Complementarity of rarity, specialisation and functional diversity metrics to assess community responses to environmental changes, using an example of spider communities in salt marshes
Boris Leroy, Isabelle Le Viol, Julien Pétillon

To cite this version:
Boris Leroy, Isabelle Le Viol, Julien Pétillon. Complementarity of rarity, specialisation and functional diversity metrics to assess community responses to environmental changes, using an example of spider communities in salt marshes. Ecological Indicators, Elsevier, 2014, 46, pp.351-357. 10.1016/j.ecolind.2014.06.037. hal-01101385

HAL Id: hal-01101385
https://hal-univ-rennes1.archives-ouvertes.fr/hal-01101385
Submitted on 8 Jan 2015

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L’archive ouverte pluridisciplinaire HAL, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d’enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.
Complementarity of rarity, specialisation and functional diversity
metrics to assess community responses to environmental changes, using
an example of spider communities in salt marshes

Boris Leroy\textsuperscript{a,b}\textsuperscript{*}, Isabelle Le Viol\textsuperscript{c} & Julien Pétillon\textsuperscript{a}

\textsuperscript{a} EA 7316 Biodiversité et Gestion des Territoires, Université de Rennes I, 263 Av. du Gal. Leclerc, 35042 Rennes Cedex, France
\textsuperscript{b} Ecologie, Systématique & Evolution, UMR CNRS 8079, Univ. Paris-Sud, Orsay Cedex FR-91405, France
\textsuperscript{c} Centre d'Ecologie et des Sciences de la Conservation (CESCO), UMR 7204 MNHN-CNRS-UPMC, 55 rue Buffon, 75005 Paris, France

\textsuperscript{*} Corresponding author: Boris Leroy; Email: leroy.boris@gmail.com; Tel.:+33 666 021 327.

Present address: Ecologie, Systématique & Evolution, UMR CNRS 8079, Univ. Paris-Sud, Orsay Cedex FR-91405, France

Number of words (abstract): 229
Number of words (full word file): 5180
Abstract

The study of community responses to environmental changes can be enhanced by the recent development of new metrics useful in applied conservation: relative rarity, ecological specialisation and functional diversity. These different metrics have been critically assessed independently, but are rarely combined in applied conservation studies, especially for less-studied taxa such as arthropods. Here we report how these different metrics can complement each other by using the response of spider communities to environmental changes in salt marshes as an example. Sampling took place using pitfall traps in salt marshes of the Mont St Michel Bay (France) during 2004 and 2007. The sampling design was spatially replicated (3 plots per treatment and 4 traps per plot) and encompassed four habitat treatments: control, sheep grazing, cutting (annual, in summer) and invasion by the plant Elymus athericus. We observed contrasting responses of spider communities to the different treatments: grazing had a negative impact on both rarity and functional diversity but a positive impact on specialisation; cutting had a negative impact on the three metrics; and invasion only had a negative impact on rarity and specialisation. These contrasting responses emphasise the necessity of using different complementary community metrics in such conservation studies. Consequently, rarity-, specialization-, and functional-based indices should be applied simultaneously more frequently, as they potentially provide additional complementary information about communities. Such complementary information is the key to better-informed conservation choices.

Key-words: Index of Relative Rarity; Community Specialisation Index; Functional Divergence; Grazing; Cutting; Spiders.
Abbreviations:

CSI: Community Specialisation Index (Julliard et al., 2006)

FDiv: Functional Divergence Index (Villéger et al., 2008)

$I_{RR}$: Index of Relative Rarity (Leroy et al. 2013)

SSI: Species Specialisation Index (Julliard et al., 2006)
1. Introduction

Addressing responses of biodiversity to environmental changes at the community rather than species level present the advantage of integrating the responses of multiple species and their interactions. Responses of communities can be assessed with different facets (taxonomic, phylogenetic or functional) for which several new indices have been developed (e.g., Devictor et al., 2010; Meynard et al., 2011; Strecker et al., 2011). However, these different facets are still rarely used in applied conservation studies, particularly for less-studied taxa such as arthropods.

