Pd2 $^+\text{-mediated base pairing in oligonucleotides}$
Oleg Golubev, Guillaume Turc, Tuomas Lönnberg

To cite this version:
Oleg Golubev, Guillaume Turc, Tuomas Lönnberg. Pd2 $^+\text{-mediated base pairing in oligonucleotides}$. Journal of Inorganic Biochemistry, Elsevier, 2016, 155, pp.36–43. <10.1016/j.jinorgbio.2015.11.008>. <hal-01263040>

HAL Id: hal-01263040
https://hal-univ-rennes1.archives-ouvertes.fr/hal-01263040

Submitted on 1 Feb 2016

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L’archive ouverte pluridisciplinaire HAL, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d’enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.
Pd$^{2+}$-Mediated Base Pairing in Oligonucleotides

Oleg Golubev$^a$, Guillaume Turc$^{a,b}$ and Tuomas Lönnberg$^a$*

$^a$ Department of Chemistry, University of Turku, Vatselankatu 2, FIN-20014 Turku, Finland

$^b$ École nationale supérieure de chimie de Rennes, 11 allée de Beaulieu – CS 50837, 35708 Rennes Cedex 7, France

* Corresponding author: E-mail: tuanlo@utu.fi; Fax: +358 2 333 6700
ABSTRACT: Two short glycol nucleic acid (GNA) oligonucleotides, having either a terminal or an intrachain nucleobase replaced by the pyridine-2,6-dicarboxamide chelate of Pd$^{2+}$, have been synthesized and their hybridization properties studied by melting temperature measurements. In the termini of a double-stranded oligonucleotide, the Pd$^{2+}$ chelates provided dramatic stabilization of the duplex relative to its metal-free counterpart, in all likelihood owing to formation of Pd$^{2+}$-mediated base pairs between pyridine-2,6-dicarboxamide and the opposing nucleobase. In contrast, no stabilization was observed when the Pd$^{2+}$ chelate was placed in the middle of the chain. Furthermore, the results could not be reproduce by adding a Pd$^{2+}$ salt in situ to the dilute oligonucleotide solutions but the palladated oligonucleotides had to be synthesized and purified prior to the hybridization studies. This behavior, presumably attributable to the relatively slow ligand-exchange reactions of Pd$^{2+}$, differs greatly from what is usually observed with more labile metal ions. The present results offer an explanation for the failure of previous attempts to incorporate Pd$^{2+}$-mediated base pairs into oligonucleotides.

1. Introduction

Metal-ion-mediated base pairing of nucleic acids has been studied extensively for more than a decade owing to its potential in DNA nanotechnology and in expanding the genetic alphabet.[1-9] So far, the most promising results have been obtained with the kinetically labile Ag$^+$, Hg$^{2+}$ or Cu$^{2+}$ as the bridging metal ion. For expanding the range of applications of metal-ion-mediated base pairing to chemotherapy, however, the low intracellular concentrations of these metal ions present a severe limitation. DNA cross-linking by the kinetically inert Pt$^{2+}$, in turn, is a widely employed strategy in cancer chemotherapy[10-12] but suffers from low sequence-specificity. Oligonucleotides capable of metal-ion-mediated hybridization even under the metal-deficient conditions of the cell would offer an elegant solution to this problem.
Along with Pt²⁺, Pd²⁺ appears an ideal candidate for metal-ion-mediated base pairing. Both of these metal ions prefer nitrogen over oxygen ligands and form highly stable square-planar complexes that are compatible with the steric requirements of the base stack of a double-helical nucleic acid. At monomeric level, several Pd²⁺-mediated base pairs have been described in the literature.[13-19] In fact, the very first artificial metal-ion-mediated base pair was formed through coordination of Pd²⁺.[20] Incorporation of a Pd²⁺-mediated base pair into an oligonucleotide, however, remains to be convincingly demonstrated. The apparent discrepancy between the results obtained at monomer and oligomer levels may be understood in terms of the relatively slow ligand exchange reactions of Pd²⁺. A double-helical oligonucleotide presents many potential donor atoms and a Pd²⁺ ion introduced in situ may well remain kinetically trapped at a nonproductive binding site.

In this article we present compelling evidence for Pd²⁺-mediated base pairing within an oligonucleotide. K₂PdCl₄ was allowed to react with short glycol nucleic acid (GNA) oligonucleotides incorporating a single high-affinity ligand. The GNA backbone was chosen as a test bed because of the relatively easy and facile synthesis of modified GNA building blocks. The metallated oligonucleotides were purified by RP HPLC before hybridization experiments. Coordination of a single Pd²⁺ ion by the modified oligonucleotides was verified by mass spectrometric analysis. Thermal melting studies with these oligonucleotides revealed the Pd²⁺-mediated base pair to be better tolerated in a terminal than in an internal position.

2. Results and discussion

2.1. Building block synthesis

The GNA phosphoramidite building blocks of the four canonical nucleobases were prepared as described previously.[21-23] For preparation of a respective building block of the metal-ion-chelating residue (1), the phenolic OH function of diethyl 4-hydroxypyridine-2,6-dicarboxylate was
first derivatized by treatment with (R)-2,2-dimethyl-1,3-dioxolane-4-methanol p-toluenesulfonate (Scheme 1). The isopropylidene protection was then removed under acidic conditions and the liberated primary OH function was protected as a 4,4'-dimethoxytrityl ether. The secondary OH of the protected intermediate 3 was phosphitylated by conventional methods. Finally, the ethoxycarbonyl substituents of the pyridine ring were converted to aminocarbonyl substituents by treatment with methanolic ammonia to afford the phosphoramidite building block 1. It is worth pointing out that GNA oligonucleotides with free OH termini will undergo cleavage under these conditions.

