

## **A test of the hierarchical model of litter decomposition**

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# 1 **A test of the hierarchical model of litter decomposition**

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37 **Our basic understanding of plant litter decomposition informs the assumptions underlying**  
38 **widely applied soil biogeochemical models, including those embedded in Earth system**  
39 **models. Confidence in projected carbon cycle-climate feedbacks therefore depends on**  
40 **accurate knowledge about the controls regulating the rate at which plant biomass is**  
41 **decomposed into products such as CO<sub>2</sub>. Here, we test underlying assumptions of the**  
42 **dominant conceptual model of litter decomposition. The model posits that a primary**  
43 **control on the rate of decomposition at regional to global scales is climate (temperature and**  
44 **moisture), with the controlling effects of decomposers negligible at such broad spatial**  
45 **scales. Using a regional-scale litter decomposition experiment at six sites spanning from**  
46 **northern Sweden to southern France – and capturing both within and among site variation**  
47 **in putative controls – we find that contrary to predictions from the hierarchical model,**  
48 **decomposer (microbial) biomass strongly regulates decomposition at regional scales.**  
49 **Further, the size of the microbial biomass dictates the absolute change in decomposition**  
50 **rates with changing climate variables. Our findings suggest the need for revision of the**  
51 **hierarchical model, with decomposers acting as both local- and broad-scale controls on**  
52 **litter decomposition rates, necessitating their explicit consideration in global**  
53 **biogeochemical models.**

54  
55 The dominant conceptual model of litter decomposition posits that the primary controls on the  
56 rate of decomposition are climate, litter quality and decomposer organisms<sup>1</sup>. These controls are  
57 hypothesized to operate hierarchically in space, with climate and litter quality co-dominant at  
58 regional to global scales<sup>2-4</sup>, and decomposers operating only as an additional local control whose  
59 effect is negligible at broader scales<sup>5</sup>. Consequently decomposers have been omitted as controls

60 from biogeochemical models, whereas a recent surge of interest in their inclusion has shown that  
61 carbon-cycle projections depend strongly on whether and how microbial decomposers are  
62 represented<sup>6-9</sup>. Yet evidence that microbial decomposers regulate decomposition rates at  
63 regional- to global-scales, independent of climate variables such as temperature and moisture, is  
64 generally lacking. One possibility for this lack of evidence is suggested by scaling theory, where  
65 the influence of mechanisms that act locally can be obscured in emergent, broad-scale patterns<sup>10</sup>.

66       Pattern and scale has been described as the central issue in ecology, where the inherent  
67 challenge to prediction and understanding lies in the elucidation of mechanisms, which  
68 commonly operate at different scales to those on which the patterns are observed<sup>10</sup>. This scale  
69 mismatch appears true for at least some ecosystem processes, such as plant productivity<sup>10,11</sup>.  
70 Decomposition processes, also, are controlled by variables operating at finer scales than those at  
71 which the variables are typically measured and evaluated<sup>1</sup>. For example, extensive empirical  
72 support for the hierarchical model of litter decomposition has been provided through multi-site  
73 climate gradient studies<sup>12-15</sup>. These multi-site studies have some common characteristics, which  
74 include collecting few observations (typically 2 to 4 per site per litter species per collection) –  
75 from which a mean decomposition rate is determined – and also use of site-mean data to estimate  
76 climatic controls<sup>1</sup>. Yet the hierarchical model, and its representation in the structure of  
77 biogeochemical models, is based on the assumption that controls act at the microsite level, by  
78 regulating the activities of decomposer organisms<sup>5,16</sup>. That is, the hierarchical model is  
79 conceptually grounded in local (i.e. microsite) dynamics, but has been developed and  
80 substantiated with site-mean data that represents climate control of decomposition as an among-  
81 site relationship.

82           Understanding controls on litter decomposition across regional scales is then necessarily  
83 intertwined with scaling theory. This body of theory<sup>10</sup> suggests that broad-scale patterns might  
84 emerge from distinct, local-scale causative relationships, which contrasts with the assumption of  
85 the hierarchical model that among-site patterns in decomposition approximate patterns operating  
86 at the microsite (Fig. 1). We refer to this as the “assumption of scale invariance” (Fig. 2a). Two  
87 lines of evidence question the validity of the assumption of scale invariance for litter  
88 decomposition. The first is that the activities of decomposer communities are shaped by  
89 environmental selection for a subset of functional traits, which then uniquely dictate how  
90 decomposition rates respond to changing climatic controls<sup>17-20</sup>. The second is that microclimate  
91 can vary widely within a site<sup>21,22</sup>. As such, site-mean climate data are likely a poor surrogate for  
92 the range in microclimate experienced by decomposer organisms within a site<sup>21</sup>. Both lines of  
93 evidence support the possibility that among-site patterns in decomposition rates emerge from  
94 distinct microsite-level relationships (the “assumption of scale dependence”, Fig. 2b).

95           We use a multi-site, litter decomposition study to test between the competing  
96 assumptions of scale invariance and dependence (Figs. 1,2). We worked across a climate  
97 gradient in Europe at six grassland sites spanning boreal climate in northern Sweden to  
98 Mediterranean climate in southern France. We predicted two specific patterns would emerge if  
99 the assumption of scale invariance were to be falsified. Prediction 1 was that relationships  
100 between climate and decomposition rates should differ when site-mean versus microsite-level  
101 climate data are analysed. That is, the emergent regional-scale pattern from microclimate data  
102 should differ from the pattern observed with site-mean climate data. Prediction 2 was that any  
103 variable expected to be an important control at the microsite-level (e.g. microbial biomass),  
104 should have a strong effect when regional-scale patterns are analysed using microsite-level data.

105 Litter quality was included in our experimental design, by using two grass species with  
106 contrasting litter functional traits, but was not under test. Instead, standardizing known  
107 controlling variables can improve estimated effects of other controls under study. In addition,  
108 litter traits are expected to interact with controls such as temperature<sup>23</sup> and so including this  
109 variable allowed us to test this possibility. In total, we measured four controls (temperature,  
110 moisture, microbial biomass and soil nitrogen availability) that naturally varied among  
111 microsites. All four variables are expected to act as strong local and, in the case of the climate  
112 variables, broad-scale controls on decomposition<sup>1,5,24,25</sup>. We then built a set of regression models,  
113 structured to represent and test between assumptions of scale invariance versus dependence in  
114 controls (see Methods), to compare the estimated effect sizes of these different variables on litter  
115 decomposition rates.

116

## 117 **Results and discussion**

118 Decomposition rates varied within and among sites and between the two litter types (Fig. 3a,b).  
119 As expected, mass carbon (C) loss over the 3-month field incubations was approximately twice  
120 as great for the higher quality *Holcus* litter ( $33.8 \pm 11.62\%$ ; mean  $\pm$  SD) than for the *Festuca* litter  
121 ( $16.8 \pm 7.15\%$ ). However, there was considerable variation, with loss rates for *Holcus* ranging  
122 from 7.72 to 53.7%, and for *Festuca* from 0.50 to 35.3%. Similarly there was marked variation in  
123 the values of the climate controls, temperature and moisture, although they had contrasting  
124 within versus among site distributions. Soil temperatures clustered within sites, meaning that  
125 variation was much greater among sites (Fig. 3c), ranging from 10.0 to 25.3°C for the most  
126 northern to southern site means. In contrast, microsite litter moisture only clustered around the  
127 site mean at the two most southern sites, where mean site moisture was lowest (11.7 and 7.5%).

128 At the most northern site the mean moisture was 51.6% but varied among microsites from 12.8  
129 to 81.3% (Fig. 3d). Microsite soil nitrogen (N) availability and microbial biomass were more  
130 clustered than moisture but within- versus among-site variation was still large (Figs. 3e,f). Soil N  
131 varied among sites from means of 9.0 to 32.8  $\mu\text{g N g soil}^{-1}$  but within the most northern site  
132 alone from 2.3 to 70.6  $\mu\text{g N g soil}^{-1}$ . Equally, microbial biomass site means varied ~2-times from  
133 0.96 to 2.03  $\mu\text{g CO}_2 \text{ g soil}^{-1} \text{ h}^{-1}$ , but within sites from about 1.6-times (most northern) to about  
134 2.75-times (most southern).

135 Prediction 1 was that emergent patterns between mean-site climate and decomposition  
136 might fail to capture relationships occurring at the microsite scale. We found no support for this  
137 prediction for temperature, with the “Microclimate” and “Site-mean climate” models (see  
138 Methods) giving similar temperature coefficients (Table 1) and effect sizes (Fig. 4a). That is, the  
139 temperature-decomposition relationship was scale invariant (Fig. 1). This perhaps is not  
140 surprising given that microsite soil temperature clustered around the site mean (Fig. 3c).  
141 Consequently the regional temperature-decomposition relationship should be, and was,  
142 approximately equivalent whether microsite or site-mean values were explored (Fig. 4a). There  
143 is evidence that microsite temperature can differ markedly to the site mean in some  
144 environmental contexts<sup>22</sup>. However across 60 sites spanning a broad range in eco-climatic  
145 conditions, Loescher *et al.*<sup>21</sup> found that microsite soil temperatures were representative of the site  
146 mean, suggesting that our finding that the temperature-decomposition relationship is scale  
147 invariant might generalize to numerous ecosystem types.

148 In contrast, the moisture-decomposition relationship was strongly scale dependent: there  
149 was a pronounced moisture-decomposition relationship for the Microsite model but a weak one  
150 for the emergent pattern estimated from the Site-mean model (Table 1, Fig. 4b). Specifically,



151 across the large observed range of microsite moisture availability (5.7 to 83.2%), the Site-mean  
152 model projected mass loss values ranging from a low of 27.4% to a high of 28.7%. In contrast,  
153 the Microclimate model estimated a shift in decomposition across the same range in moisture  
154 from 23.9 to 33.2% mass loss (Fig. 4b). Site means therefore poorly captured regional  
155 heterogeneity in microsite moisture availability, generating a scale mismatch between local  
156 mechanism and broad-scale pattern. Our data (Fig. 4b) consequently suggest that patterns  
157 emerging from among-site comparisons of site-mean moisture may fail to represent causative  
158 relationships operating at the much finer spatial scales at which decomposer organisms respond  
159 to the environment. These findings raise questions about the use of site-mean (or coarser  
160 resolution) hydroclimatic data to parameterise ecosystem models. Overall, our data suggest that  
161 assumptions of the hierarchical model about scale invariance in climatic control are variable  
162 dependent, cautioning against its general application as a conceptual and numerical  
163 representation of controls on decomposition.