The early methods used to assess communities in conservation studies were simple taxonomic diversity metrics such as species richness or abundance (e.g., Prieto-Benítez and Méndez, 2011). However these simple metrics only reflect a fraction of the biodiversity and do not take into account the identity of species and their characteristics within and between communities, even though these aspects are crucial to assess biodiversity distribution (α, β components), conservation concerns (rarity), ecosystem functioning (functional traits) and importantly the processes implied in the impact of environmental changes on this biodiversity (biotic and functional homogenisation). Hence, species and communities were attributed values with respect to the conservation goal, for example according to their rarity, or more recently to their functional characteristics. In this study, we focus on recent methodological advances regarding three aspects: rarity, ecological specialisation and functional diversity.

Rarity primarily provides an insight into the facet of species biodiversity that is most at risk of extinction (Gaston, 1994), also with respect to the maintenance of vulnerable ecosystem functions (Mouillot et al., 2013). Different axes of rarity are usually considered: restricted abundance, restricted geographic distribution and
narrow niche breadth. The study of rarity in arthropod communities has recently been improved by the proposal of new indices based on species occurrence (Leroy et al., 2012), which provide the possibility of integrating multiple spatial scales (Leroy et al., 2013). Robust metrics have also been developed to assess the average niche width, i.e. the specialisation of species communities (Devictor et al., 2010a). Using basic predictions from the ecological niche theory, specialist species should indeed be favoured in rather stable environments whereas generalists should be more able to thrive in disturbed habitats (Levins, 1968). In the same way, species functional traits are increasingly taken into account to provide a better assessment of the functional responses of communities to environmental changes. The study of this facet of biodiversity was improved thanks to the proposal of novel metrics which have been developed and analysed critically (Devictor et al., 2010a; Villéger et al., 2008). The loss of functional diversity was a criterion that had been overlooked initially, but is of increasing concern in biological conservation (e.g., Devictor et al., 2010b).

All these distinct approaches were successfully applied on arthropod communities (e.g., Leroy et al., 2013; Penone et al., 2013) to assess their responses to environmental changes. They have each been critically assessed alone, but how they complement each other in the case of applied conservation remained to be tested. In this paper, we report a case study combining different recently developed metrics (rarity, specialisation, functional diversity) to assess how they complement each other to assess the response of arthropod communities to environmental changes, by using the example of spider communities of salt marshes. The environmental changes are here the replacement of natural vegetation of salt marshes by monospecific stands of the species *Elymus athericus* (Bockelmann and Neuhaus, 1999), and two management practices likely to limit the spread of this species: annual cutting and
sheep grazing. The impact of *E. athericus* will here be termed as an invasion in accordance with previous work on this species (Pétillon et al., 2005).

Salt marshes are of important conservation value because they host stenotopic species due to the constraining environmental conditions (Pétillon et al., 2011), and geographically rare species because of the restricted distribution of salt marshes in the western Palearctic (Leroy et al., 2013). In addition, salt marshes are subject to environmental changes (invasion by *E. athericus* and management) which often result in the replacement of the single dominant plant species by another (Veeneklaas et al., 2012). These changes in vegetation in turn modify the structure and composition of arthropod communities (Ford et al., 2012). The impacts of these environmental changes on salt marsh arthropods are still poorly understood, and results from scarce literature are often contradictory (e.g., Rickert et al., 2012; van Klink et al., 2013). Consequently, we expected the application of distinct community metrics to provide new and complementary information, thus leading to a better understanding of how environmental changes impact communities. For that purpose, we compared the impacts of four treatments (control, invasion, cutting and grazing) on spider communities of salt marshes using community-level indices. Spiders were selected as a model group as they constitute one of the most abundant and diverse groups of arthropods in salt marshes (Pétillon et al., 2008) and for their well-known sensitivity to changes in habitat structure (e.g., Marc et al., 1999).
2. Methods

2.1. Sampling design

The impacts of treatments on spider communities were investigated at two sites in the Mont-Saint-Michel Bay (NW France, 48°37’N, 1°34’W), 1 kilometre apart. Four treatments were investigated: control, invasion by *E. athericus*, vegetation cutting (once a year, in July) and grazing by sheep (on average 50 sheep/ha) (Pétillon et al., 2007). Treatments are representative of the main salt-marsh habitats of the Mont St-Michel Bay (Pétillon et al., 2007), and covered all together 89% of the 4054 hectares of salt marshes (*E. athericus*-invaded areas: 35%, cutting: 19%, sheep-grazing: 25%, natural vegetation: 10%, data from 2007, Valéry and Radureau, personal communication).

