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Postoperative Spinal Implant Infections in Children: Risk Factors, Characteristics, and Outcome

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Running head title: Postoperative Spinal Implant Infections

Conflict of interest: none
**Background:** Postoperative infection is a major complication of spinal surgery with implants. We aimed to identify risk factors for, and characteristics of, postoperative spinal infections in children.

**Methods:** We performed a retrospective observational study of all children who underwent posterior spinal fusion with instrumentation in two referral hospitals in 2008-2013. Spinal infections were defined as local and/or general signs of infection that required surgical treatment in the early postoperative phase (i.e. within 30 days). Data were collected on a standardized questionnaire from medical charts.

**Results:** Of the 450 children who underwent spinal surgery, 26 (5.8%) were diagnosed with early postoperative spinal implant infection, with a median age of 14 years [interquartile range, 13-17], and a median delay of 13 days post-surgery [IQR, 7-18]. Postoperative infection was more common in children with neurologic scoliosis as compared with idiopathic scoliosis (12.2% (15/123) vs 2.4% (5/211); P<0.01). Neurologic scoliosis was an independent predictor of spinal implant infections (HR 3.87 [1.72-8.69]; P<0.001). Main pathogens were *Staphylococcus aureus* (n=14), and *Enterobacteriaceae* (n=8). All children underwent early surgery (wound exploration, debridement, lavage), and antibiotics for a median duration of 19 weeks [IQR, 12-26]. Two children (7.7%) required a second surgery. Spinal implants could be retained in all, and no relapse occurred with a follow-up of ≥24 months after antibiotic discontinuation.

**Conclusion:** Postoperative spinal implant infection is not rare in pediatric patients, especially with neurologic scoliosis. Most children may be cured with implant retention if managed with early surgery followed by a 3-months course of appropriate antibacterial agents.

**Keywords**: spinal implant; post-operative infection; *Staphylococcus aureus*; *Enterobacteriaceae*
INTRODUCTION

Spinal deformity in children may require complex surgery, including posterior spinal fusion with instrumentation. Postoperative infection is a major complication of spinal implants, reported in 0.5-3% of children operated for idiopathic scoliosis, and 3.8-13.3% of children operated for neurologic scoliosis [1-5]. Although device removal would be desirable to reduce bacterial burden and remove the biofilm, this option is not realistic in early postoperative infections, as it threatens spinal stability. Hence, management relies on early surgery with wound exploration, thorough debridement and lavage, followed by prolonged antibacterial treatment [3,6,7]. The optimal duration of antibiotics post-debridement is not defined, and some experts advocate for long-term antimicrobial therapy [8], although a landmark study in adults found that a 3-month course of antibiotics post-debridement cures most spinal implant infections [9]. We aimed to identify risk factors for early postoperative infections after spinal implantation in children, and to characterize these infections, their management, and outcomes.

MATERIALS AND METHODS

Study design

We performed a retrospective, observational study of all children <21-year old who underwent posterior spinal fusion with instrumentation during years 2008-2013 in Rennes and Nantes university hospitals, the referral centers for spinal surgery in children for Western France (population catchment area, 7 million inhabitants). Peri-operative antibiotic prophylaxis was administered in all children, 0-60 minutes before skin incision, with cefamandole from 2008 to 2011, or cefazolin in 2012-2013, 50 mg/kg (max 2 g), and was repeated once 4 hours later if the surgery was still ongoing. After surgery, children were routinely followed-up in their referral center during 24 months. Spinal infections were defined as local and/or general signs of infection that required surgical treatment in the early...
postoperative phase (i.e. within 30 days). No written protocol for spinal implant infections in children was available during the study period, but all cases were managed by a multidisciplinary team including pediatric surgeon(s), pediatrician(s), infectious diseases specialist(s), and microbiologist(s).

**Data collection**

Children who underwent spinal surgery were identified through review of pediatric surgery operation programs, the ‘Q-planner’. Postoperative infections were identified through i) department of infection control computerized database, with prospective collection of all postoperative infections since 2007 (mandatory notification); ii) pediatric surgeons registry. Data were collected by one investigator (AL) on a standardized questionnaire from medical charts, including: demographics, underlying spinal disease, surgery (duration, device implanted, antibiotic prophylaxis), delay between surgery and symptoms onset, between symptoms onset and debridement, microbiology, antibacterial treatment, and outcome. All infected patients were followed-up for ≥24 months after antibiotic discontinuation.