164         Using the “Microsite interactions” model (see Methods), we evaluated Prediction 2 that  
165 variables considered locally important should retain a strong influence at broad spatial scales.  
166 Following this prediction, the effect size of microbial biomass on decomposition rates was of  
167 similar magnitude to those for the climatic variables (Fig. 5a). Specifically, estimated  
168 decomposition rates varied by ~16% mass C loss with temperature change, ~11% with moisture  
169 change, and ~12% with microbial biomass change (Fig. 5a). Not surprisingly, given that we  
170 experimentally generated marked differences in litter quality, estimated mass loss increased  
171 ~24% (from 17 to 41%) with increasing initial litter N (Fig. 5a). The soil N effect size was by  
172 contrast small, leading to about a 2% positive change in estimated mass C loss but, as with all the  
173 other variables, the main effect coefficient was significant ( $P < 0.05$ ; Table 1, Fig. 5a). Although

174 some 2-way interaction coefficients were of comparable or greater magnitude to the main effects  
175 for temperature, moisture and microbial biomass (Table 1), qualitatively the estimated effect  
176 sizes of these variables from the Microsite interactions and Microsite main effects models were  
177 similar (Figs. 4, 5b). That is, when interactions were removed, litter quality, temperature,  
178 moisture and microbial biomass all retained strong control on decomposition at the regional scale  
179 of our study (Table 1, Supplementary Fig. 1).

180 Exclusion of soil animal decomposers does alter litter decomposition rates in at least  
181 some biomes<sup>15,24,26-28</sup> but microbial effects were not explicitly examined. However, the  
182 representation of microbial biomass or growth in biogeochemical models can improve predictive  
183 power<sup>9,29</sup> and such variables are argued to relate most directly to spatial and temporal variation in  
184 biogeochemical process rates<sup>7,8,30</sup>. In support of these arguments, the absolute size of our  
185 estimated effects of microclimate on decomposition depended strongly on microbial biomass.  
186 Specifically, using the Microsite interactions model we set microbial biomass at five values  
187 representing the observed range of microsite variation, and then varied temperature and moisture  
188 (Fig. 5c,d). Higher microbial biomass values generated a much greater absolute change in  
189 decomposition rates with increasing temperature or moisture (Fig. 5c,d). For example, estimated  
190 mass loss rates across the microsite moisture range only varied by ~5% in absolute terms when  
191 microbial biomass was low, to as much as ~25% (from 28.5 to 54.2% mass loss) when it was  
192 high. This influence of microbial biomass was primarily additive given that, when it was dropped  
193 from the modelling (giving the Microclimate model), there was minimal influence on the relative  
194 effect sizes of litter quality, temperature and moisture (Fig. 4, Table 1). An outstanding question  
195 is whether the microbial traits selected by a site's climatic context<sup>17,18</sup> in turn influence the  
196 magnitude of microclimate effects on decomposition, as is similarly observed through climate

197 selection of plant functional traits<sup>23,31</sup>. Nevertheless, our data do support emerging numerical  
198 frameworks showing that explicit representation of microbes as controlling variables can  
199 dramatically change expected effects of climate on broad-scale decomposition dynamics<sup>6,8,32</sup>.

200 We found positive but relatively weak effects of soil N availability on decomposition  
201 (Supplementary Fig. 1), despite the fact stoichiometry is considered a key control on microbial  
202 growth efficiencies and hence biogeochemical process rates<sup>33-36</sup>. The effects might have been  
203 stronger had the litter been of lower quality (e.g. <1% initial N), requiring microbes to source N  
204 from the environment for growth and enzyme production<sup>35</sup>. Such possibilities emphasize the fact  
205 that the effect sizes we report are specific to the spatial and temporal scale of our study. For  
206 example, the relative effect size of controls changes with how progressed litter decay is<sup>37-39</sup>.  
207 Future work will need to test whether the hierarchical model can approximate controls on later  
208 decomposition stages, in other biomes and at even broader spatial scales<sup>37,38</sup>, when challenged  
209 with microsite data. Where the model cannot approximate controls (i.e. where broad-scale  
210 emergent patterns do not reflect microsite relationships), new microsite-level studies will be  
211 needed to re-estimate parameter values for important controls. Such studies should test whether  
212 measuring fine-scale temporal as well as spatial variation might also necessitate a re-evaluation  
213 of how decomposition rates are controlled. Notably, our study leaves unresolved how microsite  
214 variation in litter quality might influence the nature of this co-dominant control. Further, it  
215 suggests a need to re-design multi-site litter decomposition studies but does not address the  
216 challenge of making these studies practical given the very large number of observations  
217 apparently required to test when and to what extent emergent broad-scale patterns fail to capture  
218 microsite-level mechanisms<sup>1</sup>.

219 We acknowledge that three aspects of our design may have influenced our findings:  
220 enclosing litter in mesh can alter the microclimate<sup>40</sup>; the litter species do not occur at every site;  
221 and the microsite scale we measured may also be mismatched with the litterbag scale of the  
222 response variable<sup>41</sup>. However, these caveats also apply to the multi-site litter decomposition  
223 experiments that have helped build and reinforce the hierarchical model<sup>12-15</sup>. The important  
224 caveat that we remove from these previous studies is the assumption that aggregate (i.e. site-  
225 mean) data accurately capture the relationships between decomposition and the variables  
226 regulating it that operate at local (microsite) scales. Notably, there is growing evidence that C-  
227 and N-cycling processes in soil are driven to a large extent by microsite variation in controlling  
228 variables across landscape to regional scales<sup>42-44</sup>. Those working in population and community  
229 ecology have wrestled with the insight that aggregate data may not represent local behaviour and  
230 hence lead to false conclusions and projections<sup>45</sup>; it seems the same insight may need to be  
231 grappled with in ecosystem ecology.

232

### 233 **Conclusions**

234 Scaling theory in ecology describes how emergent patterns can arise from distinct and causative  
235 relationships operating at finer-scales<sup>10</sup>. However, the issue is nested within a broader inferential  
236 challenge traditionally debated in the social sciences and increasingly so in the natural  
237 sciences<sup>42,46-48</sup>. Although apparently named without reference to the field of ecology, the issue is  
238 termed “ecological inference” and refers to the process of using aggregate data to draw  
239 conclusions about individual-level behaviour<sup>48</sup>. Causative relationships inferred from aggregate  
240 data often fail to represent the variables that control how individuals respond to and act on the  
241 environment<sup>49</sup>. By comparison, relationships inferred from site-mean data in regional- to global-

242 scale litter decomposition experiments may operate locally, or instead emerge from a set of  
243 distinct local-scale relationships and controlling variables. We have referred to these two  
244 possibilities as the assumption of scale invariance versus scale dependence (Fig. 2). Although we  
245 find temperature control scale invariant, our findings for moisture and microbial biomass control  
246 suggest that the hierarchical model may be the product of a logical inference fallacy. That is, it  
247 arises because aggregate data are falsely assumed to represent finer-scale causative  
248 relationships<sup>42,48,49</sup>. Encouragingly, the rich body of work on scaling theory and the ecological  
249 inference fallacy<sup>50</sup> provides a platform for ecosystem ecology to test and potentially reformulate  
250 its conceptual and numerical models used to explain and predict how biogeochemical processes  
251 respond to a changing environment. Our findings help reinforce calls to test and reconsider  
252 which environmental variables predominantly regulate biogeochemical process rates at regional-  
253 to global-scales, and when doing so emphasize the need to work at the microsite scales at which  
254 organisms perceive the environment.

255

## 256 **Methods**

257 **Experimental design. *Site layout.*** Our research was conducted in grasslands spanning ~20°  
258 latitude in Western Europe (Fig. 1). At each of six study sites, we established four 30-m linear  
259 transects between 50 m and up to 2 km apart. Transects were chosen to capture within-site  
260 heterogeneity in microclimate and land-use intensity (e.g. with or without grazing). Along each  
261 transect we established 20×20 cm quadrats at 5-m intervals, resulting in 7 quadrats per transect.  
262 In the context of this study, ‘quadrat’ serves as the ‘microsite scale’. Between 28 April and 16  
263 May 2015, we placed two nylon mesh bags (5×10 cm; mesh size 0.9×1 mm) at each quadrat, ~10  
264 cm apart. The mesh size presumably minimized the effect of larger soil fauna (e.g. earthworms)

265 on decomposition rates, and so our decomposition rates were likely primarily the product of  
266 microbes and micro- and mesofauna<sup>24,40</sup>. Each mesh bag contained 1 g air-dried grass foliar litter  
267 of either *Holcus lanatus* L. or *Festuca rubra* L., which differ in their litter chemical properties  
268 (see below). This resulted in a total of 6 locations × 4 transects × 7 quadrats × 2 litter types = 336  
269 litterbags. Litterbags were placed flush with the soil surface, within the existing litter layer and  
270 were retrieved after ~3 months. Of the 336 bags placed, 32 were lost in the field to such events  
271 as consumption by cows and accidental site mowing. The litter used to fill the litterbags was  
272 collected as freshly senesced material in grasslands local to the Dutch site.

273  
274 *Leaf litter.* Mean litter properties for *H. lanatus* versus *F. rubra* were pH of 6.12 vs. 5.61, %N of  
275 1.78 vs. 1.03, C:N of 24.7 vs. 43.7, and lignin, calcium, magnesium and potassium contents of  
276 157 vs. 175, 3.72 vs. 2.75, 1.31 vs. 0.79, and 6.55 vs. 1.50 mg g<sup>-1</sup>, respectively. That is,  
277 regardless of the chemical property measured, *H. lanatus* was always less recalcitrant. By  
278 including the two contrasting litter types at every site, we generated equal within and among site  
279 variation in this variable. Doing so provided a statistical control whereby the strong within-site  
280 litter type effect should be approximated by the among site effect, and so generate a scale  
281 invariant pattern (Fig. 2a). Second, standardizing known controlling variables can improve  
282 estimated effects of the controls under study (e.g. microclimate). Third, litter traits are expected  
283 to interact with other variables, such as temperature<sup>23</sup>, and so including this variable allowed us  
284 to test this possibility.