Spider communities were sampled between May and June in 2004 and 2007: the former with control, invasion and cutting treatments, and the latter with control, invasion and grazing treatments. The comparison between treatments was made in similar salt-marsh zones within each site and the only apparent varying factors (at the local and landscape scales) between plots were the presence/absence of management practices (cutting and grazing) or invasion by *E. athericus*.

The sampling protocol was designed to be comparable among treatments: within each site, each treatment was applied to three plots during the same sampling period. Plots had a surface area of 100m² and were spaced 100m apart. Within each plot, ground-dwelling spiders were sampled with four pitfall traps, set up regularly in a square grid and placed 10m part, as this is the minimum distance to avoid interference between traps (Topping and Sunderland, 1992). Traps consisted of polypropylene cups (10 cm diameter, 17 cm deep) containing ethylene-glycol as a preservative. Traps were covered with a raised wooden roof to exclude the rain and
were visited weekly, tides permitting (i.e. three times per month during May and June). To summarise, there were 36 traps per site (3 treatments * 3 plots * 4 traps) and thus a total of 72 traps for the whole sampling protocol. To verify the impacts of treatments on vegetation, percentage covers of all plant species were estimated once within a radius of 1m around all traps.

2.2. Spider community-level indices

We calculated the average rarity, specialisation, and functional diversity of each community (pitfall trap) using species characteristics obtained from spider datasets (rarity, specialisation), and the literature (hunting strategy).

Data came from i) the western France spider database, and ii) the Catalogue of Spider Species from Europe and the Mediterranean Basin (both datasets were detailed in Leroy et al., 2013).

2.2.1. Multiscale Index of Rarity

For each spider species, we calculated rarity weights \( w_{Mi} \) according to the method described in Leroy et al. (2013) (details in Appendix A). These rarity weights integrate information on the occurrence of species at two spatial scales: the western France scale and the western Palearctic scale. Weight values range from 0 to 2. The rarer the species, the higher the weights, with species which are rare at both scales receiving higher rarity weights than species which are rare at a single scale.

The Index of Relative Rarity \( I_{RR} \) of each pitfall community was then calculated as the average weight of rarity of individuals of all the species of the considered community, and was subsequently normalized to values between 0 (no rare species in the community) and 1 (all individuals of the community belong to species with the
maximum rarity weight): \( I_{RR} = (\sum (a_i \times w_{Mi})/N) - w_{\min})/(w_{\max} - w_{\min}) \) where \( a_i \) and \( w_{Mi} \) respectively are the abundance and rarity weight of the \( i^{th} \) species of the community; \( N \) is the total number of individuals in the community; and \( w_{\min} \) and \( w_{\max} \) are the minimum and maximum possible weights, respectively.

### 2.2.2. Community Specialisation Index

Each species was characterised for habitat specialisation through using the calculation of a Species Specialisation Index (SSI) according to Julliard et al. (2006). To define habitat specialisation, the western France spider database was used. For each species, the frequencies of occurrence in each habitat class were calculated (see details in Appendix B). The coefficient of variation of these frequencies of occurrence (standard deviation/average) as a measure of each Species Specialisation Index (SSI) (Julliard et al., 2006) was then calculated. The SSI varied between 0.76 (most generalist species) and 4.69 (most specialised species) (Table 1 and Appendix B). The Community Specialisation Index (CSI) of each pitfall community was calculated as the average SSI of the species detected, weighted by local species abundance, as follows: \( CSI = \sum (a_i \times SSI_i)/N \), where \( a_i \) and \( SSI_i \) respectively are the abundance and species specialisation indices of species \( i \); and \( N \) is the total number of individuals in the community (Devictor et al., 2008). The CSI was then standardised between 0 and 1 in a similar manner to the Index of Relative Rarity (Leroy et al., 2012); a CSI of 1 means that a community is composed of individuals of the most specialised species of the database, whereas a CSI of 0 means that a community is composed of individuals of the most generalist species.

### 2.2.3. Functional diversity
The functional diversity of spider communities was calculated on the basis of functional traits related to hunting modes, because of the important impacts of predator hunting modes in ecosystem functioning (Schmitz, 2009). We used functional traits at the family level on the basis of the trait matrix of spider families of Cardoso et al. (2011). The Functional Divergence (FDiv) index (Villéger et al., 2008) was calculated in order to take into account both the occupation of functional space by the different families and the relative abundance of families in communities.