**Statistics**

Continuous variables were expressed as median [interquartile range], and compared using Student t-test or Mann-Whitney U-test. Categorical variables were expressed as numbers (%), and compared using Chi² or Fisher exact tests. P<0.05 was considered statistically significant. For multivariate analysis, all variables with P<0.1 by univariate analysis were evaluated using a stepwise logistic regression model. Logistic regression analyses were performed using SPSS software, version 16.0.

**RESULTS**

From 2008 to 2013, 450 children underwent surgery for spinal deformity with implantation of spinal rods in Rennes (n=258), or Nantes (n=192). Of these, 26 (5.8%; 95% confidence interval, 3.8-8.4%) developed postoperative spinal implant infections. All children with spinal
implant infections had fever (≥38.3°C), and local signs of infection (purulent wound discharge, inflammation, and/or wound dehiscence). Median plasma C-reactive protein concentration was 63 mg/L [IQR 40-194] and median white blood cells count was 10.5 G/L [IQR 8-17]. On univariate analysis, neuromuscular scoliosis and long duration of surgery were associated with spinal implant infection (Table 1). On multivariate analysis, neuromuscular scoliosis was the only predictor of spinal implant infections (HR 3.87 [1.72-8.69]; P<0.001).

All patients underwent posterior spinal fusion with implantation of spinal titanium rods, various implants (screws, hocks and ligaments), and bone grafts. Median time between initial surgery and symptoms of infection was 13 days [IQR, 7-18]. Median time between symptoms onset and debridement was 17 days [IQR, 9-23]. Surgical treatment of postoperative infections consisted of scar incision, wound exploration, debridement and high volume-lavage (3-6 L of sterile water with chlorhexidine 0.05%). Spinal implants were retained in all patients. Multiple samples were immediately sent to the laboratory, following principles applied for prosthetic joint infections: i) ideally, 5 samples from different sites; ii) areas with macroscopic signs of infection to be prioritized; iii) routine cultures to be prolonged 7 days to identify slow-growing organisms, including coagulase-negative staphylococci, and Propionibacterium acnes. Immediately following peroperative sampling, high-doses intravenous anti-staphylococcal therapy was started in all children (vancomycin or cloxacillin, based on risk factors for meticillin-resistant S. aureus), associated with gentamicin or cefotaxime in, respectively, 13 and 12 children.

Microbiologic documentation was obtained in 25 children, including 8 (32%) polymicrobial (Table 2). Main pathogens were Staphylococcus aureus (n=14, all meticillin-susceptible S. aureus, MSSA), and Enterobacteriaceae (n=8, with no isolate resistant to third-generation cephalosporin or fluoroquinolone). Of note, all the patients who developed postoperative
*Enterobacteriaceae* infections (n=7), had neurologic scoliosis. Only two children had positive blood cultures. Twenty-four children underwent oral switch after a median duration of 17 days [IQR, 8-39] with intravenous therapy. Oral therapy was a rifampicin-fluoroquinolone combination in 17 patients (71%). Adverse events requiring treatment alterations were i) rifampicin-related nausea and vomiting (n=3), that resolved after dose reduction; ii) ciprofloxacin-related increase in the incidence of seizures in a child with preexisting convulsive encephalopathy, that required ciprofloxacin discontinuation. Median duration of antibacterial treatment was 19 weeks [IQR, 12-26] overall, but differed between the 2 hospitals, with a median duration of 12 weeks in Rennes [IQR, 6-12], versus 26 weeks in Nantes [IQR, 26-26, P<0.05].

All children were cured, with a follow-up ≥24 months after treatment discontinuation, despite spinal implants were retained in all. Two children with MSSA infections initially underwent partial surgery with limited incision and incomplete wound exploration: Both required one additional surgery with extensive wound exploration, debridement and lavage. Samples taken during the second surgery yielded MSSA again for one child and *P. acnes* for the other. Both were finally cured with an additional 12 weeks of appropriate antibacterial treatment.

