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## **Biocompatible nanoparticles containing hydrophobic nickel-bis(dithiolene) complexes for NIR-mediated doxorubicin release and photothermal therapy**

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**Keywords:** Biocompatible nanoparticles, Biodegradable block copolymers, Photothermal Controlled Drug Release, nickel-bis(dithiolene) complex, Photothermal therapy.

**Abstract:** Biocompatible nanoparticles (NPs) constituted by amphiphilic poly(ethylene glycol)-*block*-poly(benzyl malate), PEG-*b*-PMLABe, have been designed for site-specific PhotoThermal Controlled Release (PTCR) of drug thanks to the presence of a Near Infra-Red (NIR) photothermally-active nickel-bis(dithiolene) complex in the inner core of NPs, together with Doxorubicin (Dox). Nanoprecipitation technique was used to prepare well-defined nickel-bis(dithiolene) and nickel-bis(dithiolene)/Dox loaded NPs, which were characterized by dynamic light scattering (DLS), zeta-potential measurements and Transmission Electron Microscopy (TEM). We have shown that the Dox release was effectively controlled by NIR irradiation (long or pulsed NIR laser irradiation). Cytotoxicity experiments on HeLa and MDA-MB-231 cells have shown that the incorporation of more than 10 w% of nickel-bis(dithiolene) complexes does not increase the intrinsic toxicity of the polymer nanoparticles. Finally, the viability of MDA-MB-231 cells was assessed after their incubation, for 24 hours, with empty NPs, Ni<sub>4</sub>C<sub>12</sub> loaded NPs, Dox loaded NPs or Ni<sub>4</sub>C<sub>12</sub>/Dox loaded NPs, without or with NIR irradiation. Above all, the results have highlighted that the Ni<sub>4</sub>C<sub>12</sub> loaded NPs after 5 min NIR laser irradiation can induced strong cell death up to 80 % at 50 µg/mL. These results demonstrate that these NPs are good candidates for photothermal therapy.

## ***Introduction***

To overcome the drawbacks associated to the administration of drugs, notably anti-cancer drugs, which are poorly water-soluble and lack of specific targeting towards malignant cells, tissues or organs, an alternative consists in encapsulating the drug into a nanocarrier [1]. In this context, the nanocarriers used as drug delivery systems are extremely diverse, ranging from inorganic porous materials to polymer-drug conjugates and nanoparticles [2-4]. Among those nanocarriers, nanoparticles (NPs) formed by amphiphilic block copolymers, also called polymeric micelles, have been, and continue to be, the subject of numerous researches as the result of their unique properties in aqueous solution [5-7]. Indeed, thanks to their amphiphilic nature, these block copolymers are able to self-assemble in aqueous solutions to form NPs with a hydrophilic corona protecting the nanocarrier from a rapid elimination by the immune system and a hydrophobic inner core allowing the encapsulation of hydrophobic or amphiphilic molecules [5-7].

Numerous amphiphilic block-copolymers have been designed to form polymeric micelles [5-7]. The hydrophilic block is mainly a poly(ethylene glycol), PEG, which is known to be biocompatible and approved by the US Food and Drug Administration (FDA) [8]. The PEG is associated with various hydrophobic blocks such as poly( $\epsilon$ -caprolactone), poly( $\beta$ -benzyl L-aspartate), poly(lactic-co-glycolic acid) and poly(lactic acid) [5-9].

The poly(malic acid), PMLA, and its derivatives constitute an interesting polymer family to develop block-copolymers able to form (bio)degradable and biocompatible nanocarriers [10]. Moreover, because both the synthesis of  $\beta$ -substituted  $\beta$ -lactones (monomers) and the anionic ring opening polymerization reaction are well-known and mastered, it is possible to obtain a large family of PMLA derivatives with properties adapted to the selected applications [10-15]. In this context, we have synthesized and characterized amphiphilic block-copolymers based on hydrophilic methoxy-PEG and hydrophobic PMLA<sub>Be</sub> [12-15].

Besides the choice of the nature of the block-copolymer which has an influence on the nanoparticles properties in terms of diameter, stability, drug loading efficiency, etc., one has to design the nanocarriers in order that the drug release can be controlled under either internal (enzyme, pH, etc.) [16-19] or external (irradiation, magnetic or electrical fields, etc.) stimuli [20-22]. Among the molecules responding to an external stimulus, photothermal agents, more specifically the Near InfraRed (NIR) dyes, are considered as promising candidates for application in the field of controlled release. Indeed, they are able to absorb light in the NIR region, thus allowing to improve detection sensitivity, to increase image contrast, to decrease damages to healthy cells and tissues, and to have a deeper tissue penetration [23-37]. Their

photothermal effect induces an accumulation of heat into the nanocarrier leading to a drug release. Various NIR dyes are used for the PhotoThermal Controlled Release (PTCR), among them the ones based on cyanine (IndoCyanine Green (ICG) and IR780) and porphyrin are often used [38]. These compounds have a high absorption in the NIR region and photothermal efficiency comparable to the one of gold nanostructures. However, their uses are limited due to their instability and their photobleaching upon laser irradiation as a result of the reaction between the NIR-dye and the singlet oxygen molecule ( $^1\text{O}_2$ ), which is produced during the irradiation [39]. Moreover, this singlet oxygen molecule can affect the drug, thus leading to the release of an inactive form of the drug. Consequently, molecules which do not produce reactive oxygenated species (ROS) are privileged in order to allow an efficient photothermal effect together with a release of non-degraded drug. Photothermal molecules, which do not produce singlet oxygen under NIR irradiation, such as the Croconaine (Croc), have been designed [40]. But these low molar mass molecules still have several drawbacks such as a very short half-life *in vivo* [41]. In that respect, we have demonstrated that metal-bis(dithiolene) complexes display strong photothermal activity under NIR irradiation in solid-state, in gel and in solution [42-45]. These complexes are strong NIR absorbers with efficient photothermal properties. In addition, these complexes are photothermally and photochemically stable and do not produce singlet oxygen under NIR irradiation. Thus, they are good candidates for the photocontrolled release of drugs from organic nanocarriers under NIR irradiation [46]. Among them, we selected the  $[\text{Ni}(\text{dphedt})_2]^0\text{-(C}_{12}\text{H}_{26})_4$  complex, Ni4C<sub>12</sub>, carrying four long C12 carbon chains for its hydrophobic properties allowing its encapsulation into the hydrophobic inner-core of the nanoparticles based on PEG-*b*-PMLABe block copolymers [12]. Finally, we have selected the Doxorubicin (Dox) as an anti-cancer drug model to evaluate its release from the nanoparticles upon NIR irradiation.