285  
286 **Measurements.** *Field.* At each quadrat we determined microclimate at the start, after ~6 weeks  
287 and at the end of the field incubation period. We collected three measures per quadrat and time

288 point of soil temperature at 5-cm depth using a hand-held thermometer. Such repeated spot  
289 measurements are effective at characterizing relative variation in microclimate<sup>42</sup>, and so our  
290 measures are not indicative of absolute values experienced by the decomposing litters but instead  
291 capture generally warmer vs. cooler microsites, or drier vs. wetter, across the course of the study.  
292 At the mid and end time point, soil moisture content was determined gravimetrically in three soil  
293 cores (5 cm depth, 2 cm diam.) from each quadrat; cores were pooled and dried at 105°C until  
294 constant mass. We had intended to use these measures (plus initial soil moisture) to estimate  
295 microsite moisture conditions, but marked differences in soil texture from clay (Umeå) to loamy  
296 sand (Wageningen) meant that soil gravimetric moisture was a poor surrogate for litter layer  
297 moisture conditions. Instead, we used litter moisture values (see *Testing Prediction 1* below).  
298 Additionally, at the start point of the field incubations, 8-10 soil cores of the same size were  
299 taken and pooled per quadrat and were used to determine soil gravimetric moisture, microbial  
300 biomass and N availability. Initial soil samples and retrieved litterbags were shipped to the  
301 Netherlands Institute of Ecology to ensure common processing. Collectively these measures  
302 were intended to give estimates of four variables identified as important controls of  
303 decomposition either at broad-scales (i.e. temperature and moisture), or at local-scales (i.e.  
304 microbial biomass and N availability)<sup>32,35,51,52</sup>. For soil microbial biomass, it is probably fairer to  
305 consider this an estimate of the spatial variation in soil community activity, which includes  
306 invertebrate decomposers, many of which will have been able to access the litter<sup>24,40</sup>, and  
307 potentially also microbes not involved in litter decomposition.

308

309 *Laboratory*. Retrieved litter was cleaned of roots, fauna and soil, before mass was determined  
310 fresh and after drying at 65°C. It was next milled to a fine powder and analysed for total C  
311 content through elemental analysis (Flash 2000, Thermo Fisher Scientific, Bremen, Germany).

312 The initial 168 soils (6 locations × 4 transects × 7 quadrats) were passed through a 4-mm  
313 sieve and sub-sampled for gravimetric moisture, microbial biomass and N availability. We used  
314 the substrate-induced respiration (SIR) method to estimate active microbial biomass<sup>53</sup>, modified  
315 per Fierer *et al.*<sup>54</sup>. We estimated soil N availability by determining potential net N mineralization  
316 rates as the difference between salt-extractable N-NO<sub>3</sub><sup>-</sup> and N-NH<sub>4</sub><sup>+</sup> at time zero and after 14 d of  
317 incubation at 20°C and 65% water holding capacity<sup>55</sup>. Soils were extracted with 1M KCl and  
318 extracts measured using an auto-analyser (QuAAtro Segmented Flow Analyser; SEAL  
319 Analytical; Norderstedt, Germany).

320 Initial litter properties were estimated using seven randomly collected samples per  
321 species, matching the sub-sampling for the litterbags. Total C and N content were measured as  
322 described above, lignin after a chloroform/methanol extraction and hydrolysis with HCl,  
323 following Poorter & Villar<sup>56</sup>. Mineral nutrient concentrations and pH were measured following  
324 methods described in Hendry and Grime<sup>57</sup> and Cornelissen *et al.*<sup>58</sup>, respectively.

325

326 **Data and inferential analysis.** *Overview of approach.* We built a set of regression models,  
327 structured to represent and test between assumptions of scale invariance versus dependence in  
328 controls on litter decomposition (Fig. 2), to compare estimated effect sizes on decomposition of  
329 the four controlling variables under study. Specifically, we estimated the relative effect size for  
330 temperature, moisture, soil N availability and microbial biomass, across the range of observed  
331 values within and among our six sites. The relative effect size depends on the slope coefficient



332 for the specific variable, the slope coefficient for any interaction it is involved in, and the range  
333 of observed values of the variable. We generated the coefficients by fitting linear mixed-effect  
334 models (LMMs). The effect size of a variable on mass C loss was estimated using these  
335 regression parameters, while holding all other variables constant (i.e. the mean of all  
336 observations for each variable), and systematically varying the variable of interest across its  
337 measured range of values. That is, we plotted the regression equation for a model using the  
338 coefficients from the respective LMM, the mean value across all 168 quadrats for the controls  
339 not under test, and then for the control under test we estimated decomposition rates by  
340 systematically increasing the value of the control from the lowest to highest observed values  
341 across the 168 quadrats.

342         The choice of variables to measure and then include in our statistical models (described  
343 next) was based on the approach of Hobbs *et al.*<sup>59</sup>, which rejects model selection on  
344 philosophical and operational grounds. Philosophically, we investigated only variables where  
345 biological mechanism as to their influence on decomposition is firmly established. Operationally,  
346 there is subjectivity and lack of agreement in statistical model selection approaches, with  
347 different decisions leading to markedly different conclusions as to effect sizes. Instead,  
348 coefficients and hence effect sizes are generally most robust when all terms are retained,  
349 assuming that each is included with well-established biological foundation.

350

351 *Testing Prediction 1.* Prediction 1 was that relationships between climate and decomposition  
352 rates should differ when site-mean versus microsite-level climate data are analysed. This  
353 prediction was evaluated by comparing whether temperature and moisture effects on mass C loss  
354 differed when the slope coefficients were estimated from microsite versus site-mean data. We

355 established a single model structure to test Prediction 1. It included only recognized broad-scale  
356 controls as variables (i.e. temperature, moisture and litter type), but involved different data  
357 aggregation. The “Microclimate” model was tested with observations of mass C loss for each  
358 litterbag and quadrat-level microclimate. The “Site-mean climate” model was also run with all  
359 litterbag observations – to minimize changes in predictive power associated with changing  
360 values of  $n$  – but the values of the climate variables were the mean per site of the microclimate  
361 (i.e. quadrat) observations. Hence in the Microclimate model the dataset had 168 unique  
362 temperature and moisture observations, whereas in the Site-mean climate model there were only  
363 six possible values (one per site) of temperature and moisture. Specifically, microsite control  
364 values were determined from the quadrat-level measures, and site mean values determined from  
365 the mean of the 28 quadrat-measures within a site (i.e. they were based on the exact same set of  
366 measurements). To account for potential spatial auto-correlation among the quadrats within a  
367 site, we fit a random error structure accounting for the spatial hierarchy in the design (quadrat  
368 nested within transect, with transect nested within site), assuming a common slope but spatially-  
369 dependent intercept<sup>50,60</sup>.

370         Similarly, litter type was included as the litterbag-level %N value, or as the mean %N per  
371 litter type, respectively (note that climate effect sizes were independent of how litter type was  
372 included). To determine a litterbag-level initial %N value, we randomly assigned to each  
373 litterbag a %N value (to the nearest 0.1%) drawn from the measured range of initial %N values  
374 from seven additional litterbag samples (Fig. 3b). We did this to acknowledge that there was  
375 variation among litterbags in initial %N and so using the mean initial %N would give a false  
376 account of the among-bag variation. For quadrat-level temperature, we calculated the mean soil  
377 temperature across the three field measurement periods. For quadrat-level moisture, given that

378 soil gravimetric moisture was not useful given soil texture differences among sites, we calculated  
379 quadrat-level moisture as the mean of the *Holcus* and *Festuca* litterbag moisture values on  
380 collection. We acknowledge that litters were probably drier at collection than at earlier points of  
381 the field incubations, given increasing temperatures and declining precipitation across the  
382 incubations, and so these values provide an estimate of relative spatial differences in moisture  
383 only. We used the mean across the two litter types, given that species-specific moisture values  
384 are often a product of leaf litter traits and are thus correlated with litter quality<sup>4</sup>.

385

386 *Testing Prediction 2.* Prediction 2 was that any variable expected to be an important control at  
387 the microsite-level, should have a strong effect when regional-scale patterns are analysed using  
388 microsite data. Specifically, we evaluated whether effect sizes of the soil microbial biomass and  
389 N availability variables had effect sizes comparable to recognized broad-scale controls  
390 (specifically temperature and moisture). We developed three model structures. The “Microsite  
391 interactions” model included all variables (i.e. temperature, moisture, microbial biomass, N  
392 availability) and their 2-way interactions. We included two-way interactions among the main  
393 effects given expectations that the relative effects of our variables should depend on one another.  
394 For example, the decomposition rate of more recalcitrant litters is expected to be more  
395 temperature sensitive<sup>61,62</sup>. The “Microsite main effects” model removed the 2-way interactions to  
396 determine whether the effect sizes of the variables were primarily additive. The “Microclimate”  
397 model was used again but to evaluate whether dropping the soil microbial biomass and N  
398 availability terms altered inferences about temperature and moisture controls on mass C loss.  
399 Litter type (as initial %N) was again included in all models.

400

401 *Statistical model specifics.* The LMMs were fit with a Gaussian error distribution in the “lme4”  
402 package for the “R” statistical program (version 3.1.3), using the “lmer” function.  
403 Decomposition was calculated as the proportional mass C loss from the litterbags. Site, transect  
404 and quadrat were fit as random variables to the LMMs, with the finer scale variables nested  
405 within the broader scale variables, given the potential for autocorrelation caused by spatially  
406 clustering the litterbags<sup>60</sup>. Before we tested the model structures described above, we tested the  
407 data distributions. A single and highly influential observation (based on Cook’s *D*) was dropped  
408 from the dataset; it had a mass C loss value of 69.9%, far higher than any other observation (Fig.  
409 3a), and markedly affected residual fits. The remaining data conformed to assumptions of  
410 normality, and a second-order temperature term was included given the observed unimodal  
411 relationship between temperature and mass loss. Also, initial extractable N was a better choice  
412 (i.e. higher standardized coefficient) than potential N mineralization for soil N availability, and  
413 litter moisture (mean per quadrat) performed better than gravimetric soil moisture. Litter initial  
414 %N was used to represent litter quality given that it is a strong predictor of early-stage  
415 decomposition in grasses such as *H. lanatus*<sup>39,63</sup>.