**DISCUSSION**

The incidence of postoperative spinal implant infections was 5.8% overall in this cohort, in agreement with other studies [1-10]. The risk was higher in children with neuromuscular scoliosis, as previously reported [2, 4]. In addition, our study suggests for the first time that spinal implant infections in patients with neuromuscular scoliosis is more likely to be related to *Enterobacteriaceae*. Indeed, children with neuromuscular lesions, including paraplegia and anal incontinence, may be at increased risk of scar contamination with digestive microbiota, due to the proximity between the lower extremity of the scar (i.e. lumbar spine) and the anus. This pathway should be considered for the prevention of postoperative infections in children
operated for neuromuscular spinal deformity, with reinforced infection control policy, closer monitoring of surgical scars, and early change of wound dressings in case of potential contamination.

Pathogens responsible for postoperative spinal implant infections in this study are in agreement with previous reports [10], with a predominance of *Staphylococcus* sp., *Enterobacteriaceae*, *P. acnes*, and *P. aeruginosa*. Empirical treatment for postoperative spinal infections in our institutions includes piperacillin/tazobactam and gentamicin, which ensures adequate coverage, pending adjustment based on microbiologic documentation and drug susceptibility testing. Optimal duration of antibacterial treatment for spinal implant infections remains unknown, due to the absence of comparative study in the field, and the lack of validated surrogate marker for cure that could be used to guide treatment discontinuation. Hence, most experts recommend prolonged antibacterial treatment, at least 3-6 months, although shorter treatment duration may be as effective. In adults, Dubée et al. convincingly demonstrated that a 3 month-course of antibacterial treatment is sufficient for most patients with early postoperative spinal implant infections [9]. In children, Ying Li et al. recommend parenteral antibiotics for a minimum of 4-6 weeks after debridement and lavage with implant retention, followed by oral antibiotics for 2-6 months [8]. The French Society of Infectious Diseases (SPILF) recommends 2 weeks of intravenous therapy for osteoarticular infections involving foreign devices, relayed by an oral combination during 6-12 weeks [7]. In our study, antibacterial treatment was not protocolized, but most children received either 6, 12 or 24 weeks of antibiotics following debridement and lavage with retention of spinal implants, with a 100% success rate. Of note, more than two thirds of children received a combination of rifampicin and fluoroquinolone, the preferred regimen for oral step-down in foreign device-related infections when the bacteria involved are susceptible, for the following reasons: i) fluoroquinolones, and rifampicin are among the rare antibiotics with bactericidal activity on
biofilm; ii) both penetrate well in bone tissues; iii) digestive absorption is excellent, so that oral administration is similar to intravenous in terms of plasma concentration. The two children who required one additional surgical treatment for infection were those who underwent only partial surgery initially. These findings suggest that antibacterial treatment can be safely discontinued 3 months after surgical treatment of spinal implant infections, provided wound exploration, debridement and lavage, were rigorously performed. This is in line with current guidelines for the management of prosthetic joint infections in adults, with the ‘Debridement, Antibiotics, and Implant Retention’ (DAIR) approach, with cure rates >90% provided surgical intervention was performed early (i.e. <3 weeks) after infection onset. Although this recommendation is a significant step forward as compared to current recommendations of 4-6 weeks intravenous, followed by 2-6 months oral antibiotics in children [8], this may still be quite conservative, as no failure was reported among the 4 children who received only 6 weeks postoperatively in our series.

This observational study has limitations. Firstly, data were collected retrospectively from medical charts, hence important information may be lost. Secondly, sample size was limited (although among the largest reported to date). Thirdly, management was not protocolized during the study period, with significant heterogeneity between the two sites. However, this study adds significant information on risk factors for, and characteristics of this major postoperative infection, and may contribute to reduce antibacterial treatment duration, given the 100% cure rates observed after adequate debridement and lavage with implant retention, followed by 6-12 weeks of antibiotics.
REFERENCES


7. Société de Pathologie Infectieuse de Langue Française (SPIFLF), Collège des Universitaires de Maladies Infectieuses et Tropicales (CMIT), Groupe de Pathologie Infectieuse Pédiatrique (GPIP), Société Française d’Anesthésie et de Réanimation (SFAR), Société Française de Chirurgie Orthopédique et Traumatologique