In addition, we have very recently demonstrated that the proper functionalization of nickel-bis(dithiolene) complexes with PEG chains has allowed good solubility in water. More importantly, *in cellulo* experiments have demonstrated that NIR-laser irradiation of cellular culture incubated with metal-bis(dithiolene) complexes can induce high level of cell death [45]. However, these complexes display non-negligible toxicity at high concentrations (Half maximal Inhibitory Concentration, IC<sub>50</sub>, around 500  $\mu\text{g}/\text{mL}$ ). Thus, the encapsulation and the concentration of these complexes inside biocompatible nanocarriers can be a good way to decrease their toxicity toward the biological medium and to increase locally their photothermal activity.

Therefore, we describe, within this paper, the synthesis and characterization of PEG-*b*-PMLABe based nanoparticles loaded with Ni<sub>4</sub>C<sub>12</sub> complex or with Ni<sub>4</sub>C<sub>12</sub> complex and Dox. The Dox release from nanoparticles was followed with and without long NIR laser irradiation as well as laser pulses. Finally, the *in vitro* cytotoxicity of empty nanoparticles, free Dox, Ni<sub>4</sub>C<sub>12</sub> loaded nanoparticles, Dox loaded nanoparticles and Ni<sub>4</sub>C<sub>12</sub> complex and Dox loaded nanoparticles was evaluated on two cell lines: the HeLa and MDA-MB-231 cells with and without laser irradiation.

### ***Experimental part***

#### **Materials.**

PEG<sub>46</sub>-*b*-PMLABe<sub>77</sub> block copolymers were synthesized as previously described [12]. Briefly, the benzyl malolactonate, monomer obtained in four steps starting from aspartic acid [47], was polymerized by anionic ring-opening polymerization in presence of tetraethylammonium salt of  $\alpha$ -methoxy,  $\omega$ -carboxylate PEG<sub>46</sub> as initiator. After purification, the block copolymers were characterized by proton NMR (structure and molar mass of the PMLABe block, Table 1) and size exclusion chromatography in THF (molar masses and dispersity, Table 1).

Nickel-bis(dithiolene) complexes Ni<sub>4</sub>C<sub>12</sub> have been prepared as previously reported [48]. Briefly, 4,4'-bis(dodecanoxy)benzyl was sulfureted with P<sub>4</sub>S<sub>10</sub> in 1,3-dimethyl-2-imidazolidinone (DMI), followed by hydrolysis of the intermediate phosphorous thioesters in the presence of the nickel salt such as NiCl<sub>2</sub>·6H<sub>2</sub>O to afford directly the oxidized, neutral nickel complex. The compound was purified by column chromatography on silica gel and crystallized by slow evaporation dichloromethane from a CH<sub>2</sub>Cl<sub>2</sub>/MeOH mixture.

Doxorubicin and anhydrous THF were purchased from Sigma-Aldrich and used as received.

#### **Methods.**

PEG<sub>46</sub>-*b*-PMLABe<sub>77</sub> NPs preparation. Nanoparticles were prepared using the nanoprecipitation method described by Thioune et al. based on the self-assembly of hydrophobic or amphiphilic (co)polymer in aqueous medium [49]. To this end, the polymer is first solubilized in a water-miscible organic solvent, such as acetone or THF, and then rapidly added to an aqueous solution. The PEG<sub>46</sub>-*b*-PMLABe<sub>77</sub> (5 mg) was thus solubilized in 1 mL of THF and rapidly added to 2 mL of ultrapure water under vigorous stirring. The mixture was then stirred at room temperature for 10 min. In this process, the hydrophobic blocks of the copolymer (PMLABe) aggregate, leaving the hydrophilic blocks (PEG) in contact with the water, and leading to the formation of nanoparticles formed by the PEG<sub>46</sub>-*b*-PMLABe<sub>77</sub> block copolymers. The THF was

then evaporated under reduced pressure (rotary evaporator) and the final volume was adjusted to 2 mL by addition of ultrapure water.

NPs[Ni<sub>4</sub>C<sub>12</sub>] preparation. 5 mg of PEG<sub>46</sub>-*b*-PMLABe<sub>77</sub> were solubilized in 800  $\mu$ L of THF and 200  $\mu$ L of a solution of Ni<sub>4</sub>C<sub>12</sub> in THF at a concentration of 2.5 mg/mL were then added, so that the amount of complex is 10% of mass of polymer (10% / polymer), *i.e.* 0.5 mg. This solution (polymer + complex in THF) was then rapidly added in 2 mL of ultrapure water under vigorous stirring. The mixture was stirred at room temperature for 10 min, then the THF was evaporated under reduced pressure (rotary evaporator). The final volume was adjusted to 2 mL by addition of ultrapure water to give a brown solution of [Ni<sub>4</sub>C<sub>12</sub>] loaded nanoparticles ([C] polymer = 2.5 mg/mL, [C] complex = 250  $\mu$ g/mL).

NPs[Dox] preparation. A stock solution of Dox, HCl was first prepared in a THF/NEt<sub>3</sub> mixture (6 mL THF + 23  $\mu$ L NEt<sub>3</sub>), as follow 10 mg of Dox was solubilized in 4 mL of THF/NEt<sub>3</sub> ([Dox] stock solution = 2.5 mg/mL). Thereafter, 5 mg of PEG<sub>46</sub>-*b*-PMLABe<sub>77</sub> polymer were solubilized in 800  $\mu$ L of THF, and 200  $\mu$ L of the Dox solution (10% / polymer, *ie* 0.5 mg, initially introduced = 0.25 mg/mL) are added. This solution (polymer + Dox in THF) was then rapidly added in 2 mL of ultrapure water under vigorous stirring. The mixture was stirred at room temperature for 10 minutes and then the THF was evaporated under reduced pressure (rotary evaporator). The final volume is adjusted to 2 mL by adding ultrapure water. The unloaded Dox was eliminated by centrifugation using a MicroCon system as follow: the orange solution was ultracentrifuged at 15,000 g for 7 min using a filter with an exclusion limit of 10,000 Da. Then, the filter was returned and centrifuged at 1,000 g for 1 min. The concentrate was diluted with ultrapure water to obtain 2 mL of final solution. The Dox encapsulation efficiency was determined by measuring the absorbance of the Dox at 485 nm by UV-Vis spectroscopy. First, a calibration curve of free Dox in a water/DMF (20/80) mixture was realized as follows: 500  $\mu$ L of the Dox stock solution in THF/NEt<sub>3</sub> was evaporated under reduced pressure. Then, 1 mL of water/DMF (20/80) was added. This concentrated solution ([Dox] = 1.25 mg/mL) was diluted to obtain solutions with decreasing Dox concentration by adding the water/DMF mixture (20/80). The absorbance at 485 nm of the different samples of free Dox ( $V = 400 \mu$ L) was then measured by UV-Vis spectroscopy. Once the calibration curve was realized (Figure S1), 80  $\mu$ L of the NPs[Dox] suspension were diluted with 320  $\mu$ L of DMF and its absorbance was measured at 485 nm by UV-Vis spectroscopy. This value was transformed into a concentration thanks to the calibration curve. Therefore, the concentration of encapsulated Dox