416 The square-root variance inflation factors (vif) were <2 for the main effects, indicating  
417 low collinearity. As would be expected, there was a strong correlation between temperature and  
418 its second-order term, and where the effect of one variable strongly interacted with another. We  
419 reduced these ‘vif’ values by standardizing the observed value of each variable by subtracting the  
420 mean and dividing by two standard deviations<sup>64</sup>. The resulting standardized coefficients also  
421 permit coefficients to be directly compared for variables measured on different unit scales.  
422 Confirming the validity of our inferences in spite of introduced collinearity when second-order  
423 terms and interactions were permitted, variables with large effect sizes calculated on the basis of

424 the unstandardized coefficients also had large standardized coefficients. In addition, in the  
425 ‘Microsite main effects’ model all 2-way interactions were dropped, removing collinearity and  
426 concerns about over-fitting, and the relative magnitude of the coefficients were largely  
427 unchanged (Table 1).

428 All reported *P*-values are quasi-Bayesian but retain the same interpretation as frequentist  
429 *P*-values<sup>65</sup>. We considered coefficients with *P*<0.05 to be significant and coefficients with  
430 *P*<0.10 marginally significant. We calculated the  $r^2$  values for each model following Nakagawa  
431 and Schielzeth<sup>66</sup>. Calculation of  $r^2$  values is common practice when modelling decomposition  
432 and a high value associated with a specific explanatory variable is often associated with that  
433 variable having a strong effect size. This reasoning makes no sense within the context of our  
434 study because litter type was experimentally controlled and accurately measured within and  
435 among sites, whereas the other variables relied on observed variation and measurements that  
436 represented – but likely did not fully characterize – the conditions that acted on decomposer  
437 activity. The latter conditions make data more “noisy”, lowering  $r^2$  values, but in the absence of  
438 systematic bias will not change the coefficient estimates and hence effect sizes<sup>1</sup>. We therefore  
439 only report the  $r^2$  value for each model, to verify they had the potential to explain a substantive  
440 degree of the variance in decomposition rate.

441

#### 442 **Data availability**

443 Experimental data in the support of these findings and the R code for the statistical models are  
444 available via the Dryad Digital Repository (<http://dx.doi.org/10.5061/dryad.c44h0>).

445

#### 446 **References**

447 1 Bradford, M. A., Berg, B., Maynard, D. S., Wieder, W. R. & Wood, S. A. Understanding  
448 the dominant controls on litter decomposition. *J. Ecol.* **104**, 229-238 (2016).

449 2 Cornwell, W. K. *et al.* Plant species traits are the predominant control on litter  
450 decomposition rates within biomes worldwide. *Ecol. Lett.* **11**, 1065-1071 (2008).

451 3 Freschet, G. T., Aerts, R. & Cornelissen, J. H. C. A plant economics spectrum of litter  
452 decomposability. *Func. Ecol.* **26**, 56-65 (2012).

453 4 Makkonen, M. *et al.* Highly consistent effects of plant litter identity and functional traits  
454 on decomposition across a latitudinal gradient. *Ecol. Lett.* **15**, 1033-1041 (2012).

455 5 Swift, M. J., Heal, O. W. & Anderson, J. M. *Decomposition in terrestrial ecosystems.*  
456 *Studies in Ecology volume 5.* (Blackwell Scientific, 1979).

457 6 Buchkowski, R. W., Bradford, M. A., Grandy, A. S., Schmitz, O. J. & Wieder, W. R.  
458 Applying population and community ecology theory to advance understanding of  
459 belowground biogeochemistry. *Ecol. Lett.* **20**, 231-245 (2017).

460 7 Sulman, B. N., Phillips, R. P., Oishi, A. C., Shevliakova, E. & Pacala, S. W. Microbe-  
461 driven turnover offsets mineral-mediated storage of soil carbon under elevated CO<sub>2</sub>. *Nat.*  
462 *Clim. Change* **4**, 1099–1102 (2014).

463 8 Tang, J. & Riley, W. J. Weaker soil carbon–climate feedbacks resulting from microbial  
464 and abiotic interactions. *Nat. Clim. Change* **5**, 56-60 (2014).

465 9 Wieder, W. R., Bonan, G. B. & Allison, S. D. Global soil carbon projections are  
466 improved by modelling microbial processes. *Nat. Clim. Change* **3**, 909-912 (2013).

467 10 Levin, S. A. The problem of pattern and scale in ecology. *Ecology* **73**, 1943-1967 (1992).

468 11 Lauenroth, W. K. & Sala, O. E. Long-term forage production of North American  
469 shortgrass steppe. *Ecol. App.* **2**, 397-403 (1992).

470 12 Berg, B. *et al.* Litter mass-loss rates in pine forests for Europe and Eastern United States:  
471 some relationships with climate and litter quality. *Biogeochem.* **20**, 127-159 (1993).

472 13 Harmon, M. E. *et al.* Long-term patterns of mass loss during the decomposition of leaf  
473 and fine root litter: an intersite comparison. *Glob. Change Biol.* **15**, 1320-1338 (2009).

474 14 Moore, T. R. *et al.* Litter decomposition rates in Canadian forests. *Glob. Change Biol.* **5**,  
475 75-82 (1999).

476 15 Wall, D. H. *et al.* Global decomposition experiment shows soil animal impacts on  
477 decomposition are climate-dependent. *Glob. Change Biol.* **14**, 2661-2677 (2008).

478 16 Bonan, G. B., Hartman, M. D., Parton, W. J. & Wieder, W. R. Evaluating litter  
479 decomposition in earth system models with long-term litterbag experiments: an example  
480 using the Community Land Model version 4 (CLM4). *Glob. Change Biol.* **19**, 957-974  
481 (2013).

482 17 Averill, C., Waring, B. G. & Hawkes, C. V. Historical precipitation predictably alters the  
483 shape and magnitude of microbial functional response to soil moisture. *Glob. Change*  
484 *Biol.* **5**, 1957-1964 (2016).

485 18 Strickland, M. S., Keiser, A. D. & Bradford, M. A. Climate history shapes contemporary  
486 leaf litter decomposition. *Biogeochem.* **122**, 165-174 (2015).

487 19 Fierer, N. *et al.* Cross-biome metagenomic analyses of soil microbial communities and  
488 their functional attributes. *P. Natl. Acad. Sci. USA* **109**, 21390-21395 (2012).

489 20 Evans, S. E. & Wallenstein, M. D. Climate change alters ecological strategies of soil  
490 bacteria. *Ecol. Lett.* **17**, 155-164 (2014).

- 491 21 Loescher, H., Ayres, E., Duffy, P., Luo, H. & Brunke, M. Spatial variation in soil  
492 properties among North American ecosystems and guidelines for sampling designs. *PLoS*  
493 *One* **9**, e83216 (2014).
- 494 22 Scherrer, D. & Körner, C. Infra-red thermometry of alpine landscapes challenges climatic  
495 warming projections. *Glob. Change Biol.* **16**, 2602-2613 (2010).
- 496 23 Meentemeyer, V. Macroclimate and lignin control of litter decomposition rates. *Ecology*  
497 **59**, 465-472 (1978).
- 498 24 García-Palacios, P., Maestre, F. T., Kattge, J. & Wall, D. H. Climate and litter quality  
499 differently modulate the effects of soil fauna on litter decomposition across biomes. *Ecol.*  
500 *Lett.* **16**, 1045-1053 (2013).
- 501 25 Tenney, F. G. & Waksman, S. A. Composition of natural organic materials and their  
502 decomposition in the soil: IV. The nature and rapidity of decomposition of the various  
503 organic complexes in different plant materials, under aerobic conditions. *Soil Science* **28**,  
504 55-84 (1929).
- 505 26 Handa, I. T. *et al.* Consequences of biodiversity loss for litter decomposition across  
506 biomes. *Nature* **509**, 218-221 (2014).
- 507 27 Powers, J. S. *et al.* Decomposition in tropical forests: a pan-tropical study of the effects  
508 of litter type, litter placement and mesofaunal exclusion across a precipitation gradient. *J.*  
509 *Ecol.* **97**, 801-811 (2009).
- 510 28 Crowther, T. W. *et al.* Biotic interactions mediate soil microbial feedbacks to climate  
511 change. *P. Natl. Acad. Sci. USA* **112**, 7033–7038 (2015).



512 29 Lawrence, C. R., Neff, J. C. & Schimel, J. P. Does adding microbial mechanisms of  
513 decomposition improve soil organic matter models? A comparison of four models using  
514 data from a pulsed rewetting experiment. *Soil Biol. Biochem.* **41**, 1923-1934 (2009).

515 30 Hall, E. *et al.* Understanding how microbiomes influence the systems they inhabit:  
516 Insight from ecosystem ecology. *bioRxiv* <http://dx.doi.org/10.1101/065128> (2016).

517 31 Aerts, R. Climate, leaf litter chemistry and leaf litter decomposition in terrestrial  
518 ecosystems: a triangular relationship. *Oikos* **79**, 439-449 (1997).

519 32 Allison, S. D., Wallenstein, M. D. & Bradford, M. A. Soil-carbon response to warming  
520 dependent on microbial physiology. *Nat. Geosci.* **3**, 336-340 (2010).

521 33 Crowther, T. W. *et al.* Environmental stress response limits microbial necromass  
522 contributions to soil organic carbon. *Soil. Biol. Biochem.* **85**, 153-161 (2015).

523 34 Frey, S. D., Lee, J., Melillo, J. M. & Six, J. The temperature response of soil microbial  
524 efficiency and its feedback to climate. *Nat. Clim. Change* **3**, 395-398 (2013).

525 35 Schimel, J. P. & Weintraub, M. N. The implications of exoenzyme activity on microbial  
526 carbon and nitrogen limitation in soil: a theoretical model. *Soil Biol. Biochem.* **35**, 549-  
527 563 (2003).

528 36 Buchkowski, R. W., Schmitz, O. J. & Bradford, M. A. Microbial stoichiometry overrides  
529 biomass as a regulator of soil carbon and nitrogen cycling. *Ecology* **96**, 1139-1149  
530 (2015).