2.3. Analyses

Because rarity weights and specialisation indices of species describe two axes of rarity, we first analysed their correlation for the sampled species with Pearson’s correlation coefficient. Species richness and the three indices ($\text{IRR}$, CSI and FDiv) were then calculated for each community of spiders of the 72 traps. The FDiv index was calculated for only 58 out of 72 communities, because functional divergence cannot be estimated when communities contain less than three functionally singular species. The effects of treatments on the calculated metrics were then analysed using linear mixed-effects models because of the hierarchical nature of the sampling design. We fitted linear mixed-effect models with the treatment as a fixed effect, and sites nested in year as random effects. The significance of fixed effects was tested by comparing models with vs. without fixed effects using likelihood ratio tests. We then performed multiple comparisons between treatments with Tukey’s post-hoc tests with a correction on probability values by controlling the false discovery rate (Benjamini and Hochberg, 1995). Welsh two sample t-tests were used to compare the average abundance between treatments, of several species. Indices were calculated with Rarity version 1.2-1 (Leroy, 2013) and FD version 1.0-11 (Laliberté and Shipley, 2011). Linear mixed effect models were performed in R 3.0.2 (R Core Team, 2013).
with the package lme4 version 1.0-5 (Bates et al., 2013), and post-hoc tests with the package multcomp version 1.3-1 (Hothorn et al., 2008).
3. Results

3.1. Samplings

A total of 3826 spiders, representing 31 species, 10 families and 6 functional groups, was collected (Table 1). The dominance of single plant species per treatment was confirmed: *Atriplex portulacoides* in the control plots (mean±sd=66.8±11.5%, range=[55%-88%], n=12), *Elymus athericus* in the invaded plots (mean±sd=89.4±7.3%, range=[70%-93%], n=12), *Festuca rubra* in the cut plots (mean±sd=62.9±5.4%, range=[55%-70%], n=12) and *Puccinellia maritima* in the grazed plots (mean±sd=77.5±21.4%, range=[30%-90%], n=12).

3.2. Correlation between rarity weights and specialisation indices of species

We observed a positive correlation between rarity weights and specialisation indices of species (Pearson’s R = 0.75; P < 0.001; N = 31) (Fig. 1). Nevertheless, some species had very high specialisation values but low rarity weights (e.g., *Erigone longipalpis*), or had very high rarity weights but intermediate specialisation values (e.g., *Pardosa purbeckensis*).

3.3. Effects of treatments on communities

We detected a significant treatment effect on all the measured metrics (see Figure 1 and details in Appendix B). The species richness was significantly lower in traps located in invaded plots than in traps associated with other treatments ($\chi^2=13.5$, df=3, p=0.004) (Fig. 2A). The relative rarity of communities was significantly higher in control plot traps than in those located in managed or invaded plots ($\chi^2=24.1$, df=3, p<0.001) (Fig. 2B). This increase in rarity was associated with a higher mean abundance of the rare species *P. purbeckensis* in the control treatment than in other treatments ($t=5.78$, df=31.50, p<0.001) (Table 1). Conversely, although the CSI was
significantly higher in the control treatment than under invasion or cutting treatments, the highest CSI was observed in the grazed plots, and was significantly higher than in control and in invaded or cut plots ($\chi^2=24.40$, df=3, p<0.001) (Fig. 2C). The higher specialisation under the control compared to invasion and cutting treatments was associated with higher average abundance of specialised species such as *P. purbeckensis* ($t=6.60$, df=30.22, p<0.001) and *Silometopus ambiguus* ($t=2.90$, df=33.08, p=0.007) (Table 1). On the other hand, the very high specialisation value observed under the grazing treatment was associated to the high abundance of the specialised species *E. longipalpis*, with on average $13.83\pm 9.76$ individuals per trap versus $1.13\pm 2.05$ individuals per trap (mean±sd) in other treatments ($t=4.49$, df=11.20, p<0.001). However, grazing, and to a lesser extent cutting, appeared to have a negative impact on the functional divergence of communities, with a significantly lower FDiv in traps of grazed plots than in those of invaded or control traps ($\chi^2=10.4$, df=3, p=0.016) (Fig. 2D). We compared this lower functional diversity to the presence of functional groups under the grazing treatment, and we observed only two dominant functional groups (“Ground hunters” and “Other hunters”), while other treatments were more balanced, with an additional functional group (“Orb web weavers”) (Table 1).
4. Discussion

In this paper, we combined different metrics (rarity, specialisation and functional diversity) to analyse how spider communities of salt marshes respond to changes in their environment, whether induced by a biological invasion (*Elymus athericus*) or by management practices (grazing, cutting). As expected, environmental changes generally had negative impacts on spider communities, which led to decreases in rarity, specialisation and functional diversity of communities. While the general trend was similar among metrics, we observed divergent responses to the relative impacts of treatments between the three measured metrics.