2.2. Preparation of solid supports for the synthesis of GNA oligonucleotides

For immobilization on solid support, protected GNA counterparts of the canonical 2'-deoxynucleosides were first succinylated by treatment with succinic anhydride in anhydrous pyridine (Scheme 2). The crude products were then attached to long chain alkylamine-functionalized controlled pore glass (LCAA-CPG) by conventional HBTU-promoted peptide coupling. Based on trityl response on treatment with 3% dichloroacetic acid in CH$_2$Cl$_2$, loadings of the solid supports thus obtained were found to be in the typical range (30 – 50 μmol g$^{-1}$).

2.3. Oligonucleotide synthesis

The oligonucleotides (Table 1) were assembled by an automated synthesizer on a CPG-supported succinyl linker. Standard phosphoramidite strategy with 300 s coupling time (with 5-benzylthio-1H-tetrazole as the activator) was used throughout the sequences. Based on the trityl
response, the coupling yields were 44% for the modified building block 1 and more than 98% for the other building blocks. The oligonucleotides were released from the supports and the phosphate and base protections removed by treatment with either 16% aq. ammonia (for ON1 and ON2) or 20% aq. methylamine (for ON3) for 35 min at 55 °C. To minimize cleavage of the phosphodiester linkages, the aminolyses were carried out with the 3’-O-DMTr protection in place. This protecting group was finally removed in 80% aq. acetic acid (60 min at room temperature).

| Table 1 |

The crude oligonucleotides were purified by RP-HPLC and the purified products characterized by electrospray ionization mass spectrometry (ESI-MS). The concentrations of the oligonucleotides were estimated UV-spectrophotometrically based on absorbance at 260 nm. The molar absorptivities were calculated by an implementation of the nearest-neighbors method assuming the hypochromicities of GNA and DNA oligonucleotides to be equal.[24, 25] For determination of the molar absorptivity of the modified residue, compound 3 was deprotected and converted into the diamide 5 by successive treatments with hydrochloric acid and methanolic ammonia (Scheme 3). Molar absorptivity of 5 at 260 nm was determined to be 2000 M⁻¹ cm⁻¹ (UV spectrum presented as supporting information).

| Scheme 3 |

2.4. Preparation of Pd²⁺-carrying oligonucleotides

Oligonucleotide ON1, bearing the modified residue 5 in its 5’-terminus, was dissolved in 2 mM aq. K₂PdCl₄ to a final concentration of 1.3 mM. This mixture was incubated overnight at room temperature, after which it was diluted with phosphate buffer (pH 7.2). The product mixture was fractioned by RP-HPLC eluting with an MeCN gradient in phosphate buffer and the collected fractions desalted by RP-HPLC eluting with an MeCN gradient in water. Based on the HPLC profile,
**ON1** had been converted nearly quantitatively to products of lower mobility (Fig. 1). ESI-MS analysis revealed the main product, eluting at 12.5 min, to be a complex of two **ON1** molecules and two Pd$^{2+}$ ions (Fig. 2., **ON1$_2$:Pd$_2$**). Each of the Pd$^{2+}$ ions had displaced two protons, consistent with the binding mode reported previously for pyridine-2,6-dicarboxamide.[18] Given the self-complementarity of the tetrancleotide sequence in the middle of **ON1**, a homoduplex with terminal Pd$^{2+}$-mediated base pairs between the pyridine-2,6-dicarboxamide moiety of the modified residue 5 and thymine appears the most likely structure for **ON1$_2$:Pd$_2$**. Coordination of Pd$^{2+}$ to the N3 atom of thymine is accompanied by deprotonation,[18] resulting in a net charge of -1 for each of the Pd$^{2+}$-mediated base pairs. Based on ESI-MS, the second-most abundant product, eluting at 13.8 min, appeared to consist of three **ON1** molecules and three Pd$^{2+}$ ions (data not shown). The other minor products remained unidentified.

< Figure 1 >

< Figure 2 >

Oligonucleotide **ON2**, incorporating the modified residue 5 in the middle of the sequence, was dissolved in 2 mM aq. K$_2$PdCl$_4$, to a final concentration of 1.8 mM. This mixture was incubated overnight at room temperature, after which it was freeze-dried. An equimolar amount of the complementary oligonucleotide **ON3** was then added and the mixture diluted with phosphate buffer (pH 7.2). The RP-HPLC chromatogram of the product mixture (Fig. 3) consisted of two main peaks, of which the slower eluting one (t$_R$ = 13.0 min) was identified as **ON3** by HPLC-MS analysis, as well as spiking with an authentic sample. The faster-eluting peak (t$_R$ = 10.3 min), in turn, did not correspond to either of the starting oligonucleotides. In fact, hardly any unreacted **ON2** could be detected in the product mixture. When small volumes of the mixture were injected, the areas of the two peaks were approximately equal but upon injection of larger volumes the first peak grew at the expense of the second one (chromatograms presented as supporting information). Evidently the two peaks represented two compounds in equilibrium and increasing the concentration shifted this equilibrium
to favor the faster-eluting compound. The most likely explanation for such behavior is that the first peak, eluting faster than either ON2 or ON3, corresponded to a heterodimer of these oligonucleotides. This interpretation was confirmed by reinjection of the collected peak – in this chromatogram, peaks of the fast-eluting product and oligonucleotide ON3, but not ON2, were detected (Fig. 3D). The absence of the peak of ON2 could be explained if ON2 eluted as the Pd$^{2+}$ complex ON2:Pd and the retention time of this metallo-oligonucleotide was very similar to that of the heterodimer product. Attempts to desalt the product fraction were unsuccessful, presumably due to denaturation of the ON2:Pd:ON3 duplex and subsequent loss of the bridging Pd$^{2+}$ ion at the low ionic strength employed. A small sample was, however, treated as described above for ON1$_2$:Pd$_2$ and analyzed by ESI-MS (data presented as supporting information). Oligonucleotides ON2, ON3 and ON2:Pd were detected, suggesting that Pd$^{2+}$ is coordinated at the desired site but the ON2:Pd:ON3 duplex is not quite as stable as ON1$_2$:Pd$_2$.