531 37 Adair, E. C. *et al.* Simple three-pool model accurately describes patterns of long-term  
532 litter decomposition in diverse climates. *Glob. Change Biol.* **14**, 2636-2660 (2008).

533 38 Currie, W. S. *et al.* Cross-biome transplants of plant litter show decomposition models  
534 extend to a broader climatic range but lose predictability at the decadal time scale. *Glob.*  
535 *Change Biol.* **16**, 1744-1761 (2010).

536 39 Smith, V. C. & Bradford, M. A. Litter quality impacts on grassland litter decomposition  
537 are differently dependent on soil fauna across time. *Appl. Soil Ecol.* **24**, 197-203 (2003).

538 40 Bradford, M. A., Tordoff, G. M., Eggers, T., Jones, T. H. & Newington, J. E. Microbiota,  
539 fauna, and mesh size interactions in litter decomposition. *Oikos* **99**, 317-323 (2002).

540 41 Bokhorst, S. & Wardle, D. A. Microclimate within litter bags of different mesh size:  
541 Implications for the 'arthropod effect' on litter decomposition. *Soil Biol. Biochem.* **58**,  
542 147-152 (2013).

543 42 Bradford, M. A. *et al.* Climate fails to predict wood decomposition at regional scales.  
544 *Nat. Clim. Change* **4**, 625-630 (2014).

545 43 Keiser, A. D., Knoepp, J. D. & Bradford, M. A. Disturbance decouples biogeochemical  
546 cycles across forests of the southeastern US. *Ecosystems* **19**, 50-61 (2016).

547 44 Waring, B., Adams, R., Branco, S. & Powers, J. S. Scale-dependent variation in nitrogen  
548 cycling and soil fungal communities along gradients of forest composition and age in  
549 regenerating tropical dry forests. *New Phyt.* **209**, 845-854 (2016).

550 45 Schmitz, O. J. *Resolving ecosystem complexity.* (Princeton Univ. Press, 2010).

551 46 Oakes, M. J. Commentary: individual, ecological and multilevel fallacies. *Inter. J.*  
552 *Epidem.* **38**, 361-368 (2009).

553 47 Robinson, W. S. Ecological correlations and the behavior of individuals. *Am. Socio. Rev.*  
554 **15**, 351-357 (1950).

555 48 Schuessler, A. A. Ecological inference. *P. Natl. Acad. Sci. USA* **96**, 10578-10581 (1999).

556 49 Gelman, A., Shor, B., Bafumi, J. & Park, D. Rich state, poor state, red state, blue state:  
557 what's the matter with Connecticut? *Qu. J. Poli. Sci.* **2**, 345-367 (2007).

558 50 Gelman, A. & Hill, J. *Data analysis using regression and multilevel/hierarchical models*.  
559 (Cambridge Univ. Press, 2007).

560 51 Rousk, J. Biomass or growth? How to measure soil food webs to understand structure and  
561 function. *Soil Biol. Biochem.* **102**, 45-47 (2016).

562 52 Allison, S. D. *et al.* Microbial abundance and composition influence litter decomposition  
563 response to environmental change. *Ecology* **94**, 714-725 (2013).

564 53 Anderson, J. P. E. & Domsch, K. H. A physiological method for the quantitative  
565 measurement of microbial biomass in soils. *Soil Biol. Biochem.* **10**, 215-221 (1978).

566 54 Fierer, N., Schimel, J. P. & Holden, P. A. Influence of drying-rewetting frequency on soil  
567 bacterial community structure. *Microb. Ecol.* **45**, 63-71 (2003).

568 55 Robertson, G. P. *et al.* in *Standard soil methods for long-term ecological research* (eds G  
569 P Robertson, D C Coleman, C S Bledsoe, & P Sollins) 258-271 (Oxford University Press,  
570 1999).

571 56 Poorter, H. & Villar, R. in *Plant resource allocation* (eds F A Bazzaz & J Grace) 39-72  
572 (Academic Press, 1997).

573 57 Hendry, G. A. F. & Grime, J. P. *Methods in comparative plant ecology* (Chapman &  
574 Hall, 1993).

575 58 Cornelissen, J. H. C. *et al.* Foliar pH as a new plant trait: can it explain variation in foliar  
576 chemistry and carbon cycling processes among subarctic plant species and types?  
577 *Oecologia* **147**, 315-326 (2006).

578 59 Hobbs, N. T., Andren, H., Persson, J., Aronsson, M. & Chapron, G. Native predators  
579 reduce harvest of reindeer by Sámi pastoralists. *Ecol. App.* **22**, 1640-1654 (2012).

580 60 Bolker, B. M. *et al.* Generalized linear mixed models: a practical guide for ecology and  
581 evolution. *Trends Ecol. Evol.* **24**, 127-135 (2009).

582 61 Fierer, N., Craine, J. M., McLauchlan, K. & Schimel, J. P. Litter quality and the  
583 temperature sensitivity of decomposition. *Ecology* **86**, 320-326 (2005).

584 62 Conant, R. T. *et al.* Temperature and soil organic matter decomposition rates – synthesis  
585 of current knowledge and a way forward. *Glob. Change Biol.* **17**, 3392-3404 (2011).

586 63 Smith, V. C. & Bradford, M. A. Do non-additive effects on decomposition in litter-mix  
587 experiments result from differences in resource quality between litters? *Oikos* **102**, 235-  
588 242 (2003).

589 64 Gelman, A. Scaling regression inputs by dividing by two standard deviations. *Stat. in*  
590 *Med.* **27**, 2865-2873 (2008).

591 65 Baayen, R. H., Davidson, D. J. & Bates, D. M. Mixed-effects modeling with crossed  
592 random effects for subjects and items. *J. Mem. Lang.* **59**, 390-412 (2008).

593 66 Nakagawa, S. & Schielzeth, H. A general and simple method for obtaining R<sup>2</sup> from  
594 generalized linear mixed-effects models. *Meth. Ecol. Evol.* **4**, 133-142 (2013).

595

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605

## 606 **Authorship**

607 MAB and GFV contributed equally to this work. They designed the study, co-wrote the  
608 manuscript, constructed litterbags and carried out the lab analyses. All authors established,  
609 maintained and collected data from the field sites. MAB, GFV, DSM and SAW analysed data.  
610 All authors contributed to data interpretation and paper writing.

611

## 612 **Additional information**

613 **Supplementary information** is available for this paper.

614 **Correspondence and requests for materials** should be addressed to M.A.B.

615

## 616 **Competing interests**

617 The authors declare no competing financial interests.

618

619 **Figure 1 | Study design and site characteristics.** Spatial organisation and operational  
620 definitions of the study extent and observational grain are given in the hierarchical figure (site to  
621 microsite). Sites are named for the closest city and their climate data are from climatedata.eu for  
622 the months (May-June) of litterbag incubation, giving the range across months in the average  
623 high and low temperature and precipitation. Soil data are the mean soil temperature and litter

624 moisture data measured across the study period. Latitude and longitude data are for one transect  
625 in each site.

626 **Figure 2 | Competing assumptions for how decomposer communities affect relationships**  
627 **between climate and decomposition rates at regional to global scales.** Ecosystem theory  
628 holds that soil decomposer communities influence functional relationships between controls and  
629 decomposition rates in a spatially invariant manner. For example, broad-scale patterns among  
630 site-mean climate conditions are representative of a common relationship operating at finer  
631 spatial scales (a): the assumption of *scale invariance*. Increasingly there is empirical evidence  
632 that decomposer communities can be functionally distinct, meaning that broad-scale patterns  
633 may instead emerge from distinct fine-scale (in this case within-site) relationships (b): the  
634 assumption of *scale dependence*.

635 **Figure 3 | Measured variation in decomposition rates and controlling variables within and**  
636 **among sites.** The response variable (decomposition) is shown in (a), litter quality in (b), climate  
637 variables in (c) and (d), soil nitrogen availability in (e) and an estimate of the active decomposer  
638 biomass in (f). Points represent individual observations ( $n=303$ ) and are jiggered around the site  
639 number to help prevent similar observations obscuring one another. Sites are described in Fig. 1.

640 **Figure 4 | Estimated effects of temperature and moisture controls on decomposition rates.**  
641 Effect sizes are estimated for temperature (a) and moisture (b) using the coefficients from the  
642 models presented in Table 1. Specifically, these coefficients were used in a regression equation,  
643 along with the mean value across all 168 quadrats for the controls not under test, and then for the  
644 control under test by systematically increasing the control from the lowest to highest observed  
645 values across the 168 quadrats. Comparisons of effect sizes between the Microclimate versus  
646 Site-mean climate models test whether patterns between site-mean climate and decomposition

647 rates (effect sizes from the Site-mean climate model) approximate those operating at the  
648 microsite scales at which decomposer organisms perceive the environment (effect sizes from the  
649 Microclimate model). The temperature-decomposition relationship appears scale invariant  
650 whereas the moisture-decomposition relationship is scale dependent (Fig. 2). The two Microsite  
651 models ask whether inclusion of microbial biomass and N availability as additional variables  
652 alters the estimated effects of temperature and moisture. Their inclusion does not appear to  
653 strongly affect the climate-decomposition relationships.