Influence of community composition on metrics

In particular, the most interesting divergence was observed for the effect of grazing treatment. Specialisation was higher in grazed plots than in control plots, whereas the opposite was observed for rarity and functional diversity. This is surprising given the observed positive relationship between rarity weights and specialisation indices at the species level. However, this relationship has exceptions such as the widespread but specialist species *E. longipalpis*. Incidentally, *E. longipalpis* was highly abundant in the grazed plot, leading to a divergence between specialisation and rarity indices. This pattern shows that metrics expected to vary conjointly can produce divergent outcomes because of their sensitivity to the composition of communities. This sensitivity is very important to highlight unexpected impacts of environmental changes on communities.

Another important property of the metrics used here is that they are not biased by the species richness or abundance: because the metrics are based on the average trait (rarity, specialisation) of the species in the community, then a species-poor community *can* have a higher value than a species-rich community (Julliard et al.,
If we take the example of the CSI, a community with only a few individuals from very specialised species will have a higher CSI than a community with many individuals of generalist species. Because of this averaging property, a decrease in CSI may result from a decrease of specialists and/or from an increase of generalist species. Typically disturbances are expected to negatively impact specialist species while fostering generalist species, as both predicted by theory and empirically observed (see Devictor and Robert, 2009). As a consequence it is expected that changes in community composition resulting from environmental disturbances will generally result in lower CSI values. However, in extreme cases, the reverse might be observed: for example a disturbance corresponding to the extreme values of the abiotic gradient may have positive impacts on only a few specialist species with particular adaptation to this disturbance (e.g., Doxa et al., 2010), which would thus result in a higher CSI value. Though this is a particular case, it highlights the importance of verifying, as a second step, how community composition changed. This impact is similar on the functional divergence index: the loss of species occupying intermediate functions will increase the functional value. However, this is not relevant in our case study since spider functional groups are strongly divergent; thus, a decrease in abundance of any functional group will decrease the functional value.

To summarise, the three applied metrics accurately synthesise how changes in community composition affect different facets of biodiversity; such changes cannot be detected by species richness alone. However, understanding these changes imply to look at changes in community composition as a second step.

**Implications for salt marsh conservation**

The indices showed that grazing induced a decrease in the average rarity and functional diversity of communities, but an increase in their specialisation. This
pattern is mainly due to the increase in widespread but specialist species such as *E. longipalpis*. This positive impact of grazing on specialist species of salt marshes such as *E. longipalpis* was reported by Ford et al. (2012), although they did not investigate species rarity. The negative effect of grazing, and to a lesser extent cutting, on functional diversity is clearly due to a reduction in the number of vegetation strata, which negatively impacts spiders living in higher vegetation such as orb web weavers (Uetz et al., 1999). On the other hand, no significant impact of grazing was detected on species richness alone, unlike Rickert et al. (2012) and van Klink et al. (2013). In fact, a negative impact on species richness was only observed for the invaded treatment. We also found higher specialisation and rarity in control plots than in invaded and cut plots, in accordance with a previous study that revealed more continental, non-specialised, spider species in cut or invaded salt marshes (Pétillon et al., 2005).

To summarise, our results suggest that undisturbed salt marshes generally support a higher specialisation, rarity and functional diversity than disturbed salt marshes, with the notable exception of the positive impact of grazing on specialised species. To preserve all the facets of spider diversity, we would recommend maintaining the grazing management in areas invaded by *E. athericus* rather than the cutting treatment.

**Conclusion**

We therefore showed contrasting responses of spider communities to the different environmental changes (invasion and management measures). These contrasts emphasise the necessity of using different and complementary community metrics in such conservation studies. As a consequence, rarity-, specialization- and functional-based indices should be more frequently applied together, as they can potentially
contribute a wide range of complementary information about species communities. Such complementary information is the key to better-informed conservation choices.