2.5. Thermal stability of the modified oligonucleotide duplexes

Melting temperatures for the Pd$^{2+}$-containing duplexes ON1$_2$:Pd$_2$ and ON2:Pd:ON3, as well as their non-metallated counterparts ON1$_2$ and ON2:ON3, were measured in either a 20 mM cacodylate buffer (pH = 7.4, for ON1$_2$ and ON1$_2$:Pd$_2$) or a 20 mM phosphate buffer (pH = 7.2, for ON2:ON3 and ON2:Pd:ON3). The oligonucleotide concentrations were 6.0 μM for ON1 (3.0 μM for ON1$_2$) and 3.0 μM for ON2 and ON3. The ionic strength of the samples was adjusted to 0.1 M with NaClO$_4$. With both of the non-metallated oligonucleotide duplexes, typical sigmoidal melting curves were obtained, the $T_m$ being 45.9 °C for ON1$_2$ and 36.3 °C for ON2:ON3 (Fig. 4). The palladated duplexes, on the other hand, behaved quite differently: the melting temperature of ON1$_2$:Pd$_2$ was too high to be determined whereas with ON2:Pd:ON3 no stabilization over ON2:ON3 was observed. Similar results have previously been obtained with 2’-O-methyl-RNA oligonucleotides incorporating
2,6-bis(3,5-dimethylpyrazol-1-yl)purine as the metal-ion-chelating nucleobase. In that case, 10 °C stabilization upon addition of 1 eq. of K₂PdCl₄ was obtained with uracil as the opposite base, but only when the metal-ion-mediated base pair was placed at a terminal position.[26] Presumably both of these Pd²⁺-mediated base pairs deviate from the optimal geometry required by the base stack and this deviation is better tolerated in a terminal than in an intrastrand position.

While impossible to quantify accurately, it is evident that the difference between the thermal stabilities of ON₁₂:Pd₂ and ON₁₂ is vastly greater than what has been previously reported for any oligonucleotide structure with Pd²⁺ as the bridging metal ion. The reason for this discrepancy could be the different preparation of the samples. In the present study, Pd²⁺-carrying duplexes were synthesized and purified prior to the measurements, in contrast to the more common method of simply adding Pd²⁺ salt to a dilute sample containing both strands of the duplex. As discussed above, this method may fail to afford the thermodynamic product due to the relatively slow ligand-exchange reactions of Pd²⁺. To test this hypothesis, the melting temperature of ON₁₂ was measured in the presence of 2 eq. (6.0 μmol L⁻¹) of K₂PdCl₄ (Fig. 4A). The Tₘ was somewhat lower than the one obtained in the absence of Pd²⁺ (41.5 and 45.9 °C, respectively). Clearly, mixing dilute solutions of ON₁ and K₂PdCl₄ at neutral pH does not result in significant formation of the desired ON₁₂:Pd₂ duplex.

2.6. Secondary structure of the modified oligonucleotide duplexes

To probe the secondary structure of the modified duplexes and the possible changes induced by Pd²⁺ coordination, CD spectra of ON₁₂, ON₂:ON₃, ON₁₂:Pd₂ and ON₂:Pd:ON₃ were recorded under the same conditions as described above for the Tₘ measurements (Figs. 5 and 6). In all cases, a negative band at 275 nm was observed at low temperatures, consistent with an (S)-GNA
double helix.[27] The other previously reported negative band (at 220 nm) was hardly detectable but
\( \text{ON1}_2 \) exhibited a prominent negative band at 247 nm. This band was not observed with \( \text{ON1}_2: \text{Pd}_2 \) and the band at 275 nm was also less prominent, suggesting partial unwinding of the double helix upon formation of the terminal \( \text{Pd}^{2+} \)-mediated base pairs. In contrast, the spectra of \( \text{ON2:ON3} \) and \( \text{ON2:Pd:ON3} \) were very similar. As expected, heating resulted in gradual decrease of the CD signals with all the oligonucleotides. In the case of \( \text{ON1}_2: \text{Pd}_2 \), however, some ellipticity persisted even at 90 °C, in line with the high thermal stability of this oligonucleotide. In other words, while the terminal \( \text{Pd}^{2+} \)-mediated base pairs of \( \text{ON1}_2: \text{Pd}_2 \) disrupt the helical secondary structure, they still greatly stabilize the duplex. This stabilization is attributable to the two Pd-N bonds formed between the terminal thymine residues and the \( \text{Pd}^{2+}: \text{pyridine-2,6-dicarboxamide} \) chelates. Evidently their bond energy is greater than the energy cost for (partial) unwinding of the double helix.

\[ \text{Figure 5} \]

\[ \text{Figure 6} \]

2.7. Conclusions

Formation of \( \text{Pd}^{2+} \)-mediated base pairs within an oligonucleotide duplex has been demonstrated. At the termini of a short duplex, such base pairs may be highly stabilizing even if they are not easily accommodated in the base stack of the double helix. For realization of its potential as the bridging metal ion, however, \( \text{Pd}^{2+} \) has to be firmly attached to a predetermined site of one strand of the duplex prior to hybridization. Attempts to introduce \( \text{Pd}^{2+} \) to mixtures of oligonucleotides in situ under typical conditions of hybridization studies are not likely to afford the desired \( \text{Pd}^{2+} \)-mediated duplex in a practical yield.
3. Experimental part

3.1. General

Preparation of the GNA phosphoramidite building blocks of the four canonical nucleobases has been described in the literature.[21-23] The other reagents were commercial products that were used as received. Solvents were dried over 3 Å molecular sieves and trimethylamine over calcium hydride. $^1$H, $^{13}$C and $^{31}$P NMR spectra: Bruker Avance 500- or 400-MHz NMR spectrometer; δ in ppm rel. to Me$_4$Si (for $^1$H and $^{13}$C) or orthophosphoric acid (for $^{31}$P) as external standard, J in Hz. HR-ESI-MS: Bruker Daltonics micrOTOF-Q mass spectrometer; in m/z. UV spectra: Perkin-Elmer Lambda 35 UV-vis spectrophotometer. CD spectra: Applied Photophysics Chirascan spectropolarimeter.