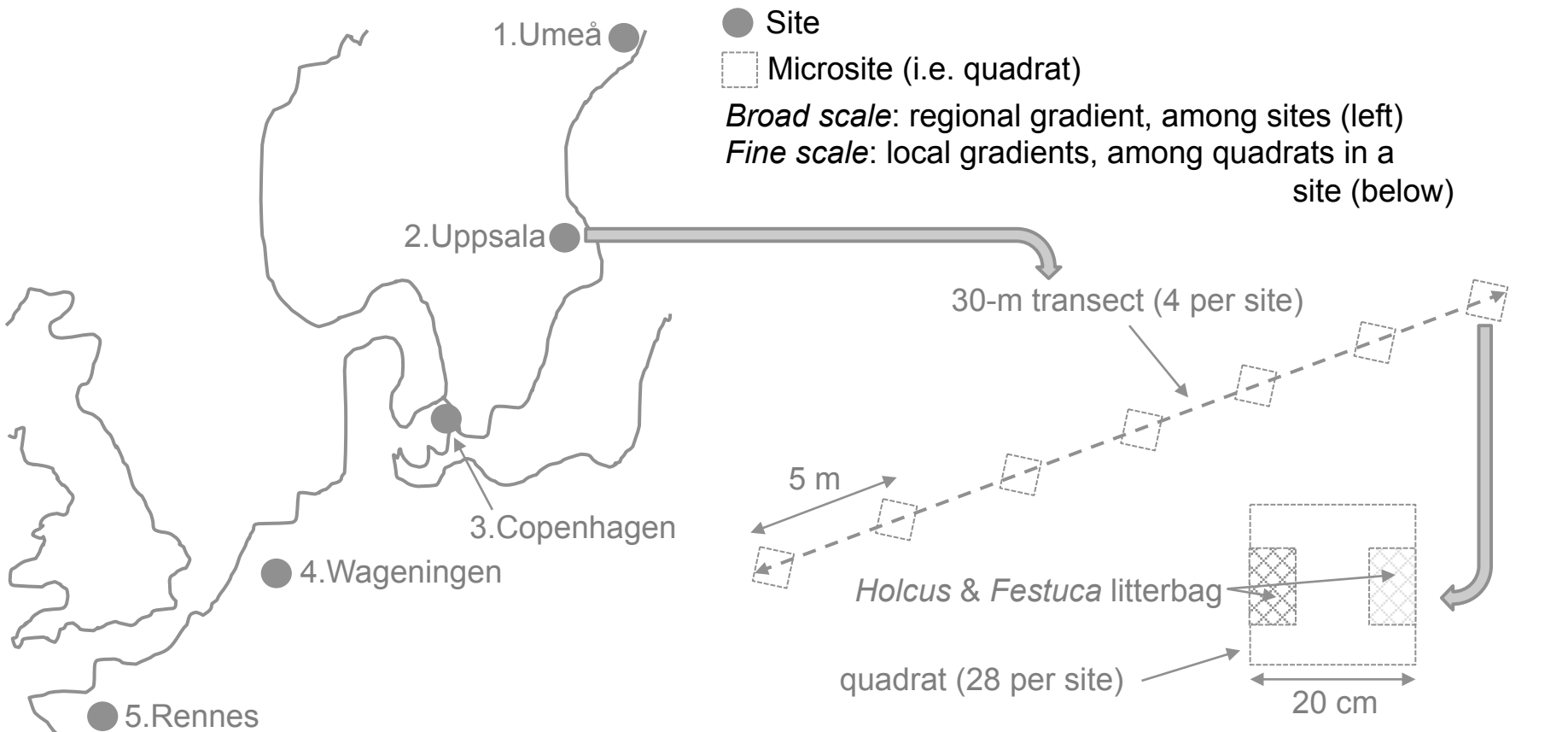
654 **Figure 5 | Estimated effects of controls on decomposition rates.** Effect sizes are estimated  
655 from the Microsite interactions model presented in Table 1, and in (b) also from the Microsite  
656 main effects model, following the procedure described in the legend of Fig. 4. In (a), plots for  
657 each variable are generated using unstandardized coefficients from the “Microsite interactions”  
658 model and the measured range in microsite conditions. The levels of each variable are  
659 relativized, ranging from the minimum (0%) to maximum (100%) measured value, revealing that  
660 microbial biomass (Microbe) has an effect size approximately equivalent to both temperature and  
661 moisture. In (b), comparison of the two models asks whether the effect size of the microbial  
662 biomass is additive or non-additively dependent on the other controlling variables. Its effect  
663 seems primarily additive, given the similarity in the two plots. However, the effect sizes plotted  
664 in (c) and (d) reveal that this additive effect of microbial biomass can still strongly determine  
665 temperature and moisture effects on decomposition rates. The level of microbial biomass is  
666 relativized, with five values shown ranging from the minimum (0%) to maximum (100%)  
667 observed value. There are much stronger absolute decomposition responses to temperature and  
668 moisture when microbial biomass values are greater.

669

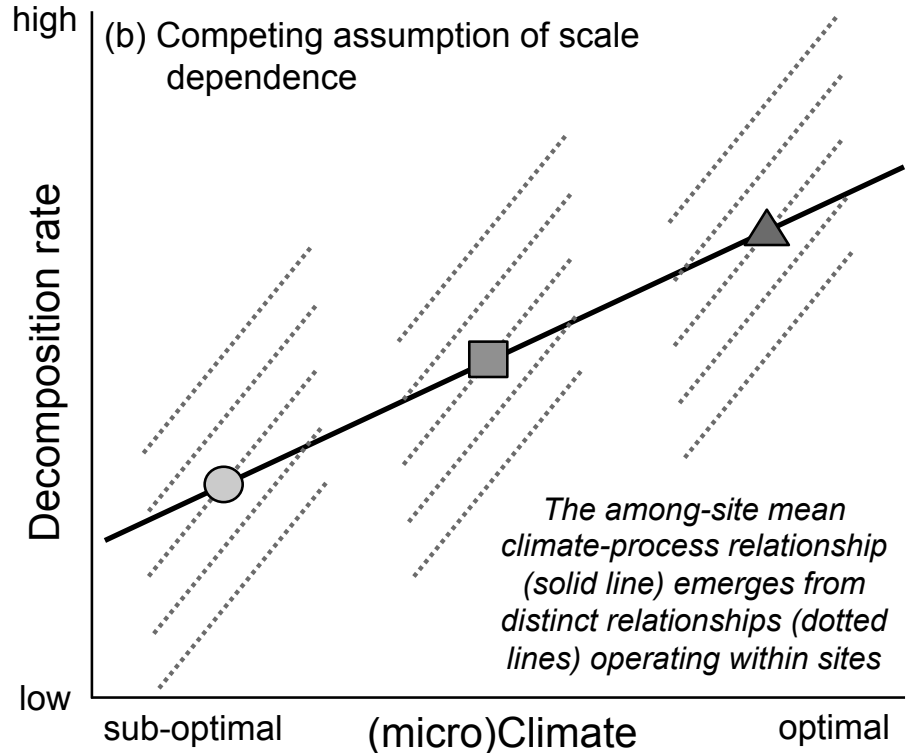
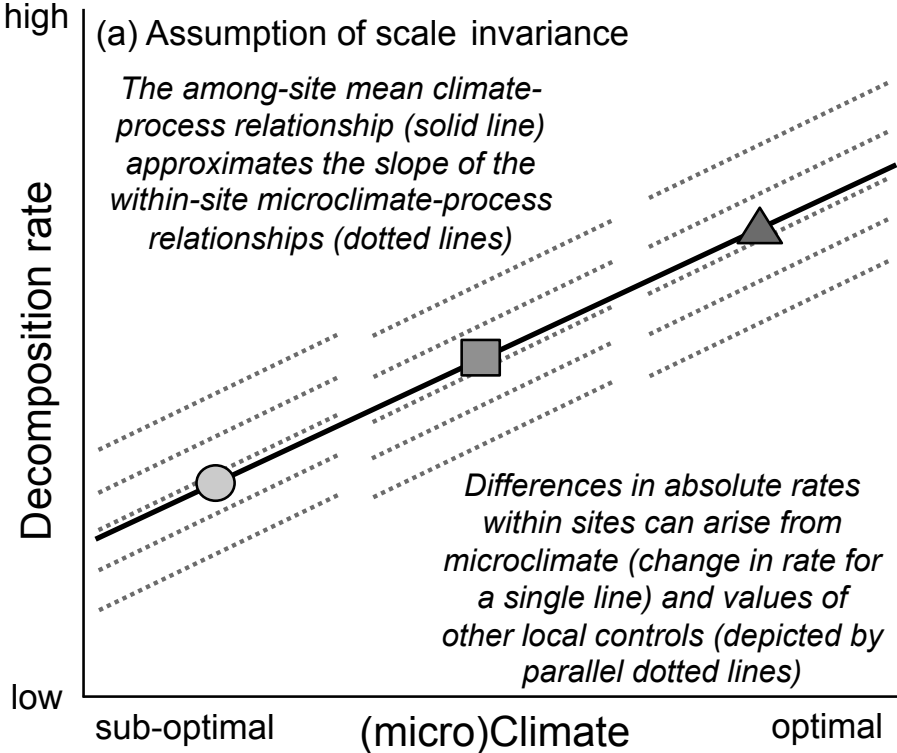
670 **Table 1** | Coefficients, significance and  $r^2$  values for the linear mixed models used to evaluate  
671 controls on litter decomposition rates.