Acknowledgements

We thank Simon Potier, Alain Radureau and Vincent Schricke for fruitful discussion, Damien Nouguès and Mathieu de Flores for their contribution in field work and spider identification. Aldyth Nys provided useful editing of the English of an earlier draft. Loïc Valéry provided data on the spatial distribution of treatments. We thank two anonymous reviewers for useful additions in an earlier draft.
References


Table 1. Multiscale rarity weights, specialisation indices and functional groups of the sampled spider species, and their average ± sd abundance in pitfall traps. The functional groups are based on Cardoso et al. (2011).

<table>
<thead>
<tr>
<th>Species</th>
<th>Multiscale rarity weights</th>
<th>Species specialisation indices</th>
<th>Functional groups</th>
<th>Abundance ± sd</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>Control</td>
<td>Invasion</td>
</tr>
<tr>
<td>Family Araneidae</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Argiope bruennichi</em></td>
<td>0.009</td>
<td>1.272</td>
<td>Orb web weavers</td>
<td>0.00±0.00</td>
</tr>
<tr>
<td><em>Larinioides cornutus</em></td>
<td>0.004</td>
<td>1.225</td>
<td>Orb web weavers</td>
<td>0.04±0.20</td>
</tr>
<tr>
<td><em>Neoscona adianta</em></td>
<td>0.002</td>
<td>1.271</td>
<td>Orb web weavers</td>
<td>0.00±0.00</td>
</tr>
<tr>
<td>Family Clubionidae</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Clubiona stagnatilis</em></td>
<td>0.144</td>
<td>1.877</td>
<td>Other hunters</td>
<td>0.00±0.00</td>
</tr>
<tr>
<td>Family Dictinidae</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Argenna patula</em></td>
<td>0.344</td>
<td>3.247</td>
<td>Ground hunters</td>
<td>0.71±1.16</td>
</tr>
<tr>
<td>Family Gnaphosidae</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Drassyllus pusillus</em></td>
<td>0.031</td>
<td>1.429</td>
<td>Ground hunters</td>
<td>0.04±0.20</td>
</tr>
<tr>
<td><em>Silometopus ambiguus</em></td>
<td>0.697</td>
<td>3.164</td>
<td>Other hunters</td>
<td>3.29±3.38</td>
</tr>
<tr>
<td><em>Zelotes latreillei</em></td>
<td>0.026</td>
<td>1.749</td>
<td>Ground hunters</td>
<td>0.00±0.00</td>
</tr>
<tr>
<td>Family Linyphiidae</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Agyneta conigera</em></td>
<td>0.551</td>
<td>4.286</td>
<td>Sheet web weavers</td>
<td>0.00±0.00</td>
</tr>
<tr>
<td><em>Bathyphantes gracilis</em></td>
<td>0.012</td>
<td>1.368</td>
<td>Sheet web weavers</td>
<td>0.21±0.51</td>
</tr>
<tr>
<td><em>Collinsia inerrans</em></td>
<td>0.525</td>
<td>2.222</td>
<td>Other hunters</td>
<td>0.04±0.20</td>
</tr>
<tr>
<td><em>Erigone atra</em></td>
<td>0.010</td>
<td>1.346</td>
<td>Other hunters</td>
<td>0.46±0.78</td>
</tr>
<tr>
<td><em>Erigone dentipalpis</em></td>
<td>0.003</td>
<td>1.195</td>
<td>Other hunters</td>
<td>0.17±0.38</td>
</tr>
<tr>
<td><em>Erigone longipalpis</em></td>
<td>0.234</td>
<td>4.690</td>
<td>Other hunters</td>
<td>2.50±2.69</td>
</tr>
<tr>
<td><em>Oedothorax fuscus</em></td>
<td>0.022</td>
<td>1.224</td>
<td>Other hunters</td>
<td>4.83±6.68</td>
</tr>
<tr>
<td><em>Oedothorax retusus</em></td>
<td>0.028</td>
<td>1.353</td>
<td>Other hunters</td>
<td>0.58±0.78</td>
</tr>
<tr>
<td><em>Pelecopsis parallela</em></td>
<td>0.051</td>
<td>1.830</td>
<td>Other hunters</td>
<td>0.00±0.00</td>
</tr>
<tr>
<td><em>Savignia frontata</em></td>