3.2. Syntheses

Diethyl (S)-4-[(2,2-dimethyl-1,3-dioxolan-4-yl)methoxy]pyridine-2,6-dicarboxylate (2). 4-Hydroxypyridine-2,6-dicarboxylate (0.50 g, 2.09 mmol), (R)-2,2-Dimethyl-1,3-dioxolane-4-methanol $p$-toluenesulfonate (0.72 g, 2.5 mmol) and potassium carbonate (1.03 g, 7.45 mmol) were suspended in anhydrous DMF (20 mL). The mixture was stirred at 70 °C for 96 h, after which it was evaporated to dryness. The residue was dissolved in CH$_2$Cl$_2$ (100 mL) and washed with water (100 mL). The organic layer was dried with Na$_2$SO$_4$ and evaporated to dryness. The crude product thus obtained was purified on a silica gel column eluting with a mixture of MeOH and CH$_2$Cl$_2$ (6:94, v/v). Yield 0.61 g (82%). $^1$H-NMR (400 MHz, CDCl$_3$): 7.77 (s, 2H, Pyr-H3 & -H5), 4.48 (m, 1H, dioxolane-H5), 4.43 (q, $J=7.1$, 4H, CO$_2$C$_3$H$_7$), 4.20-4.09 (m, 3H, dioxolane-H4 & PyrOCH$_2$), (dd, $J=5.8$, 8.6, 1H, dioxolane-H4), 1.42 (s, 3H, dioxolane-CH$_3$), 1.41 (t, $J=7.1$, 6H, CO$_2$CH$_2$CH$_3$), 1.36 (s, 3H, dioxolane-CH$_3$). $^{13}$C-NMR (100 MHz, CDCl$_3$): 166.5 (2C, CONH$_2$), 164.5 (Pyr-C4), 150.3 (2C, Pyr-C2 & -C6), 114.3 (2C, Pyr-C3 & -C5), 110.1 (dioxolane-C2), 73.4 (dioxolane-C5), 69.3 (PyrOCH$_2$), 66.3 (dioxolane-C4), 62.3 (2C,
\( \text{CO}_2\text{CH}_2\text{CH}_3, 26.7 \text{ (dioxolane-CH}_3), 25.2 \text{ (dioxolane-CH}_3), 14.2 \text{ (2C, CO}_2\text{CH}_2\text{CH}_3). \) HR-ESI-MS: 354.1547 ([M + H]^+, C_{17}H_{24}NO_7; \text{calc.} 354.1547).