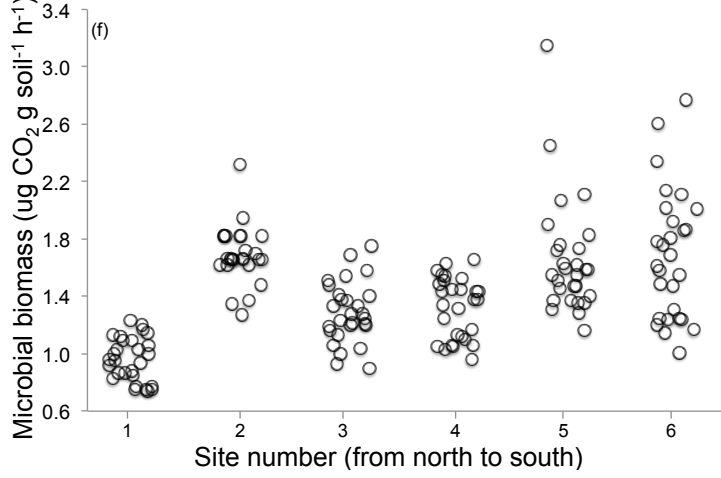
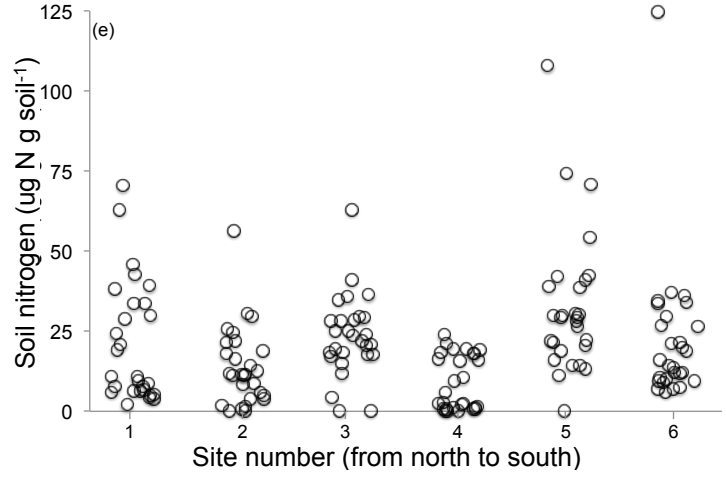
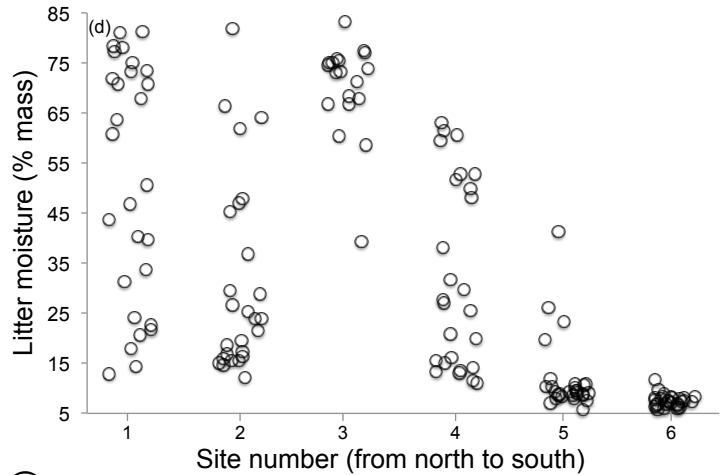
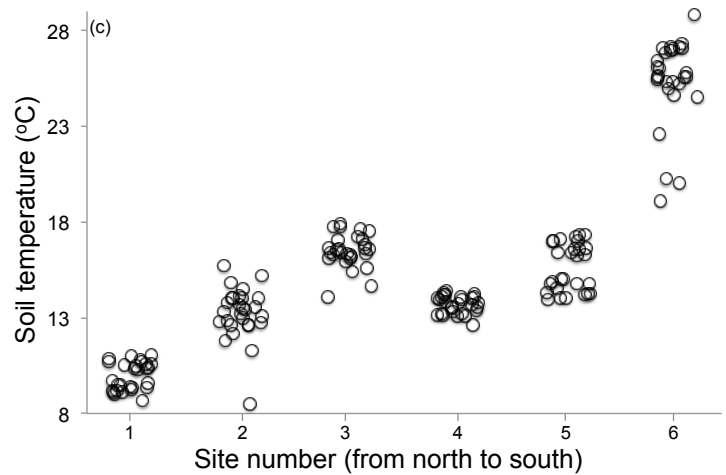
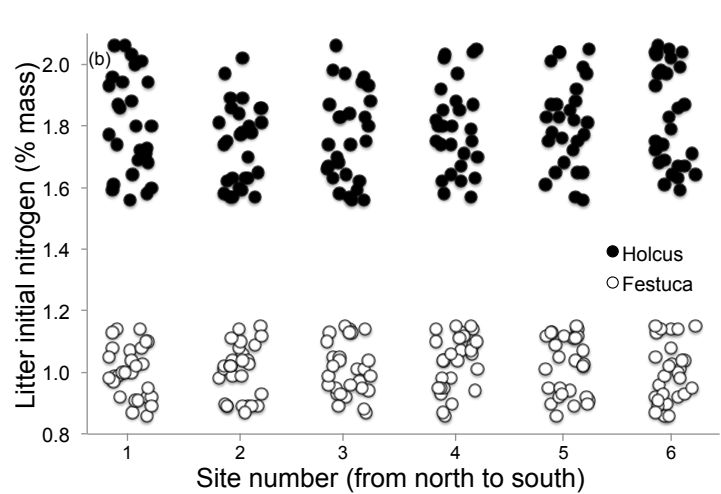
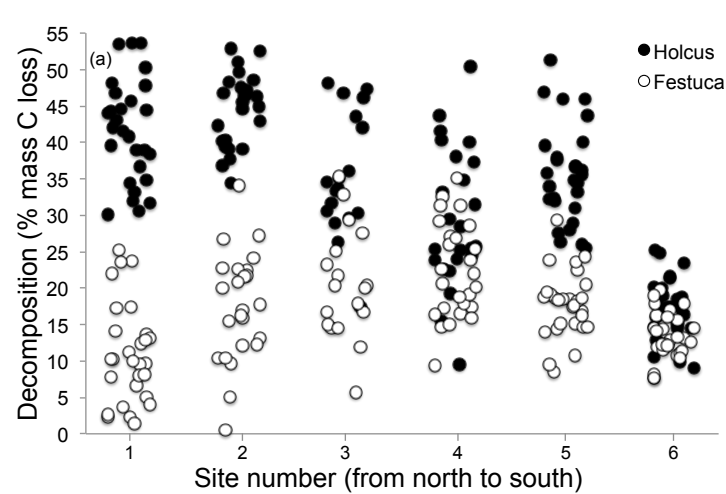


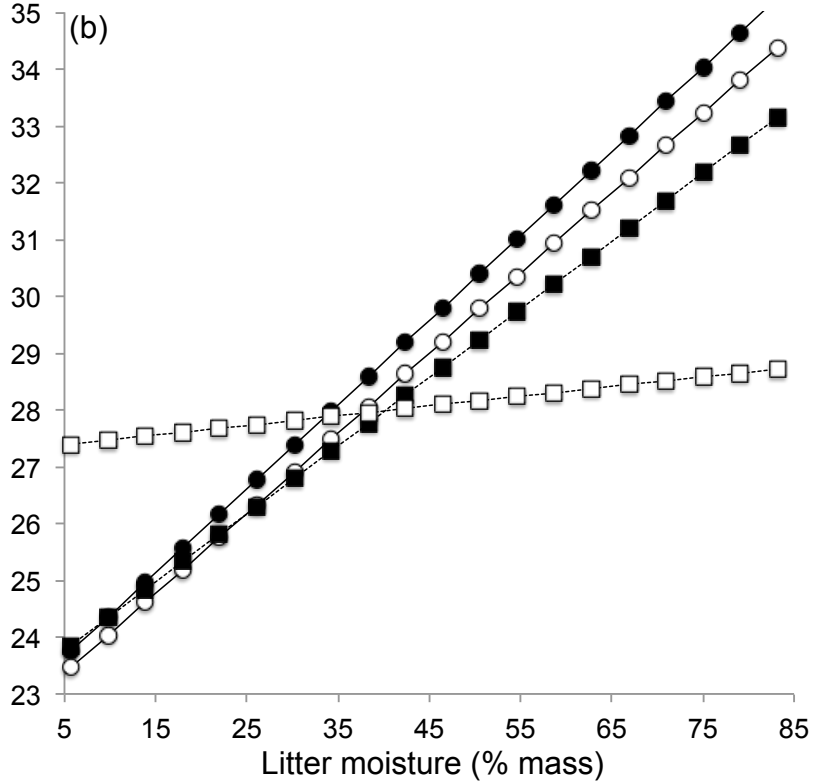
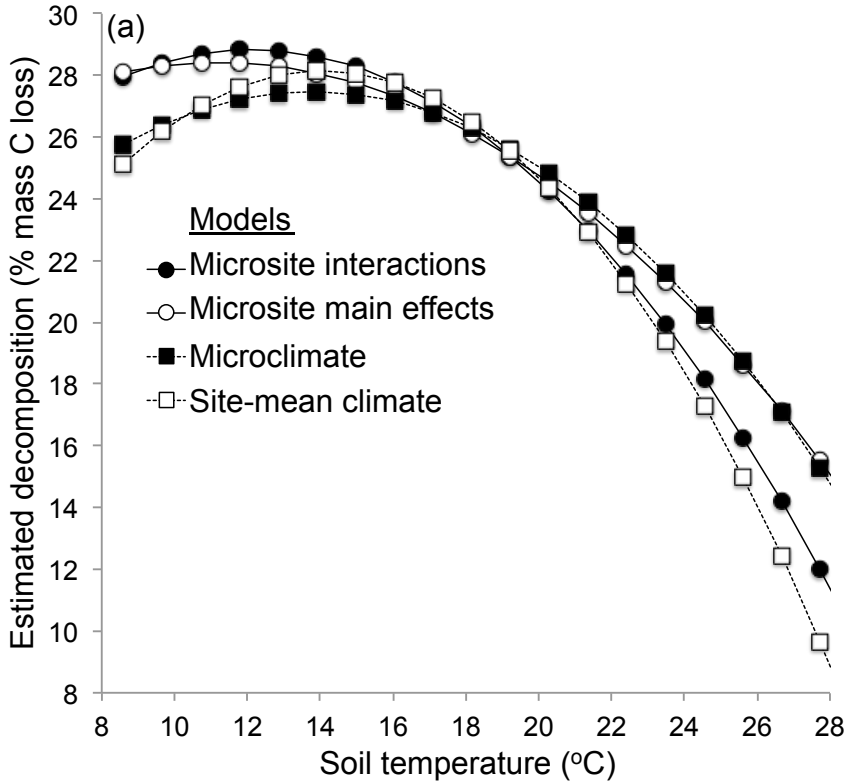
## Operational definitions of scale and variance