<td>0.325</td>
<td>3.720</td>
<td>Other hunters</td>
<td>0.00±0.00</td>
</tr>
<tr>
<td>Species</td>
<td>Abundance</td>
<td>Activity Type</td>
<td>Abundance Mean ± SD</td>
<td></td>
</tr>
<tr>
<td>-------------------------------</td>
<td>-----------</td>
<td>---------------------</td>
<td>---------------------</td>
<td></td>
</tr>
<tr>
<td><em>Stemonyphantes lineatus</em></td>
<td>0.031</td>
<td>Other hunters</td>
<td>0.04±0.20 0.13±0.34 0.00±0.00 0.00±0.00</td>
<td></td>
</tr>
<tr>
<td><em>Tenuiphantes tenuis</em></td>
<td>0.003</td>
<td>Sheet web weavers</td>
<td>0.75±1.03 0.75±1.11 0.50±0.67 1.33±0.78</td>
<td></td>
</tr>
<tr>
<td><strong>Family Liocranidae</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Agroeca lusatica</em></td>
<td>0.555</td>
<td>Ground hunters</td>
<td>0.33±0.56 0.42±0.83 0.08±0.29 0.00±0.00</td>
<td></td>
</tr>
<tr>
<td><strong>Family Lycosidae</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Alopecosa pulverulenta</em></td>
<td>0.007</td>
<td>Ground hunters</td>
<td>0.00±0.00 0.08±0.28 0.00±0.00 0.00±0.00</td>
<td></td>
</tr>
<tr>
<td><em>Arctosa fulvolineata</em></td>
<td>0.560</td>
<td>Ground hunters</td>
<td>3.13±2.42 2.17±1.97 4.00±3.49 2.00±1.28</td>
<td></td>
</tr>
<tr>
<td><em>Pardosa palustris</em></td>
<td>0.035</td>
<td>Ground hunters</td>
<td>0.04±0.20 0.00±0.00 0.08±0.29 0.08±0.29</td>
<td></td>
</tr>
<tr>
<td><em>Pardosa prativaga</em></td>
<td>0.023</td>
<td>Ground hunters</td>
<td>0.04±0.20 0.04±0.20 0.00±0.00 0.00±0.00</td>
<td></td>
</tr>
<tr>
<td><em>Pardosa proxima</em></td>
<td>0.019</td>
<td>Ground hunters</td>
<td>0.00±0.00 0.08±0.28 0.17±0.39 0.00±0.00</td>
<td></td>
</tr>
<tr>
<td><strong>Family Tetragnathidae</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Pachygnatha clercki</em></td>
<td>0.010</td>
<td>Orb web weavers</td>
<td>0.00±0.00 0.00±0.00 0.33±0.65 0.00±0.00</td>
<td></td>
</tr>
<tr>
<td><em>Pachygnatha degeeri</em></td>
<td>0.006</td>
<td>Orb web weavers</td>
<td>2.54±2.28 2.83±4.57 1.83±2.29 0.33±0.65</td>
<td></td>
</tr>
<tr>
<td><strong>Family Theridiidae</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Enoplognatha mordax</em></td>
<td>0.214</td>
<td>Space web weavers</td>
<td>0.08±0.28 0.04±0.20 0.08±0.29 0.08±0.29</td>
<td></td>
</tr>
<tr>
<td><strong>Family Thomisidae</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Ozyptila simplex</em></td>
<td>0.091</td>
<td>Ambush hunters</td>
<td>0.04±0.20 0.00±0.00 0.00±0.00 0.08±0.29</td>
<td></td>
</tr>
<tr>
<td><strong>Total abundances</strong></td>
<td></td>
<td></td>
<td>41.00±14.63 18.71±11.96 27.42±11.77 37.67±10.53</td>
<td></td>
</tr>
</tbody>
</table>
Figure 1. Correlation between specialisation indices and rarity weights of the sampled species. Two species highly abundant in some of the sampled sites and with high weight values are indicated: *Erigone longipalpis* and *Pardosa purbeckensis*. 
Figure 2. Average values of indices (A. Species richness, B. Multiscale Index of Relative Rarity, C. Community Specialisation Index, and D. Functional Divergence) and 95% confidence intervals (estimated by linear mixed-effects models (LMMs)) for the four treatments. Different successive letters indicate significant differences in means (revealed by corrected Tukey’s post-hoc test on treatment effect following LMMs, with correction for the false discovery rate).