Table 1. Risk factors for spinal implant infections: univariate analysis

<table>
<thead>
<tr>
<th>Variable</th>
<th>Total (n=450)</th>
<th>Spinal implant infections (n=26)</th>
<th>No spinal implant infection (n=424)</th>
<th>P</th>
<th>RR (CI95%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, years (mean ± SD)</td>
<td>13.5 ± 3.6</td>
<td>14.8 ± 3.5</td>
<td>13.5 ± 3.6</td>
<td>0.064</td>
<td>NA</td>
</tr>
<tr>
<td>Duration of surgery, minutes (mean ± SD)</td>
<td>248 ± 80.8</td>
<td>281 ± 58.1</td>
<td>246 ± 81.6</td>
<td>0.031</td>
<td>NA</td>
</tr>
<tr>
<td>Male sex, n (%)</td>
<td>144 (32)</td>
<td>10 (38.5)</td>
<td>134 (31.6)</td>
<td>0.47</td>
<td>1.33 (0.62-2.85)</td>
</tr>
<tr>
<td>Etiology of scoliosis, n (%)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Idiopathic</td>
<td>211 (46.9)</td>
<td>5 (19.2)</td>
<td>206 (48.6)</td>
<td>&lt;0.01</td>
<td>0.27 (0.10-0.70)</td>
</tr>
<tr>
<td>Neuromuscular</td>
<td>123 (27.3)</td>
<td>15 (57.7)</td>
<td>108 (25.5)</td>
<td>&lt;0.01</td>
<td>3.57 (1.71-7.67)</td>
</tr>
<tr>
<td>Congenital</td>
<td>75 (16.7)</td>
<td>4 (15.4)</td>
<td>71 (16.75)</td>
<td>0.86</td>
<td>0.91 (0.32-2.56)</td>
</tr>
<tr>
<td>Others *</td>
<td>41 (9.1)</td>
<td>2 (7.7)</td>
<td>39 (9.2)</td>
<td>0.80</td>
<td>0.83 (0.20-3.39)</td>
</tr>
</tbody>
</table>

SD, standard deviation; NA, Non applicable

* Spondylolisthesis, trauma, cancer, benign tumor, miscellaneous genetic diseases
Table 2. Microbiology of spinal implant infections (n=26)

<table>
<thead>
<tr>
<th>Microbiology</th>
<th>Number of isolates</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Staphylococcus aureus</strong></td>
<td>14</td>
</tr>
<tr>
<td>Meticillin-resistant</td>
<td>0</td>
</tr>
<tr>
<td>Coagulase-negative staphylococci</td>
<td>4</td>
</tr>
<tr>
<td>Meticillin-resistant</td>
<td>3</td>
</tr>
<tr>
<td><strong>Enterococcus faecalis</strong></td>
<td>1</td>
</tr>
<tr>
<td><strong>Enterobacteriaceae</strong></td>
<td>8</td>
</tr>
<tr>
<td><strong>Escherichia coli</strong></td>
<td>3</td>
</tr>
<tr>
<td><strong>Klebsiella pneumoniae</strong></td>
<td>2</td>
</tr>
<tr>
<td><strong>Proteus mirabilis</strong></td>
<td>2</td>
</tr>
<tr>
<td><strong>Serratia marcescens</strong></td>
<td>1</td>
</tr>
<tr>
<td><strong>Pseudomonas aeruginosa</strong></td>
<td>2</td>
</tr>
<tr>
<td>Ciprofloxacin-resistant</td>
<td>0</td>
</tr>
<tr>
<td><strong>Propionibacterium acnes</strong></td>
<td>4</td>
</tr>
<tr>
<td>Polymicrobial infection *</td>
<td>8</td>
</tr>
</tbody>
</table>

*K. pneumoniae* and *P. aeruginosa; Granulicatella adiacens* and *Micromonas micros; E. faecalis* and meticillin-susceptible *S. aureus; P. acnes* and meticillin-susceptible *S. aureus; K. pneumoniae* and *P. acnes* and meticillin-resistant *S. epidermidis; Providencia stuartii* and meticillin-susceptible *S. epidermidis; P. acnes* and *Finegoldia magna; P. mirabilis* and *E. coli* and meticillin-susceptible *S. aureus;*