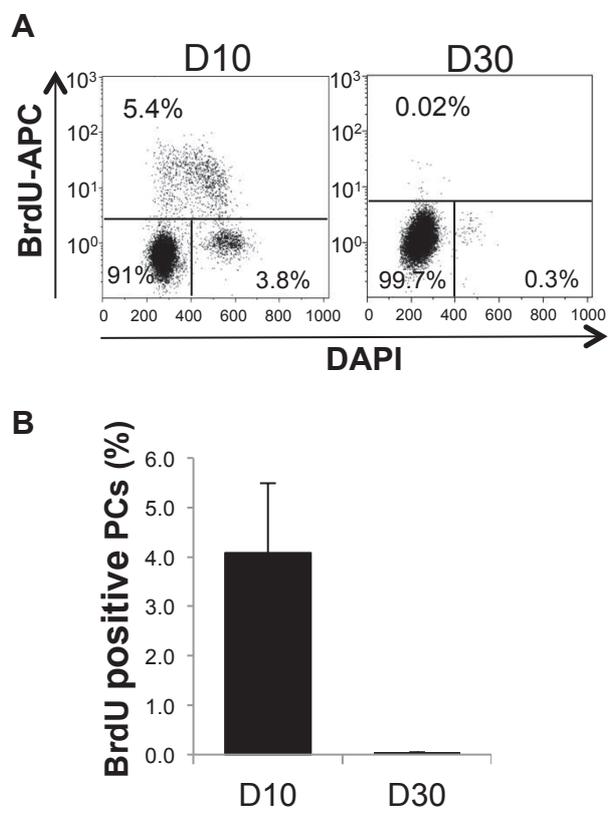


Figure 5

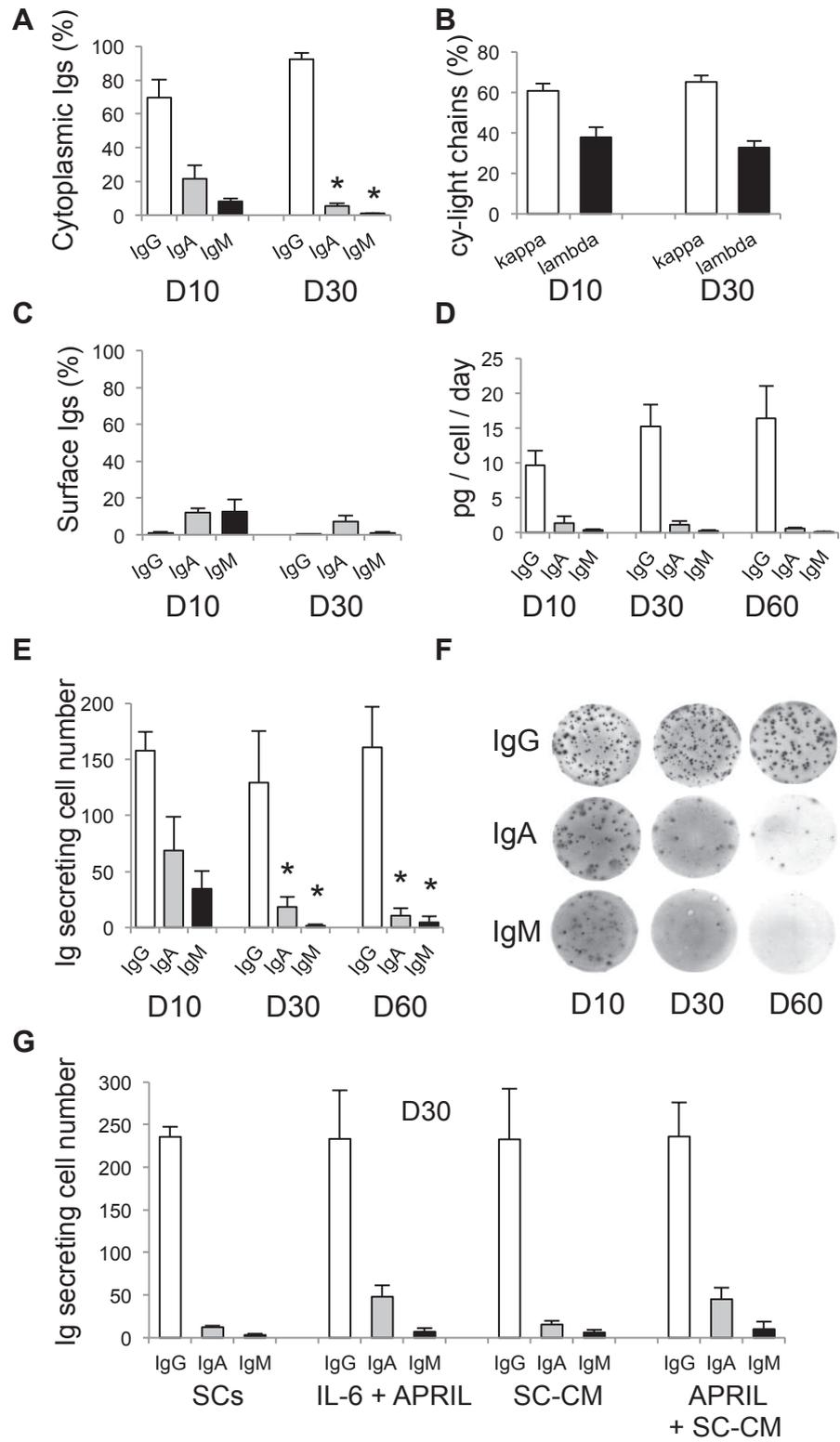


Figure 6