\( (S)-1\text{-[\{(4,4'-Dimethoxytrityl)oxy\}3-\{[2,6-bis(ethoxycarbonyl)pyridin-4-yl]oxy\}propan-2-yl}  \ (2\text{-cyanoethyl) diisopropylphosphoramidite (4).} \) Compound 2 (0.61 g, 1.73 mmol) was dissolved in a mixture of 37% aq. HCl (0.40 mL), MeCN (4.0 mL) and water (4.0 mL). The mixture was stirred at room temperature for 16 h, after which it was evaporated to dryness. The residue was coevaporated from toluene (10 mL) and anhydrous pyridine (3 \times 10 mL) and finally redissolved in anhydrous pyridine (10 mL). 4,4'-Dimethoxytrityl chloride (0.65 g, 1.9 mmol) was added and the resulting mixture stirred at room temperature for 16 h, after which it was concentrated under reduced pressure. The oily residue was diluted with \text{CH}_2\text{Cl}_2 (100 mL) and washed with saturated aq. NaHCO_3 (100 mL). The organic layer was dried with Na_2SO_4 and evaporated to dryness. The crude product thus obtained was purified on a silica gel column eluting with a mixture of Et_3N, MeOH and \text{CH}_2\text{Cl}_2 (1:3:96, \nu/\nu). The purified intermediate 3 (0.274 g, 0.445 mmol), anhydrous Et_3N (311 \mu L, 2.23 mmol) and 2-cyanoethyl-\text{N,N-diisopropylchlorophosphoramidite} (120 \mu L, 0.496 mmol) were dissolved in anhydrous \text{CH}_2\text{Cl}_2 (1.5 mL). The resulting mixture stirred under \text{N}_2 at room temperature for 2 h, after which it was diluted with \text{CH}_2\text{Cl}_2 (100 mL) and washed with saturated aq. NaHCO_3 (100 mL). The organic layer was dried with Na_2SO_4 and evaporated to dryness. NMR analysis revealed the product thus obtained to be of sufficient purity. Yield 0.3484 g (25%) as two diastereomers in approximately 1:1 ratio. \(^1\text{H-NMR (400 MHz, CDCl}_3, \text{diastereomer A)}: 7.78 (s, 2\text{H, Pyr-H3} \& -H5), 7.44 (m, 2\text{H, Ph-H2} \& -H6), 7.31 (m, 4\text{H, MeOPh-H2} \& -H6), 7.27 (m, 2\text{H, Ph-H3} \& -H5), 7.23 (t, J = 5.0, 1\text{H, Ph-H4}), 6.80 (d, J = 8.5, 4\text{H, MeOPh-H3 and} \& -H6), 4.48 (q, J = 7.1, 4\text{H, CO}_2\text{CH}_2\text{CH}_3), 4.37-4.18 (m, 3\text{H, PyrOCH}_2 \& \text{CH}_2\text{CH(OP)CH}_2), 3.78 (s, 6\text{H, OCH}_3), 3.74 (m, 1\text{H, POCH}_2), 3.70 (m, 1\text{H, POCH}_2), 3.66-3.47 (m, 2\text{H, NCHCH}_3), 3.36 (dd, J = 9.6, 5.1, 1\text{H, DMTroOCH}_2), 3.29 (dd, J = 9.4, 6.2, 1\text{H, DMTroOCH}_2), 2.61 (t, J = 6.4, 2\text{H, CH}_2\text{CN}), 1.46 (t, J = 7.0, 6\text{H, CO}_2\text{CH}_2\text{CH}_3), 1.14 (d, J = 5.6, 6\text{H, NCHCH}_3), 1.07 (d, J = 6.7, 6\text{H, NCHCH}_3). \(^1\text{H-NMR (400 MHz, CDCl}_3, \text{diastereomer B)}: 7.74 (s, 2\text{H, Pyr-H3} \& -H5), 7.42 (m, 2\text{H, Ph-H2} \& -H6), 7.31 (m, 4\text{H, MeOPh-H2} \& -H6), 7.27 (m, 2\text{H, Ph-H3} \& -H5), 7.19 (t, J = 6.3, 1\text{H, Ph-H4}), 6.82
(d, J = 8.1, 4H, MeOPh-H3 and & -H5), 4.48 (q, J = 7.1, 4H, CO2CH2CH3), 4.37-4.18 (m, 3H, PyrOCH2 & CH2CH(OP)CH2), 3.82 (m, 1H, POCH2), 3.78 (s, 6H, OCH3), 3.75 (m, 1H, POCH2), 3.66-3.47 (m, 2H, NCHCH3), 3.40 (m, 2H, DMTrOCH2), 2.47 (t, J = 6.5, 2H, CH2CN), 1.46 (t, J = 7.2, 6H, CO2CH2CH3), 1.18 (d, J = 7.2, 6H, NCHCH3), 1.16 (d, J = 6.1, 6H, NCHCH3). 13C-NMR (100 MHz, CDCl3, diastereomer A): 166.8 (2C, CONH2), 164.7 (Pyr-C4), 158.5 (2C, MeOPh-C4), 150.2 (2C, Pyr-C2 & -C6), 144.7 (Ph-C1), 135.9 (MeOPh-C1), 135.8 (MeOPh-C1), 130.0 (2C, MeOPh-C2 & -C6), 128.1 (2C, Ph-C3 & -C5), 127.8 (2C, Ph-C2 & -C6), 126.8 (Ph-C4), 117.6 (CN), 114.5 (2C, Pyr-C3 & -C5), 113.1 (4C, MeOPh-C3 & -C5), 86.2 (Ar3C), 71.5 (d, J = 16.9, CH2CH(OP)CH2), 69.4 (d, J = 4.5, PyrOCH2), 63.1 (d, J = 4.1, DMTrOCH2), 62.4 (2C, CO2CH2CH3), 58.2 (d, J = 18.4, POCH2), 55.2 (2C, OCH3), 43.2 (d, J = 12.7, 2C, NCHCH3), 24.6 (2C, NCHCH3), 24.5 (2C, NCHCH3), 20.2 (d, J = 7.1, CH2CN), 14.2 (2C, CO2CH2CH3). 13C-NMR (100 MHz, CDCl3, diastereomer B): 166.7 (2C, CONH2), 164.7 (Pyr-C4), 158.6 (2C, MeOPh-C4), 150.2 (2C, Pyr-C2 & -C6), 144.6 (Ph-C1), 135.8 (MeOPh-C1), 135.8 (MeOPh-C1), 130.0 (4C, MeOPh-C2 & -C6), 128.1 (2C, Ph-C3 & -C5), 127.8 (2C, Ph-C2 & -C6), 126.9 (Ph-C4), 117.5 (CN), 114.3 (2C, Pyr-C3 & -C5), 113.1 (4C, MeOPh-C3 & -C5), 86.3 (Ar3C), 71.2 (d, J = 16.0, CH2CH(OP)CH2), 69.2 (d, J = 2.3, PyrOCH2), 63.0 (d, J = 3.9, DMTrOCH2), 62.4 (2C, CO2CH2CH3), 58.3 (d, J = 18.8, POCH2), 55.2 (2C, OCH3), 43.2 (d, J = 12.5, 2C, NCHCH3), 24.7 (2C, NCHCH3), 24.6 (2C, NCHCH3), 20.3 (d, J = 6.9, CH2CN), 14.2 (2C, CO2CH2CH3). 31P-NMR (162 MHz, CDCl3, diastereomer A): 149.9. 31P-NMR (162 MHz, CDCl3, diastereomer B): 150.1. HR-ESI-MS: 838.3376 ([M + Na]+, C44H54N3NaO10P+; calc. 838.3439).