was 108  $\mu\text{g/mL}$ , meaning that 22% of the initially introduced Dox has been encapsulated (4% relative to the polymer mass).

NPs[Ni<sub>4</sub>C<sub>12</sub>,Dox] preparation. 5 mg of PEG<sub>46</sub>-*b*-PMLABe<sub>77</sub> polymer were solubilized in 600  $\mu\text{L}$  of THF, 200  $\mu\text{L}$  of the complex stock solution at a concentration of 2.5 mg/mL in THF (10% / polymer, ie 0.5 mg of Ni<sub>4</sub>C<sub>12</sub> ) and 200  $\mu\text{L}$  of the Dox solution (10% / polymer, ie 0.5 mg, initially introduced = 0.25 mg/mL) are added. This solution (polymer + complex + Dox in THF) was then rapidly added in 2 mL of ultrapure water under vigorous stirring. The mixture was stirred at room temperature for 10 minutes and then the THF was evaporated under reduced pressure (rotary evaporator). The final volume is adjusted to 2 mL by adding ultrapure water. The unloaded Dox was eliminated by centrifugation using a MicroCon system as described above. The Dox encapsulation efficiency was determined by measuring the absorbance of the Dox at 485 nm by UV-Vis spectroscopy using a calibration curve of free Dox in a water/DMF (20/80) mixture realized as described above (Figure S1); then, 80  $\mu\text{L}$  of the NPs[Ni<sub>4</sub>C<sub>12</sub>,Dox] suspension were diluted with 320  $\mu\text{L}$  of DMF and its absorbance was measured at 485 nm by UV-Vis spectroscopy. This value was transformed into a concentration thanks to the calibration curve. Therefore, the concentration of encapsulated Dox was 126  $\mu\text{g/mL}$ , meaning that 25% of the initially introduced Dox has been encapsulated (5% relative to the polymer mass).

DLS experiments. The hydrodynamic diameters ( $D_h$ ) and polydispersity indexes (PDI) of the prepared NPs were analyzed by dynamic light scattering (DLS) with a Zetasizer Nano-ZR90 (Malvern) apparatus at 25°C using a He6Ne laser at 633 nm and a detection angle of 90°.

TEM Measurements. TEM analyses of the NPs were realized at the “Institute des Sciences Chimiques de Rennes”. Each sample (5 $\mu\text{L}$ ) was deposited on a Formvar-carbon film coated on a copper grid (300 mesh). After 6 min, the excess of water was removed by absorption with filter paper and the sample was stained with phosphotungstic acid (0.1 v%) for 30 s. The grid was then placed under the electron beam. The microscope used was a JEOL 2010 LaB6 operating at 80 kV. The camera, on which the images were recorded, was a GATAN Orius 200D CCD (Charge Coupled Device) camera.

UV-vis-NIR titration. UV-Vis-NIR absorption spectra in solution were recorded on a Shimadzu UV3600 Plus spectrophotometer. Samples were placed in 1 cm path length quartz cuvettes.

Photothermal studies. For the photothermal studies, 1 mL of NPs suspensions was irradiated through a quartz cuvette with a 940 nm-wavelength semiconductor laser (BWT Beijing LTD) for 10 min. The power intensity of the laser could be adjusted externally (0-10 W). The output power was independently calibrated using an optical power meter. A thermocouple with an accuracy of  $\pm 0.1$  °C connected to an Agilent U1253B multimeter was inserted into the solution. The thermocouple was inserted at such a position that the direct irradiation of the laser was avoided. The temperature was measured every 1s.

Measurement of Dox release from the NPs. The Dox release was directly studied in a quartz cuvette using a home-made dialysis system. For this, a dialysis membrane with a cut-off threshold of 3,500 Da was immersed in milliQ water for 30 minutes and then rinsed with clean water. The dialysis film was cut to the desired size and then placed on a 100  $\mu$ L Eppendorf tip of which has been cut to 2.9 cm from the top. The dialysis membrane was held on the tip with a plastic paraffin film (Parafilm). The system was then introduced into a quartz cuvette having a circular aperture and containing 2.7 mL of water. 150  $\mu$ L of the NPs suspension were then placed in the modified tip. The absorption spectra for the Dox release in water were recorded at room temperature using a Shimadzu UV3600 Plus spectrophotometer. For measurements under laser irradiation, the samples were irradiated at 5 W/cm<sup>2</sup> at 940 nm for a given time by the top of the quartz cuvette.

Cell culture. The HeLa and MDA-MB-231 cells were cultured in DMEM supplemented with 10 % fetal bovine serum (FBS), 100 U/mL penicillin and 100  $\mu$ g/mL streptomycin at 37 °C in a humidified 5 % CO<sub>2</sub> atmosphere.