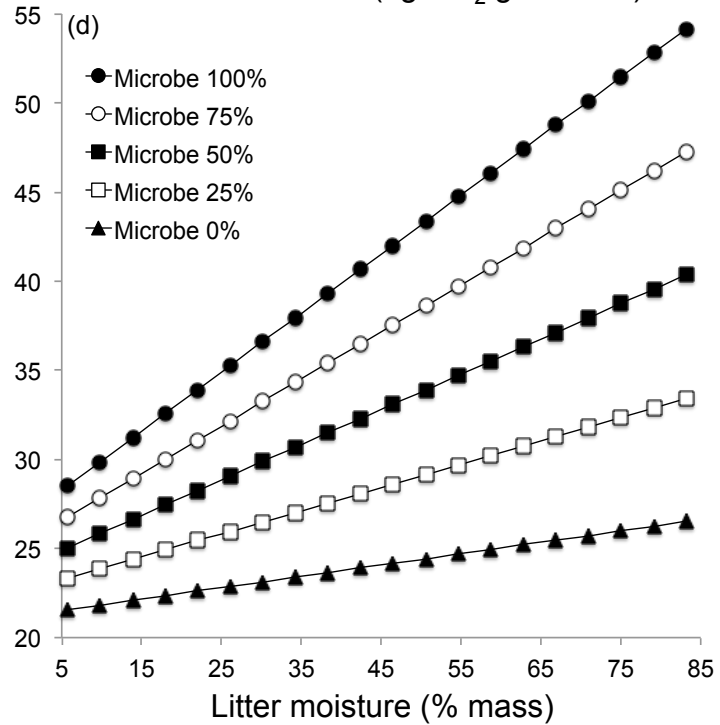
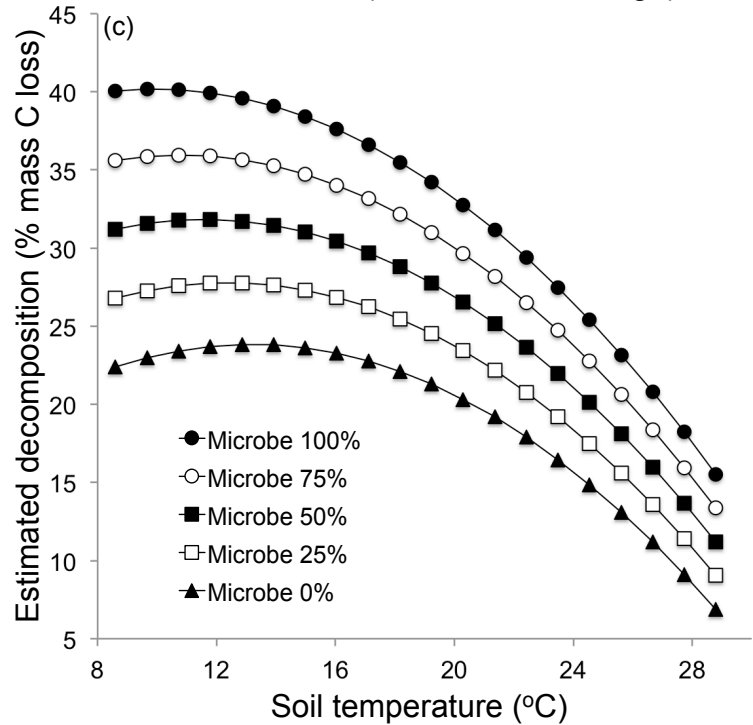
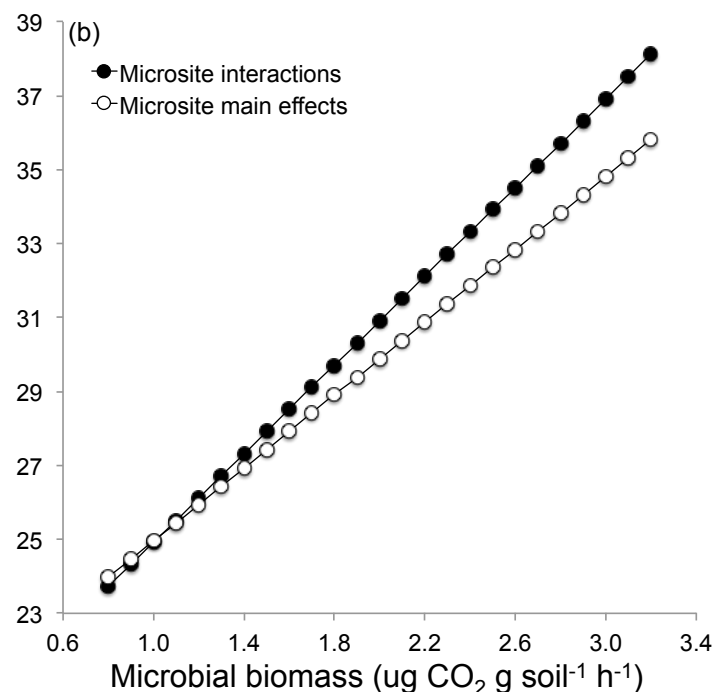
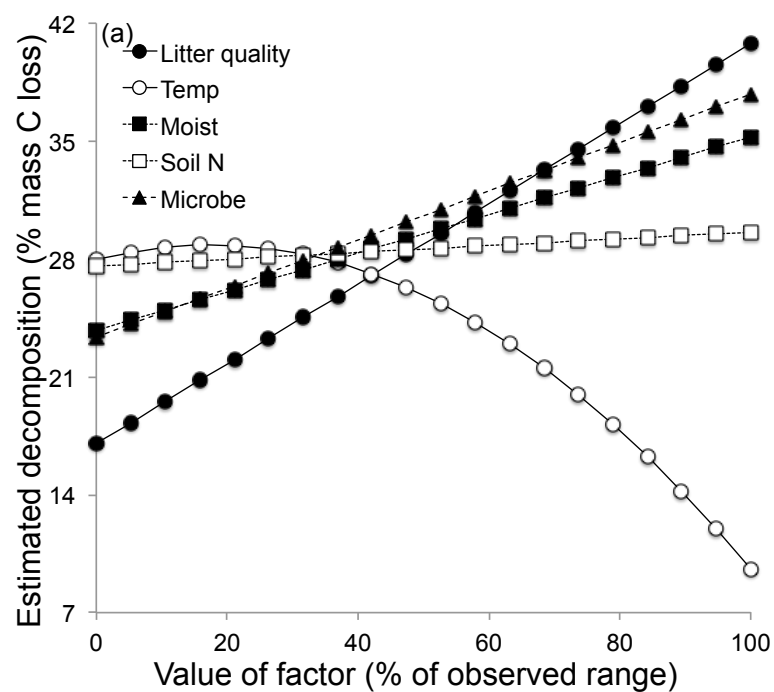


Variable	Site						
	<i>North</i>	1.	2.	3.	4.	5.	<i>South</i>
High temp. (°C)		12-20	16-22	15-20	17-21	18-23	22-29
Low temp. (°C)		2-10	6-13	7-13	8-13	8-13	11-17
Precip. (mm)		28-63	30-72	42-68	48-65	47-51	48-11
Soil temp. (°C)		10.0	13.5	16.6	13.7	15.6	25.3
Litter moist. (%)		51.5	29.3	70.5	31.8	13.2	7.2
Latitude (N)		63°48'	59°46'	55°22'	52°04'	48°11'	43°55'
Longitude (E)		020°14'	017°34'	011°34'	005°45'	-001°46'	003°05'









**Table 1** Coefficients, significance and  $r^2$  values for the linear mixed models used to evaluate controls on litter decomposition rates<sup>1</sup>. Shown in the second column are standardized coefficients for the full model, where “Microsite” refers to the level at which the variables were observed, and “interactions” to the inclusion of all 2-way interactions among the predictors.

Unstandardized coefficients were used when plotting Figs. 4, 5 and Supplementary Fig. 1. The consequence of aggregating microsite variation to generate “Site means” for the predictor variables was examined, but microsite variation in the response variable was retained to maintain the number of observations ( $n=303$ ). Significant ( $P<0.05$ ) and marginally-significant ( $P<0.1$ ) coefficients are shown in bold and italic fonts, respectively.

Variables	Model				
	Microsite interactions	<i>Unstandardized coefficients</i>			
		Microsite interactions	Microsite main effects	Microclimate	Site-mean climate
Intercept	27.0±0.689	-70.0±14.629	-17.1±6.264	-15.6±6.365	-24.1±6.960
Litter N	<b>16.1±0.856</b>	<b>45.3±5.998</b>	<b>19.3±1.173</b>	<b>19.2±1.198</b>	<b>22.6±1.283</b>
Temperature	<b>-4.49±1.600</b>	<b>5.03±1.344</b>	<b>1.05±0.702</b>	<b>1.73±0.681</b>	<b>2.81±0.759</b>
Temp <sup>2</sup>	<b>-6.84±3.285</b>	<b>-0.069±0.033</b>	<b>-0.047±0.018</b>	<b>-0.063±0.018</b>	<b>-0.100±0.021</b>
Moisture	<b>7.23±1.256</b>	<b>0.240±0.156</b>	<b>0.141±0.023</b>	<b>0.120±0.022</b>	0.017±0.028
Soil N	<b>0.732±1.075</b>	<b>0.151±0.158</b>	<i>0.014±0.028</i>	na	na
Microbe	<b>4.59±1.165</b>	<b>4.70±7.575</b>	<b>4.93±1.477</b>	na	na
Lit × Temp	<b>-13.9±1.888</b>	<b>-1.72±0.233</b>	na	na	na
Lit × Moist	-0.275±2.057	-0.007±0.049	na	na	na
Lit × soilN	1.58±1.666	0.053±0.056	na	na	na
Lit × Mic	0.347±1.997	0.535±3.077	na	na	na
Temp × Moist	<i>-7.03±4.157</i>	<i>-0.014±0.008</i>	na	na	na
Temp × soilN	<i>-3.09±2.035</i>	<i>-0.009±0.006</i>	na	na	na
Temp × Mic	<i>-1.46±2.172</i>	<i>-0.185±0.276</i>	na	na	na
Moist × soilN	-3.02±2.536	-0.002±0.001	na	na	na
Moist × Mic	4.55±2.923	0.111±0.071	na	na	na
soil N × Mic	-0.409±1.226	-0.014±0.042	na	na	na
model $r^2$	66.3	66.3	57.1	55.2	57.6

<sup>1</sup>Mean coefficients, their SD and significance are estimated using an MCMC sampling approach, and model  $r^2$  values using a method that retains the random effects structure (see Methods).

Model  $r^2$  values were identical for the fixed and full (i.e. fixed + random) effects.

Note: In the standardized Microsite interactions model, all sqrt VIFs were  $<2$  except Temperature<sup>2</sup> which was 2.98 and Temperature  $\times$  Moisture which was 2.30. In the unstandardized Microsite interactions model, all sqrt VIFs were  $<10$  except Temperature which was 16.0 and Temperature<sup>2</sup> which was 14.8.

In the unstandardized Microsite main effects model, all sqrt VIFs were  $<2$  except Temperature and Temperature<sup>2</sup>; and the same was observed with the Microclimate model, and the Site-mean climate model.

na = not applicable