(S)-1-[(4,4´-Dimethoxytrityl)oxy]-3-[(2,6-dicarbamoylpyridin-4-yl)oxy]propan-2-yl (2-cyanoethyl) diisopropylphosphoramide (1). Compound 4 (115 mg, 0.141 mmol) was dissolved in 7 M methanolic ammonia. The mixture was stirred at room temperature for 16 h, after which all volatiles were removed in vacuum. Yield 106 mg (near quantitative). 1H-NMR (500 MHz, CDCl3, diastereomer A): 7.87 (s, 2H, Pyr-H3 & -H5), 7.67 (d, J = 3.4 Hz, 2H, CONH), 7.45 (m, 2H, Ph-H2 & -H6), 7.33 (m, 4H, MeOPh-H2 & -H6), 7.28 (m, 2H, Ph-H3 & -H5), 7.22 (t, J = 7.3, 1H, Ph-H4), 6.85 (d, J = 8.5, 4H, MeOPh-H3 and & -H5), 5.94 (d, J = 3.7, 2H, CONH), 4.46 (m, 1H, CH2CH(OP)CH3), 4.33 (m,
2H, PyrOCH$_3$), 3.82 (s, 6H, OCH$_3$), 3.77 (m, 1H, POCH$_3$), 3.71 (m, 1H, POCH$_3$), 3.67-3.50 (m, 2H, NCH$_3$), 3.36 (dd, $J = 9.9, 4.9, 1H$, DMTrOCH$_3$), 3.29 (dd, $J = 9.4, 5.9, 1H$, DMTrOCH$_3$), 2.50 (ddd, $J = 6.5, 6.3, 2.1, 2H$, CH$_2$CN), 1.17 (d, $J = 7.0, 6H$, NCH$_3$), 1.08 (d, $J = 6.7, 6H$, NCH$_3$). $^1$H NMR (500 MHz, CDCl$_3$, diastereomer B): 7.85 (s, 2H, Pyr-H3 & -H5), 7.66 (d, $J = 3.7$ Hz, 2H, CONH), 7.44 (m, 2H, Ph-H2 & -H6), 7.33 (m, 4H, MeOPh-H2 & -H6), 7.28 (m, 2H, Ph-H3 & -H5), 7.23 (t, $J = 7.3, 1H$, Ph-H4), 6.83 (d, $J = 8.8, 4H$, MeOPh-H3 and & -H5), 5.92 (d, $J = 3.7, 2H$, CONH), 4.32 (m, 2H, PyrOCH$_3$), 4.26 (m, 1H, CH$_2$C$(OP)$CH$_2$), 3.84 (m, 1H, POC$_3$H$_2$), 3.81 (s, 6H, OC$_3$H$_3$), 3.78 (m, 1H, POC$_3$H$_2$), 3.67-3.50 (m, 2H, NC$_3$H$_3$), 3.43 (dd, $J = 9.7, 4.4, 1H$, DMTrOCH$_3$), 3.29 (dd, $J = 10.5, 5.6, 1H$, DMTrOCH$_3$), 2.65 (ddd, $J = 6.6, 0.7, 2H$, CH$_2$CN), 1.19 (d, $J = 7.1, 6H$, NCH$_3$), 1.18 (d, $J = 7.3, 6H$, NCH$_3$). $^{13}$C-NMR (125 MHz, CDCl$_3$, diastereomer A): 167.6 (Pyr-C4), 165.7 (2C, CONH), 158.5 (2C, MeOPh-C4), 150.5 (2C, Pyr-C2 & -C6), 144.7 (Ph-C1), 135.8 (2C, MeOPh-C1), 130.0 (2C, MeOPh-C2 & -C6), 130.0 (2C, MeOPh-C2 & -C6), 128.1 (2C, Ph-C3 & -C5), 127.8 (2C, Ph-C2 & -C6), 126.8 (Ph-C4), 117.7 (CN), 113.1 (4C, MeOPh-C3 & -C5), 111.9 (2C, Pyr-C3 & -C5), 86.2 (Ar$_2$C), 71.1 (d, $J = 16.1$, CH$_2$CH(OP)CH$_2$), 69.3 (d, $J = 2.2$, PyrOCH$_3$), 63.1 (d, $J = 3.8$, DMTrOCH$_3$), 58.3 (d, $J = 18.4$, POCH$_3$), 55.2 (2C, OCH$_3$), 43.2 (d, $J = 12.6$, 2C, NCH$_3$), 24.6 (2C, NCH$_3$), 24.5 (2C, NCH$_3$), 20.3 (d, $J = 6.9$, CH$_2$CN). $^{13}$C-NMR (125 MHz, CDCl$_3$, diastereomer B): 167.7 (Pyr-C4), 165.6 (2C, CONH), 158.5 (2C, MeOPh-C4), 150.5 (2C, Pyr-C2 & -C6), 144.7 (Ph-C1), 135.9 (MeOPh-C1), 135.8 (MeOPh-C1), 130.1 (4C, MeOPh-C2 & -C6), 128.1 (2C, Ph-C3 & -C5), 127.9 (2C, Ph-C2 & -C6), 126.9 (Ph-C4), 117.5 (CN), 113.1 (4C, MeOPh-C3 & -C5), 111.8 (2C, Pyr-C3 & -C5), 86.3 (Ar$_2$C), 71.4 (d, $J = 16.8$, CH$_2$CH(OP)CH$_2$), 69.5 (d, $J = 4.7$, PyrOCH$_3$), 63.0 (d, $J = 3.1$, DMTrOCH$_3$), 58.3 (d, $J = 18.9$, POCH$_3$), 55.2 (2C, OCH$_3$), 43.2 (d, $J = 12.5$, 2C, NCH$_3$), 24.7 (2C, NCH$_3$), 24.6 (2C, NCH$_3$), 20.2 (d, $J = 7.1$, CH$_2$CN). $^{31}$P-NMR (202 MHz, CDCl$_3$, diastereomer A): 149.8. $^{31}$P-NMR (202 MHz, CDCl$_3$, diastereomer B): 150.1. HR-ESI-MS: 780.3130 ([M + Na]$^+$, C$_{46}$H$_{48}$N$_{5}$NaO$_{8}$P$^+$; calc. 780.3138).

(R)-4-(2,3-Dihydroxypropoxy)pyridine-2,6-dicarboxamide (5). Compound 2 (0.50 g, 1.41 mmol) was dissolved in a mixture of 37% aq. HCl (0.50 mL), MeCN (5.0 mL) and water (5.0 mL). The mixture was stirred at room temperature for 16 h, after which it was evaporated to dryness. The
residue was dissolved in 7 M methanolic ammonia and the mixture stirred at room temperature for 16 h, after which all volatiles were removed in vacuum. Yield 0.36 g (near quantitative). $^1$H NMR (500 MHz, (D$_6$)DMSO): 8.85 (s, 2H, NH$_2$), 7.71 (s, 2H, NH$_2$), 7.67 (s, 2H, Pyr-H3 & -H5), 5.09 (s, 1H, CH$_2$CH(OH)CH$_2$), 4.76 (s, 1H, CH$_2$OH), 4.25 (dd, 1H, $J$ = 10.2, 3.0, PyrOCH$_3$), 4.09 (dd, 1H, $J$ = 9.0, 6.6, PyrOCH$_3$), 3.84 (m, H, CH$_2$CH(OH)CH$_2$), 3.47 (m, 2H, HOC$_2$H). $^{13}$C NMR (125 MHz, (D$_6$)DMSO): 167.8 (Pyr-C4), 165.7 (2C, CONH$_3$), 151.6 (2C, Pyr-C2 & -C6), 110.7 (2C, Pyr-C3 & -C5), 70.9 (PyrOCH$_3$), 70.1 (CH$_2$CH(OH)CH$_2$), 62.8 (HOCH$_2$). HR-ESI-MS: 278.0742 ([M + Na]$^+$, C$_{10}$H$_{13}$N$_3$NaO$_5$$^+$; calc. 278.0747).