*In vitro* evaluation of cytotoxicity. The effect of NPs on cell growth was assessed in 96 wells plates (Microtest<sup>TM</sup>-96-Becton Dickinson). The cells were seeded at 2,500 cells per well of full medium (100  $\mu$ L/well). The NPs suspensions at the appropriate concentration (50, 125, 250  $\mu$ g/mL) were added 24 hours after seeding for further 24 h. After exposure to the NPs, cell viability was determined using the MTT (thiazolyl-blue-tetrazolium-bromide, MTT from Sigma) assays [50]. In parallel, the effect of radiation was studied after the wells were irradiated with a 940 nm diode laser at 5 W/cm<sup>2</sup> for 5 min from the top. Then, the MTT tests were undertaken. The absorbance of the solutions was measured at wavelength of 570 nm using a BMG Labtech FLUOstar Optima plate reader. Triplicates were done for each group of treated cells



## Results and Discussion

Recently, we have demonstrated that the Ni-bis(dithiolene) complexes are promising photo-active agents for PTCR from liposomes under NIR irradiation, as a result of their good thermal and photo-stability, and their high photothermal activity [46]. To overcome the problems linked to the fast elimination *in vivo* of phospholipids, we proposed to encapsulate the hydrophobic derivatives of Ni-bis(dithiolene) into the inner-core of biocompatible nanoparticles constituted by the amphiphilic block copolymers PEG<sub>46</sub>-*b*-PMLABe<sub>77</sub> (the number representing the number of monomer units in each block, i.e. 46 units of ethylene glycol (molar mass of PEG = 2,015 g/mol) and 77 units of MLABe (molar mass of PMLABe = 15,860 g/mol)). The PEG<sub>46</sub>-*b*-PMLABe<sub>77</sub> block copolymers used in this study were synthesized by anionic ring-opening polymerization of the benzyl malolactonate, MLABe, using an  $\alpha$ -methoxy, $\omega$ -carboxylate PEG initiator, as described previously [12-15]. The molar mass of the PMLABe block was fixed by the ratio MLABe/PEG to a value of 77, corresponding to a theoretical PMLABe molar mass of 15,000 g/mol. After purifications, the PEG<sub>46</sub>-*b*-PMLABe<sub>77</sub> block copolymers were analyzed, as described previously [12], by <sup>1</sup>H NMR and Size Exclusion Chromatography (SEC); the NMR spectrum of the block copolymer allowed us to calculate the molar mass of the PMLABe block, knowing the one of the commercially available PEG block, while the SEC analysis gave access to the average molar mass and the dispersity (Table 1).

Theoretical Molar Mass PMLABe (g/mol) <sup>a</sup>	Given Molar Mass PEG (g/mol) <sup>b</sup>	Calculated Molar Mass PMLABe (g/mol) <sup>c</sup>	Molar Mass PEG- <i>b</i> -PMLABe (g/mol)	M <sub>w</sub> (g/mol) <sup>d</sup>	D <sup>d</sup>
15,000	2,015	15,860	17,880	5,460	1.38

*a. Determined by the ratio MLABe/initiator; b. PEG bought from PEG Iris Biotech; c. Molar mass calculated from <sup>1</sup>H NMR spectrum of the copolymer; d. Measured by SEC in THF (1ml/min, 40°C, Poly(styrene) standards).*

**Table 1.** Characteristics of the PEG<sub>46</sub>-*b*-PMLABe<sub>77</sub> block copolymer used in this study.

This PEG<sub>46</sub>-*b*-PMLABe<sub>77</sub> block copolymer allowed us to prepare the corresponding nanoparticles (NPs) using the nanoprecipitation method, initially described by Thioune et al. [49]. The block-copolymer, the Ni<sub>4</sub>C<sub>12</sub> and the Dox are dissolved in an organic solvent miscible with water, in our study THF, and this solution is rapidly added to water under vigorous stirring.

As a result of their nature, the block-copolymer chains are able to immediately self-assemble leading to the formation of well-defined nanoparticles, after evaporation of THF under reduced pressure. When the nanoprecipitation is realized in presence of Ni4C<sub>12</sub>, Dox or Ni4C<sub>12</sub>/Dox, these non-encapsulated molecules were eliminated by ultracentrifugation/filtration using Microcon system. The solution contained in the Microcon, after ultracentrifugation, was kept and correctly diluted. The empty, Dox, Ni4C<sub>12</sub> and Ni4C<sub>12</sub>/Dox loaded nanoparticles were then characterized by dynamic light scattering (DLS), and zetametry. The two analytical methods give access to the hydrodynamic diameter, polydispersity and surface charge of the nanoparticles. The amount of encapsulated Ni4C<sub>12</sub> and Dox was evaluated using UV analysis. The characteristics of the obtained NPs are gathered in Table 2.

	DLS <sup>a</sup>		Zetametry (mV) <sup>a</sup>	wt% Ni4C <sub>12</sub> <sup>b</sup>	wt% Dox <sup>c</sup>
	Average diameter (nm)	Dispersity			
Empty NPs	90	0.14	-25	--	--
NPs[Dox]	80	0.24	-24	--	4
NPs[Ni4C <sub>12</sub> ]	110	0.18	-27	10	--
NPs[Ni4C <sub>12</sub> ,Dox]	100	0.22	-26	10	5

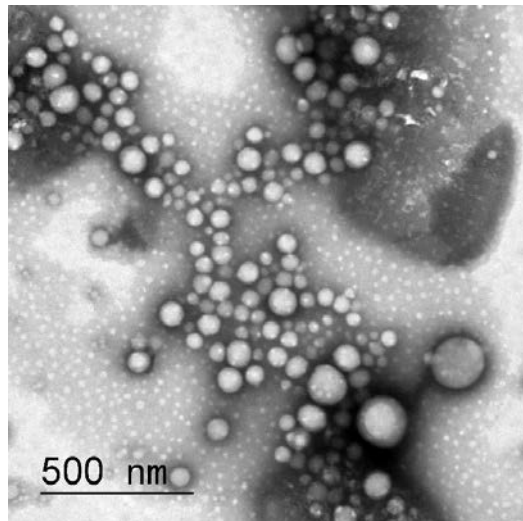
*a. Measurements realized on a Nanosizer Nano-ZR90 (Malvern); b. Confirmed by UV-vis-NIR absorption measurements performed on the filtrate; c. Determined by UV at 485nm using a standard curve established with Dox solutions with various concentrations.*

**Table 2.** Characteristics of empty, Dox, Ni4C<sub>12</sub> and Ni4C<sub>12</sub>/Dox loaded NPs.

DLS measurements confirmed the formation of nano-objects with an average size of about 100 nm and a quite narrow dispersity of around 0.1-0.2 (Table 2). Due to the encapsulation of Ni4C<sub>12</sub> complex, the diameter of the NPs[Ni4C<sub>12</sub>] is greater (110 nm) than that of the empty polymeric nanoparticles (90 nm) (Figure S2-S3). The dispersity of the NPs[Ni4C<sub>12</sub>] (0.18) and the negative zeta potential (-27 mV) indicate that the nanoparticles are homogeneous and do not form any aggregate in solution. DLS measurements performed on the NPs[Dox] and the NPs[Ni4C<sub>12</sub>,Dox] suspensions show that the encapsulation of the Dox has little influence on the final size of the nanoparticles (Figure S4).