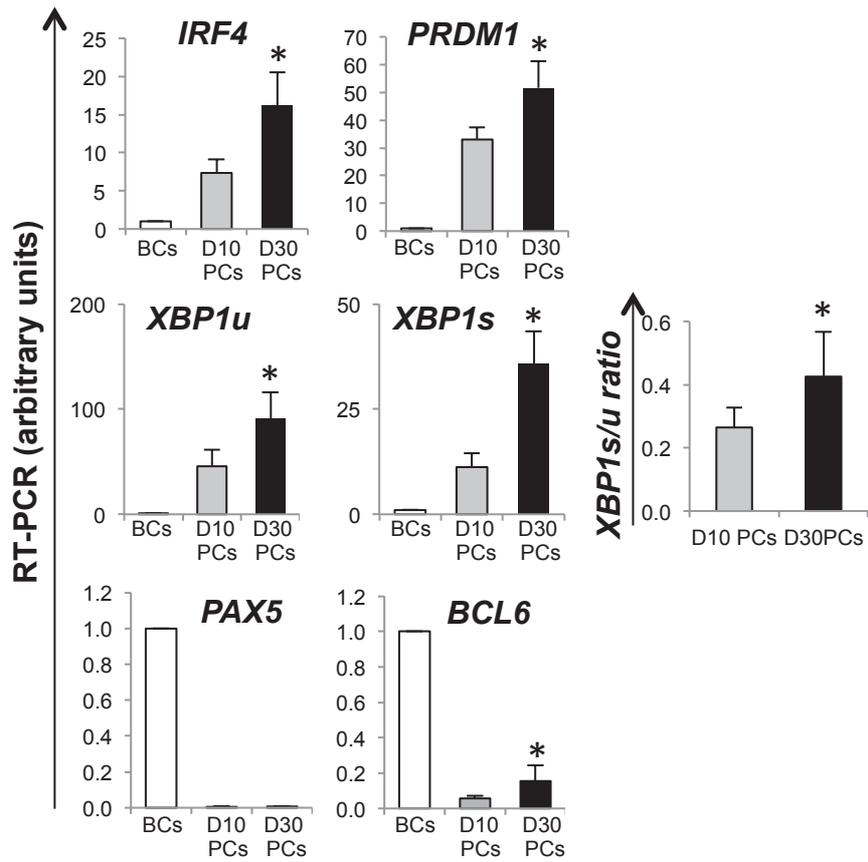


Figure 7

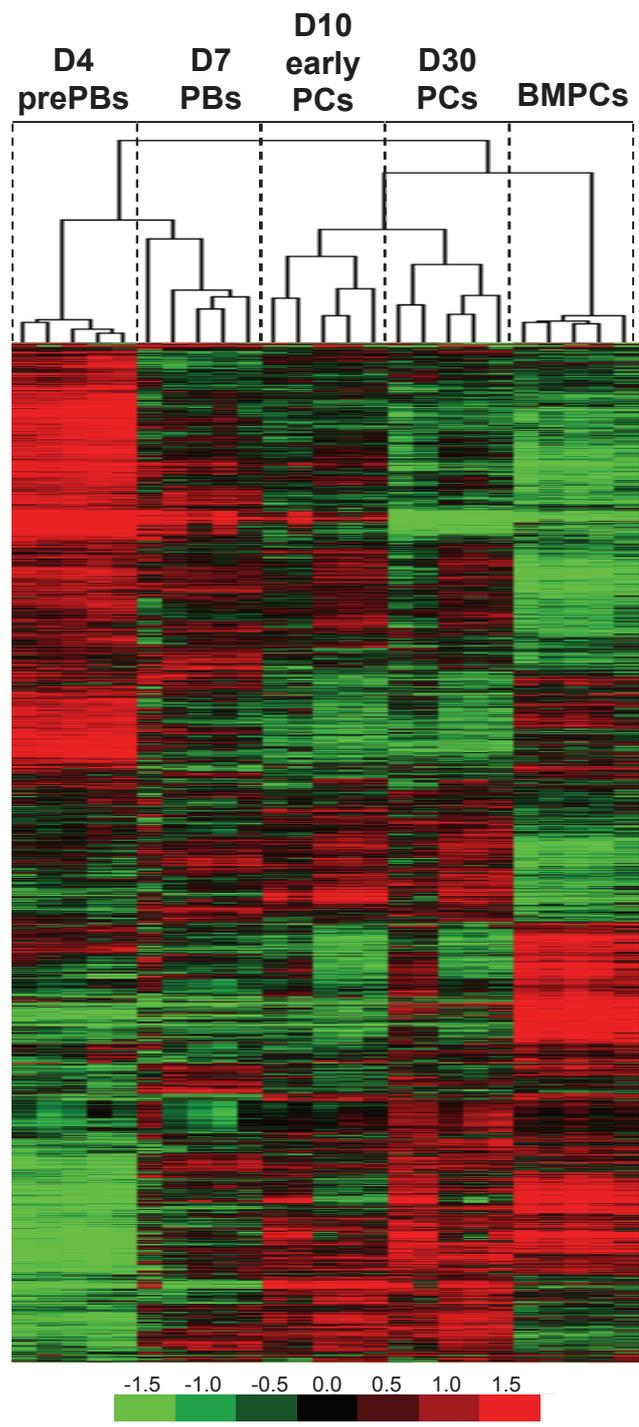


Figure 8