**General procedure for preparation of solid supports for the synthesis of GNA oligonucleotides.** An appropriately protected GNA counterpart of a 2’-deoxynucleoside (0.15 mmol) was dissolved in anhydrous pyridine (8.0 mL). Succinic anhydride (0.60 mmol) and DMAP (cat.) were added and the resulting mixture stirred at room temperature for 16 h, after which it was evaporated to dryness. The residue was dissolved in CH$_2$Cl$_2$ (40 mL) and washed with saturated aq. NaHCO$_3$ (40 mL). The organic layer was dried with Na$_2$SO$_4$ and evaporated to dryness. An approximately 20 μmol portion of the crude product thus obtained was dissolved in anhydrous DMF (0.50 mL). HBTU (7.6 mg, 20 μmol) and DIPEA (7.0 μL, 41 μmol) were added and the resulting solution transferred to a microcentrifuge tube containing long chain alkylamine-CPG (100 mg). The tube was shaken at room temperature for 24 h, after which the CPG was transferred to an oligonucleotide synthesizer column and washed with anhydrous DMF (15 mL) and MeCN (15 mL). The unreacted amino groups of the CPG support were acetylated by treatment with a mixture of 2,6-lutidine (0.20 mL), Ac$_2$O (0.20 mL), N-methylimidazole (0.32 mL) and THF (3.28 mL) at room temperature for 3 min. Finally, the support was washed with anhydrous MeCN (15 mL) and CH$_2$Cl$_2$ (15 mL).

For quantification of its loading, 5.0 mg of the CPG support was placed in a microcentrifuge tube and 3% solution of dichloroacetic acid in CH$_2$Cl$_2$ (1.0 mL) was added. The resulting orange solution was separated and the above treatment repeated until the acidic solution remained colourless. The aliquots were then combined and diluted to 25.0 mL with CH$_2$Cl$_2$. The absorbance of
this solution was then measured at 503 nm and the loading calculated based on the known molar absorptivity of 0.076 L μmol⁻¹ cm⁻¹. The loadings obtained by this method were 46.1, 48.6, 41.9 and 38.9 μmol g⁻¹ for A, C, G and T, respectively.

**Oligonucleotide synthesis.** The oligonucleotides were assembled by an automated synthesizer on CPG supports prepared as described above. Conventional phosphoramidite strategy with 5-benzylthio-1H-tetrazole as the activator and 300 s coupling time was used throughout the sequences. Based on the trityl response, the coupling yields for the modified building block 1 and the canonical building blocks were 44% and more than 98%, respectively. Oligonucleotides ON1 and ON2 were released from the supports and the phosphate and base protections removed by a 35 min treatment with 16% aq. ammonia at 55 °C. In the case of ON3, 20% aq. methylamine was used instead. To prevent nucleophilic attack of 3'-OH on the vicinal phosphate during aminolysis, this step was carried out with the 3'-O-DMTr protection in place. This protecting group was finally removed by a 60 min treatment with 80% aq. acetic acid at room temperature.

The crude oligonucleotides were purified by RP-HPLC on a Thermo Scientific Aquasil C18 column (150 × 4 mm, 5 μm), eluting with a mixture of acetonitrile (linear gradient of 0 to 30% over 30 min) and 0.10 M triethylammonium acetate. The flow rate was 1.0 mL min⁻¹ and the detection wavelength 260 nm. The purified oligonucleotides were characterized by electrospray ionization mass spectrometry (ESI-MS, mass spectra presented as supporting information) and their concentrations estimated UV-spectrophotometrically based on absorbance at 260 nm. The molar absorptivities were calculated by an implementation of the nearest-neighbors method assuming the hypochromicities of GNA and DNA oligonucleotides to be equal.[24, 25]

**Preparation of the Pd²⁺-carrying oligonucleotide ON1₂:Pd₂.** A 215 μM aq. solution of oligonucleotide ON1 (300 μL, 64.5 nmol) was lyophilized and the residue dissolved in 2.0 mM aq. solution of K₂PdCl₄ (48.4 μL, 96.8 nmol). After vigorous vortexing, the mixture was incubated at room temperature for 16 h. Phosphate buffer (48.4 μL, 120 mM, pH 7.2) was added and the mixture
incubated at room temperature for another 12 h. The product mixture was fractioned by RP HPLC on a Thermo Scientific Aquasil C18 column (150 × 4 mm, 5 μm), eluting with a mixture of acetonitrile (linear gradient of 0 to 30% over 30 min) and 60 mM phosphate buffer (pH 7.2). The flow rate was 1.0 mL min⁻¹ and the detection wavelength 260 nm. The main fraction, eluting at 12.5 min, was collected and desalted by RP HPLC on the same column, eluting with a mixture of acetonitrile (0% for the first 6 min, followed by a linear gradient to 60% over 0.5 min and isocratic elution at 60% for 9 min) and water. Formation of the desired structure ON1₂:Pd₂ was verified by ESI-MS (mass spectrum presented as supporting information).

Preparation of the Pd²⁺-carrying oligonucleotide ON2:Pd:ON3. A 399 μM aq. solution of oligonucleotide ON2 (300 μL, 119.7 nmol) was lyophilized and the residue dissolved in 2.0 mM aq. solution of K₂PdCl₄ (65.8 μL, 131.6 nmol). After vigorous vortexing, the mixture was incubated at room temperature for 16 h and then lyophilized. The residue was dissolved in a 280 μM aq. solution of oligonucleotide ON3 (427.5 μL, 119.0 nmol) and the resulting solution diluted with phosphate buffer (427.5 μL, 120 mM, pH 7.2). After incubation for 1 h at room temperature, the product mixture was fractioned by RP HPLC under the same conditions as described above for ON1₂:Pd₂. The main fraction, eluting at 10.4 min, was collected and used as such in the hybridization studies. Attempts to desalt ON2:Pd:ON3 were unsuccessful. For characterization, a small sample was treated as described above for ON1₂:Pd₂ and analyzed by ESI-MS (data presented as supporting information). Oligonucleotides ON2, ON3 and ON2:Pd were detected, consistent with formation of the desired structure ON2:Pd:ON3.