UV-Vis-NIR absorption measurements have revealed that using 10 wt% of complex /polymer, all the complex is encapsulated inside the nanoparticles. In fact, no NIR signal has been detected in the filtrate (Figure S5). The Ni4C<sub>12</sub> complex encapsulated in PEG<sub>46</sub>-*b*-PMLABe<sub>77</sub> NPs (NPs[Ni4C<sub>12</sub>]) and suspended in water has an absorption maximum at 914 nm and a molar

absorption coefficient of about 10,200 L/mol/cm. It should be noted that these values are lower than that of the free Ni<sub>4</sub>C<sub>12</sub> complex in solution in dichloromethane ( $\lambda_{\text{max}} = 933 \text{ nm}$  and  $\epsilon = 31,300 \text{ L/mol/cm}$ ). Attempts have been made to increase the amount of encapsulated complex. Introduction of 50 wt% of complex has been tried in the polymer NPs (50 wt% complex/polymer). However, only 13wt% of the Ni<sub>4</sub>C<sub>12</sub> polymer was efficiently encapsulated as revealed by UV-vis-NIR measurements performed on the aqueous suspension. Due to the insolubility of the complex in water, the excess product remains adhered to the walls of the round-bottom flask during preparation. Moreover, the DLS measurements reveal the presence of large nanoparticles with a size of 240 nm. This increase in size is probably due to the presence of nano-aggregates of non-encapsulated Ni<sub>4</sub>C<sub>12</sub> in the suspension of NPs[Ni<sub>4</sub>C<sub>12</sub>]. Therefore, it can be concluded that a maximum of 13 wt% of Ni<sub>4</sub>C<sub>12</sub> complex can be incorporated in the polymeric nanovectors and thus the use of 10 wt% Ni<sub>4</sub>C<sub>12</sub> /polymer is a good compromise for PTCR applications and photothermal therapy. UV-Vis-NIR measurements have also revealed that 25 wt% of the initially introduced Dox has been encapsulated, evaluated to 5 wt% relative to the polymer mass.

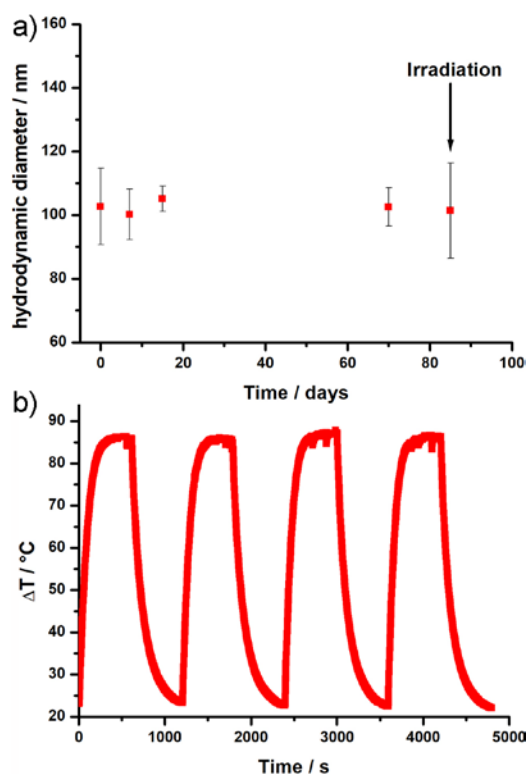


**Figure 1.** Image obtained by TEM of the NPs[Ni<sub>4</sub>C<sub>12</sub>] using a Jeol 2010 LaB6 apparatus operating under a voltage of 80KeV. The sample was previously stained with 0.1v% of phosphotungstic acid.

The empty and Ni<sub>4</sub>C<sub>12</sub> loaded NPs were analyzed by transmission electron microscopy, Figure S6 and Figure 1, respectively. As shown by Figures 1 and S6, both the empty and Ni<sub>4</sub>C<sub>12</sub> loaded

nanoparticles have a spherical shape. The images show a quite large size repartition with an average diameter of 60 nm with several small NPs and few larger ones. The average diameter measured by TEM is lower than the one measured by DLS because the hydration layer is not visible by TEM.

One important evaluated parameter is the stability of Ni<sub>4</sub>C<sub>12</sub> loaded PEG<sub>46</sub>-*b*-PMLABe<sub>77</sub> NPs. For that, we have followed the evolution of the NPs diameter with time by DLS measurements (Figure 2a). As shown in Figure 2a, the average diameter remained stable at around 110 nm over three months, highlighting a good stability of the NPs in aqueous solution. Moreover, we have irradiated the solution containing the NPs[Ni<sub>4</sub>C<sub>12</sub>] after 86 days in suspension (Figure 2a). Suspensions of NPs[Ni<sub>4</sub>C<sub>12</sub>] have also been irradiated following several irradiation cycle given in Figure 2b. Irradiation has been performed with a 940-nm laser wavelength which is close to the absorption maxima of the Ni<sub>4</sub>C<sub>12</sub> complex in the NIR region (Figure S5). Several conclusions can be drawn from the results presented in Figure 2.



**Figure 2. a.** Evolution of the average diameter of NPs[Ni<sub>4</sub>C<sub>12</sub>] in water as a function of time ( $C = 2.5$  mg/mL). After 86 days, identical hydrodynamic diameters have been measured before and after laser irradiation (940 nm, 10 min, 5 W/cm<sup>2</sup>) (error bars have been determined on three different batches); **b.** Measurement of the temperature rise of a suspension of NPs[Ni<sub>4</sub>C<sub>12</sub>] in water as a function of time and under laser irradiation ( $C = 2.5$  mg/mL of

*nanoparticles, C = 0.25 mg/mL of Ni4C12 complex), 940 nm, 5 W/cm<sup>2</sup>, 10 min ON and 10 min OFF, 4 cycles).*

First, we observed that the average diameter of the NPs[Ni4C12] before and after irradiation (Figure 2a) remains unchanged (110 nm), highlighting the stability of the nanoparticles under NIR irradiation. TEM experiments performed after irradiation have confirmed that the morphology of the NPs is not affected by the laser irradiation (Figure S7). UV-vis-NIR experiments have also revealed that the complex is not degraded during the irradiation (Figure S8). Second, these results highlight the photothermal effect of the NPs[Ni4C12] with a temperature increase of more than 85°C. It should be also noted that this photothermal effect is constant and reproducible even after several irradiation cycles. Moreover, the NIR irradiation of pure water and empty NPs suspension only lead to a moderated increase of temperature (+15°C), while the irradiation of a Ni4C12 loaded NPs suspension induces a drastic increase in temperature of about 75°C (Figure S9). Therefore, the strong photothermal property of NPs[Ni4C12] is a result of the presence of Ni4C12 within the NPs inner core. Finally, the photothermal efficiency ( $\eta$ ) of the NPs[Ni4C12] was determined using the equation given by Roper et al. [50] (Figure S10): the calculated  $\eta$  has a value of 26%. Such value is higher than the one calculated for liposomes encapsulating, in its inner aqueous core, another PEGylated nickel-bis(dithiolene) complex [46], probably due the encapsulation in the hydrophobic core which prevents deactivation by the surrounding water molecules.

In conclusion, all these results highlighted that the PEG<sub>46</sub>-*b*-PMLABe<sub>77</sub> based NPs encapsulating the complex Ni4C12 were stable both in time and under laser irradiation, together with a good photothermal efficiency.

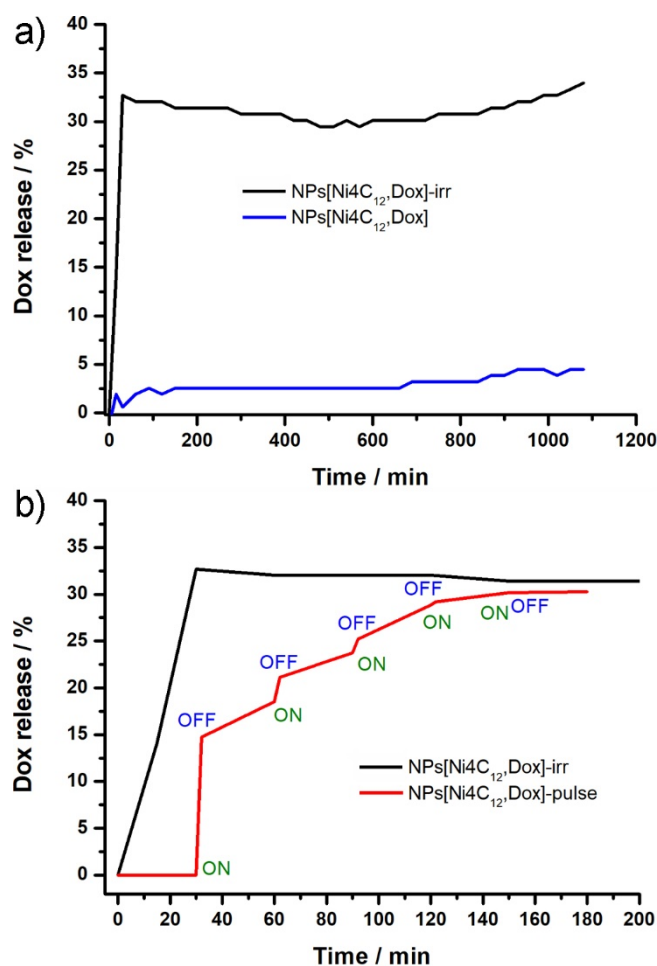
The Dox release from NPs[Ni4C12,Dox] upon NIR-irradiation was evaluated by dialysis with and without laser irradiation at 940 nm. The absorbance of the out-side solution containing the released Dox was measured regularly by UV at 485 nm. As shown by Figure 3, several experiments were realized: Dox release without NIR irradiation, Dox release after a prolonged NIR irradiation of 15 min at T<sub>0</sub> and a NIR irradiation of 2 min every 30 min. The percent of Dox released with time was calculated using the Equation 1:

$$\% \text{ of released Dox} = \frac{A_t - A_0}{A_f} * 100$$

*Where A<sub>t</sub>: absorbance measured at t, A<sub>0</sub>: absorbance measured at t=0 and A<sub>f</sub>: absorbance measured at the end of the experiment by adding a drop of DMF to break the NPs and release the Dox still encapsulated.*

**Equation 1.** Calculation of Dox release from the absorbance measured by UV at 485 nm.

On Figure 3a, the amount of Dox released from the NPs[Ni4C<sub>12</sub>,Dox] without NIR irradiation was lower than 5%, while upon 15 min of NIR irradiation at the beginning of the release process more than 30% of Dox were released after 30 min. However after 18 hours of incubation, no more Dox was released from the nanocarrier, probably as a result of the low solubility of Dox in aqueous medium preventing its diffusion outside the polymer network constituting the NPs or by a higher affinity of the Dox for the hydrophobic PMLABe inner core. Nevertheless, this experiment has highlighted that the NIR irradiation induces a larger and faster Dox release from the NPs, probably because of a change in the NPs permeability resulting from the temperature increase due to the irradiation of the NIR-sensitive dye.



**Figure 3.** Dox release (%) as a function of time from a solution of PEG<sub>46</sub>-b-PMLABe<sub>77</sub> based NPs containing Dox and Ni4C<sub>12</sub>: a. without and with a NIR irradiation (940 nm, 5 W/cm<sup>2</sup>, 15 min at the beginning); b. with an initial 15 min NIR irradiation (940 nm, 5 W/cm<sup>2</sup>, 15 min) or with several intermittent NIR irradiations (940 nm, 5 W/cm<sup>2</sup>, 2 min ON, 30 min OFF).