References


Table 1 Structures of the GNA oligonucleotides used in this study.

Scheme 1 Synthesis of the modified GNA phosphoramidite building block 1. Reagents and conditions: (a) K₂CO₃, DMF; (b) HCl, MeCN, H₂O; (c) DMTrCl, pyridine; (d) 2-cyanoethyl-N,N-diisopropylchlorophosphoramidite, Et₃N, CH₂Cl₂; (e) NH₃, MeOH.

Scheme 2 Preparation of solid supports for the synthesis of GNA oligonucleotides. Reagents and conditions: (a) succinic anhydride, DMAP, pyridine; (b) LCAA-CPG, HBTU, DIPEA, DMF.

Scheme 3 Synthesis of the unprotected monomer 5. Reagents and conditions: (a) HCl, MeCN, H₂O; (b) NH₃, MeOH.

Figure 1 HPLC traces for (A) purified ON1 and (B) product mixture of the reaction of ON1 with 1.5 eq. of K₂PdCl₄; Thermo Scientific Aquasil C18 column (150 × 4 mm, 5 μm); flow rate = 1.0 mL min⁻¹; linear gradient (0 to 30% over 30 min) of MeCN in 60 mM phosphate buffer (pH = 7.2).

Figure 2 (A) Proposed structure of ON1₂:Pd₂ and (B) calculated and observed mass spectra for its tetraanion.

Figure 3 HPLC traces for (A) purified ON2, (B) purified ON3, (C) product mixture of the reaction of ON2 and 1.1 eq. of K₂PdCl₄, followed by 1.0 eq. of ON3 and (D) reinjection of the faster-eluting peak
of chromatogram C; Thermo Scientific Aquasil C18 column (150 × 4 mm, 5 μm); flow rate = 1.0 mL min⁻¹; linear gradient (0 to 30% over 30 min) of MeCN in 60 mM phosphate buffer (pH = 7.2).

**Figure 4** UV melting profiles for (A) \( \text{ON}_1\text{ ON}_2 \) (●), \( \text{ON}_1\text{ ON}_2:\text{Pd}_2 \) (□) and \( \text{ON}_1 \) with 2.0 eq. of \( \text{K}_2\text{PdCl}_4 \) added \textit{in situ} (△) and (B) \( \text{ON}_2:\text{ON}_3 \) (●) and \( \text{ON}_2:\text{Pd}:\text{ON}_3 \) (□); \([\text{ON}_1] = 6.0 \ \mu\text{M}; [\text{ON}_2] = [\text{ON}_3] = 3.0 \ \mu\text{M}; [\text{Pd}^{2+}] = 0 / 3.0 / 6.0 \ \mu\text{M}; I(\text{NaClO}_4) = 0.10 \ \text{M}; \text{pH} = 7.4.

**Figure 5** CD spectra of oligonucleotide duplexes (A) \( \text{ON}_1\text{ ON}_2 \) and (B) \( \text{ON}_1\text{ ON}_2:\text{Pd}_2 \), recorded at 10 °C intervals between 10 and 90 °C; \([\text{ON}_1\text{ ON}_2] = [\text{ON}_1\text{ ON}_2:\text{Pd}_2] = 3.0 \ \mu\text{M}; \text{pH} = 7.4 \ (20 \ \text{mM cacodylate buffer}); I(\text{NaClO}_4) = 0.10 \ \text{M}.

**Figure 6** CD spectra of oligonucleotide duplexes (A) \( \text{ON}_2\text{ ON}_3 \) and (B) \( \text{ON}_2\text{ Pd:ON}_3 \), recorded at 10 °C intervals between 10 and 90 °C; \([\text{ON}_2\text{ ON}_3] = [\text{ON}_2\text{ Pd:ON}_3] = 3.0 \ \mu\text{M}; \text{pH} = 7.2 \ (20 \ \text{mM phosphate buffer}); I(\text{NaClO}_4) = 0.10 \ \text{M}. 
Figure 1
Figure 2

A

\[
5' - \text{X CGCGT} - 3' \\
\text{Pd} \quad \text{Pd} \\
3' - \text{TGGCCX} - 5' \\
\]

\[\text{ON1}_{2}\text{Pd}_2\]

B

Calculated for \(\text{ON1}_{2}\text{Pd}_2\) 

Observed
Figure 3
Figure 4
Figure 5
Figure 6
Scheme 1
Scheme 2
Scheme 3
Table 1

<table>
<thead>
<tr>
<th>Sequence</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>ON1</strong></td>
<td>5’-XCGCGT-3’</td>
</tr>
<tr>
<td><strong>ON2</strong></td>
<td>5’-CGCGXGC-3’</td>
</tr>
<tr>
<td><strong>ON3</strong></td>
<td>5’-GCCTGCXGC-5’</td>
</tr>
</tbody>
</table>

The symbol 'X' represents a specific modification in the sequence.
Graphical abstract

Terminal Pd\(^{2+}\)-mediated base pairs between pyridine-2,6-dicarboxamide and thymine greatly enhance the thermal stability of short GNA oligonucleotides relative to their metal-free counterparts. The effect was only observed with metallated oligonucleotides isolated prior to the hybridization studies, not when Pd\(^{2+}\) salt was added to the oligonucleotide solution \textit{in situ}.
HIGHLIGHTS

- Pd\(^{2+}\)-mediated base-pairing within oligonucleotides has been demonstrated.
- At the termini of a duplex, Pd\(^{2+}\)-mediated base-pairing is highly stabilizing.
- Mixing dilute oligonucleotide and Pd\(^{2+}\) solutions may not afford the desired structure.