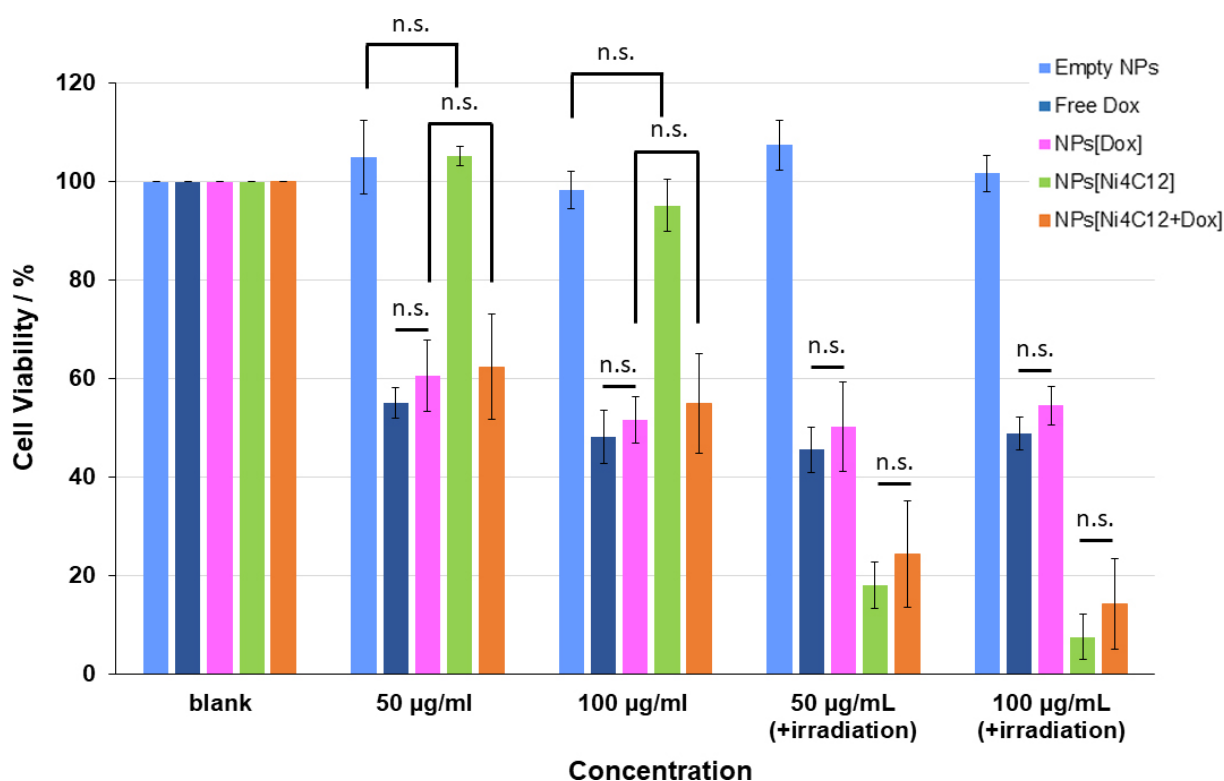
More interestingly, additional experiments were realized with a sequential NIR irradiation of the NPs[Ni<sub>4</sub>C<sub>12</sub>,Dox] suspension (Figure 3b). As shown by the graph of Figure 3b, 15% of Dox was released after the first NIR irradiation. When the NIR irradiation was stopped, the Dox release slowed down and accelerated again during the next step of NIR irradiation until the plateau at 30% of Dox release is reached. At that time, the effect of NIR irradiation is minor. The maximum Dox release is achieved, in this case, after 3 hours in suspension. The exposition of NPs to the laser light probably increases definitively their permeability. Indeed, a slow Dox release is observed during the period without NIR irradiation, release which is, moreover, faster than the one observed for the NPs[Ni<sub>4</sub>C<sub>12</sub>,Dox] not submitted to NIR irradiation. It should be noticed that UV-vis-NIR titration performed on the dialysate after irradiation did not show absorption in the NIR region, meaning that this hydrophobic Ni<sub>4</sub>C<sub>12</sub> nickel-bis(dithiolene) complex remains within the polymer nanoparticles.

From these results, we can conclude that the NPs[Ni<sub>4</sub>C<sub>12</sub>] might be used for PTCR. Moreover, depending on the duration and the number of NIR irradiation, the speed of the drug release can be tuned according to the drug delivery needed for a specific therapy in term of quantity and duration. However, studies to optimize the system are still to be considered. Indeed, the rate of release can probably be further decreased, using lower irradiation powers.

In parallel, we have studied the *in cellulo* cytotoxicity of the NPs[Ni<sub>4</sub>C<sub>12</sub>] on two cell lines, the MDA-MB-231 and the HeLa cells, using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assays. As shown by Figure S11, the NPs[Ni<sub>4</sub>C<sub>12</sub>] have a higher cytotoxicity on HeLa cells than on MDA-MB-231 cells. Indeed, we have observed a half maximal inhibitory concentration (IC<sub>50</sub>, being the concentration at which 50% of the cells were dead) around 300 µg/mL in HeLa cells, while, only a 80% maximal inhibitory concentration (IC<sub>80</sub>) of 300 µg/mL was observed with the MDA-MB-231 cell line (Figure S11). To further investigate the impact of the presence of the Ni<sub>4</sub>C<sub>12</sub> complexes and of the Dox combined or not with laser irradiation on the *in vitro* cytotoxicity, additional experiments have been performed on the more robust MDA-MB-231 cells.

The cell viabilities of the MDA-MB-231 cells incubated with empty NPs, free Dox, NPs[Dox], NPs[Ni<sub>4</sub>C<sub>12</sub>] and NPs[Ni<sub>4</sub>C<sub>12</sub>,Dox] are presented in Figure 4. These experiments have been performed at concentrations up to 100 µg/mL where the toxicity of the NPs remains negligible. First, comparison of the cells viability between the empty NPs and the NPs[Ni<sub>4</sub>C<sub>12</sub>] without laser irradiation shows that the introduction of 10 wt% of Ni<sub>4</sub>C<sub>12</sub> complexes inside the NPs does not induce additional toxicity since the cell viability was almost identical for all the tested

concentrations. Moreover, the cells viability is not affected by the application of a laser irradiation in the absence of Ni<sub>4</sub>C<sub>12</sub> complexes inside the NPs, meaning that the laser irradiation alone has no effect on the cell viability.



**Figure 4.** Relative cell viability of MDA-MB-231 cells incubated at 50 and 100 µg/mL of empty NPs NPs[Dox], NPs[Ni<sub>4</sub>C<sub>12</sub>] and NPs[Ni<sub>4</sub>C<sub>12</sub>,Dox] without or with laser irradiation (5 min, 5 W/cm<sup>2</sup>, 940 nm). Free Dox was introduced at 2.5 and 5 µg/mL which corresponds to 5 w% of the NPs concentration. n.s.: non-significant. Statistical analyses were realized based on ANOVA Post Hoc Test (Tukey multiple comparisons of means or Wilcoxon rank sum test), only the non-significant values were marked on the figure for clarity's reasons.

Below 25 µg/mL of NPs[Dox] or equivalent 1.25 µg/mL of free dox, no significant effect have been detected on the cell viability. However, the cell viability drops to 80 % at 25 µg/mL, 60 % at 50 µg/mL and 50 % at 100 µg/mL with the Dox loaded NPs. These results show that the Dox is likely naturally released from the NPs after one day of incubation and that Dox starts to be toxic at concentrations of NPs[Dox] higher than 25 µg/mL. Comparison of the results obtained on NPs[Dox] and equivalent quantities of free Dox shows that all the Dox has been



release from the nanoparticles after one day incubation. Laser irradiation does not provide any additional effect.

In presence of Ni<sub>4</sub>C<sub>12</sub> complexes, NPs[Ni<sub>4</sub>C<sub>12</sub>] or NPs[Ni<sub>4</sub>C<sub>12</sub>,Dox], higher cell death can be induced under laser irradiation above 50 µg/mL. The cell viability after laser irradiation decreases from 60 to 20 % at 50 µg/mL and from 50 to 15 % at 100 µg/mL. In absence of Ni<sub>4</sub>C<sub>12</sub> complexes, laser irradiation does not affect the cell viability. The synergetic effect of the Dox and the laser irradiation is not noticeable. At this stage, based on the statistical analyses (Figure 4), there is no clear evidence of a potential cumulative effect of the presence of dox in addition to the already highly efficient photothermal effect induced by the presence of the Ni<sub>4</sub>C<sub>12</sub> complex under laser irradiation. The strong cell death of the MDA-MB-231 cells upon laser irradiation in the presence of NPs[Ni<sub>4</sub>C<sub>12</sub>] was also confirmed by optical microscopy (Figure S12). The phase contrast observation of the cells was not markedly affected after incubation with NPs[Ni<sub>4</sub>C<sub>12</sub>] without irradiation whereas after irradiation, the cells adopted a round shape with loss of adherence, a sign of cell death. These last results clearly highlight the high potential of NPs[Ni<sub>4</sub>C<sub>12</sub>] to be used as efficient photothermal agents in addition to their photocontrolled drug release ability. The photothermal effect is effective above a concentration threshold of 50 µg/mL of NPs[Ni<sub>4</sub>C<sub>12</sub>], corresponding to a Ni<sub>4</sub>C<sub>12</sub> concentration of 5 µg/mL. Washing of the cell wells, prior to irradiation, have also revealed that no cell death can be induced under laser irradiation, meaning that the nanoparticles are not internalized inside the MDA-MB-231 cells (Figure S13). In this case, the cell death is only due to the presence of the Dox and not to the photothermal effect generated by NPs[Ni<sub>4</sub>C<sub>12</sub>] under laser irradiation. These results highlight that there is no need to internalize the nanoparticles inside the malignant cells to induce strong cell death by photothermal effect, i.e. a close proximity is sufficient.

### ***Conclusions***

We have demonstrated the possibility to co-encapsulate a NIR photothermal dye (Ni<sub>4</sub>C<sub>12</sub>) and an anti-cancer drug (Dox) within nanoparticles based on biocompatible amphiphilic block copolymers (PEG<sub>46</sub>-*b*-PMLABe<sub>77</sub>). We have also proved that NIR irradiation of polymer nanoparticles containing nickel-bis(dithiolene) complexes allows for the fine control release of drugs in solution. The Ni<sub>4</sub>C<sub>12</sub> and Dox loaded NPs have physico-chemical properties close to the ones of empty NPs, Dox loaded NPs or Ni<sub>4</sub>C<sub>12</sub> loaded NPs. The presence of the NIR sensitive dye has an influence on the Dox release upon NIR irradiation of the solution containing the NPs and on-demand drug delivery can also be achieved by applying short intermittent laser irradiation pulses. More importantly, cytotoxicity experiments have shown

that the trapping of an hydrophobic nickel-bis(dithiolene) complex into the hydrophobic inner-core of the NPs does not induce any additional toxicity to the NPs. In addition, above 50  $\mu\text{g/mL}$ , a clear photothermal effect can be observed under laser irradiation, showing that these nickel-bis(dithiolene) complex loaded polymer nanoparticles can be good candidates to develop new agents for photothermal therapy.

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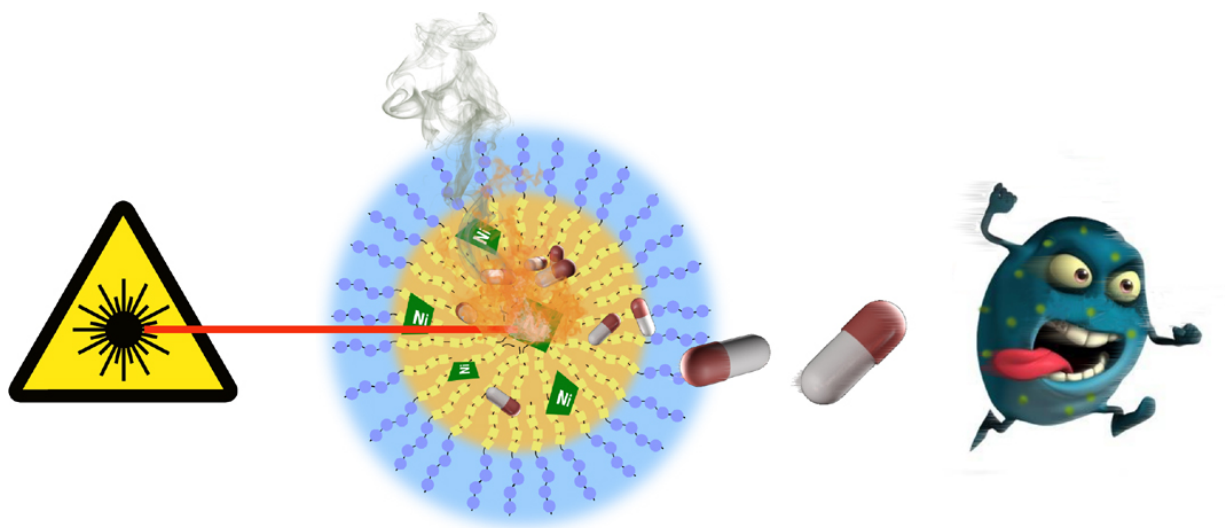
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TOC



Encapsulation of photoactive nickel-bis(dithiolene) complexes into biocompatible and biodegradable block copolymers nanoparticles allows for the fine controlled release of their drug content and photothermal therapy under NIR laser irradiation.