



# Endocrine Disrupting-Chemicals and Biochemical Recurrence of Prostate Cancer after Prostatectomy A cohort study in Guadeloupe (French West Indies)

Laurent Brureau, Elise Emeville, Carole Helissey, Jean Pierre Thome, Luc  
Multigner, Pascal Blanchet

## ► To cite this version:

Laurent Brureau, Elise Emeville, Carole Helissey, Jean Pierre Thome, Luc Multigner, et al.. Endocrine Disrupting-Chemicals and Biochemical Recurrence of Prostate Cancer after Prostatectomy A cohort study in Guadeloupe (French West Indies). International Journal of Cancer, 2020, 146 (3), pp.657-663. 10.1002/ijc.32287 . hal-02091212

**HAL Id: hal-02091212**

**<https://univ-rennes.hal.science/hal-02091212>**

Submitted on 11 Apr 2019

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

# Endocrine Disrupting-Chemicals and Biochemical Recurrence of Prostate Cancer after Prostatectomy: A cohort study in Guadeloupe (French West Indies)

Laurent BRUREAU <sup>a,\*</sup>, Elise EMEVILLE <sup>b,\*</sup>, Carole HELISSEY <sup>c</sup>, Jean Pierre THOME <sup>d</sup>,  
Luc MULTIGNER <sup>b,\*\*</sup>, Pascal BLANCHET <sup>a</sup>

\* Equal contribution

## Affiliations

<sup>a</sup> CHU de Pointe-à-Pitre, Univ Antilles, Univ Rennes, Inserm, EHESP, Irset (Institut de recherche en santé, environnement et travail) - UMR\_S 1085, F-97110 Pointe-à-Pitre, France.

<sup>b</sup> Univ Rennes, Inserm, EHESP, Irset (Institut de recherche en santé, environnement et travail) - UMR\_S 1085, F-35000 Rennes, France.

<sup>c</sup> Département d'Oncologie Médicale, Unité de Recherche Clinique, Hôpital d'Instruction Militaire Begin, F-94160 Saint Mandé, France.

<sup>d</sup> Université de Liège, LEAE-CART (Laboratoire d'Ecologie Animale et d'Ecotoxicologie-Centre de Recherche Analytique et Technologique), B- 4000 Liège, Belgium.

**\*\* Corresponding author:** Luc Multigner, Inserm U1085-IRSET, 9 Avenue du Professeur Léon Bernard, 35000 Rennes, France. Telephone: + 33 223 59 28 28. E-mail: luc.multigner@inserm.fr

**Keywords:** Biochemical recurrence; Endocrine-disrupting chemicals; Prostate cancer

**Abbreviations:** DDE, *p,p'*-dichlorodipenyldichloroethylene ; PCB-153, polychlorinated biphenyl congener 153; EDCs, endocrine-disrupting chemicals ; POPs, persistent organic

pollutants ; LOD, limit of detection; HR, hazard ratio; CI, confidence interval ; PSA, prostate-specific antigen.

## Research Article

**Novelty and Impact:** Environmental chemicals with hormonal properties, also called endocrine-disrupting chemicals are suspected to favor the occurrence of prostate cancer. These chemicals are obviously still present at the beginning of therapy, and beyond, and consequently they could influence the treatment outcome. Here we report for the first time that exposure to the estrogenic insecticide chlordecone, determined at the time of diagnosis, increases the risk of biochemical recurrence after radical prostatectomy of organ-confined disease.

## Abstract

Previous studies have suggested that exposure to environmental chemicals with hormonal properties, also called endocrine disrupting chemicals, may be involved in the occurrence of prostate cancer. Such exposure may also influence the treatment outcome as it is still present at the time of diagnosis, the beginning of therapy, and beyond. We followed 326 men in Guadeloupe (French West Indies) who underwent radical prostatectomy as primary treatment of localized prostate cancer. We analyzed the relationship between exposure to the estrogenic chlordecone, the anti-androgenic dichlorodiphenyldichloroethylene (DDE, the main metabolite of the insecticide DDT), and the non-dioxin-like polychlorinated biphenyl congener 153 (PCB-153) with mixed estrogenic/anti-estrogenic properties and the risk of biochemical recurrence after surgery. After a median follow-up of 6.1 years after surgery, we found a significant increase in the risk of biochemical recurrence, with increasing plasma chlordecone concentration (adjusted hazard ratio = 2.51; 95% Confidence Interval: 1.39-4.56 for the highest versus lowest quartile of exposure;  $p$  trend = 0.002). We found no associations for DDE or PCB-135. These results shown that exposure to environmental estrogens may negatively influence the outcome of prostate cancer treatment.

## Introduction

Prostate cancer is the most common hormone-sensitive malignancy in men.<sup>1</sup> Endogenous steroid sex hormones play a key role in the initiation and progression of this disease.<sup>2</sup>

*In vitro* and *in vivo* experimental research has increasingly suggested that environmental chemicals with hormonal properties, also called endocrine-disrupting chemicals (EDCs), particularly those that mimic steroid sex hormones, may modulate the risk of developing prostate cancer.<sup>3-5</sup> Among these, persistent organic pollutants (POPs) have attracted attention because of their long-term presence in the environment and for some of them their ability to interfere with steroid sex hormone-regulated processes.<sup>6-10</sup>

Several epidemiological studies using exposure biomarkers have been conducted to investigate the relationship between human exposure to POPs with hormonal properties during adulthood and the risk of prostate cancer.<sup>11-19</sup> Conclusions have been divergent, sometimes even demonstrating associations in opposite directions, depending on the chemical considered or the aggressiveness of the disease at the time of diagnosis. Such disparities may be explained by the different hormonal properties of these compounds, agonistic or antagonistic, on different members of the steroid sex receptor superfamily, the clinical and molecular heterogeneity of the disease, or differences in exposure windows.

The influence of human exposure to EDCs on the development of prostate cancer is still an open question. Nonetheless, patients are still exposed at the time of diagnosis and subsequent treatment. This raises the question of the influence of EDCs on disease progression after treatment. Surgical resection of the entire gland, radical prostatectomy, is a common therapeutic option, for curative purposes, for men with organ confined prostate

cancer. However, approximately 20-40% of patients will present biochemical recurrence (rising prostate-specific antigen, PSA) of the disease. Many studies, have identified clinical and pathological features to be significant risk factors of biochemical recurrence<sup>20-22</sup> but nothing is known about the influence of environmental chemicals

We sought to analyze the association between blood concentrations of various POPs at the time of diagnosis and the risk of biochemical recurrence after radical prostatectomy, a common surgical procedure for the complete removal of the prostate for men with clinically organ-confined prostate cancer.

Among POPs, we have focused our attention on the most prevalent ones such as the estrogenic insecticide chlordecone, the anti-androgenic *p*, *p'*-dichlorodiphenyldichloroethylene (DDE, the main and most stable metabolite of the insecticide *p*, *p'*-dichlorodiphenyltrichloroethane, DDT), and the non-dioxin-like polychlorinated biphenyl congener 153 (PCB-153), which displays both estrogenic and anti-estrogenic properties.

## **Material and Methods**

### ***Study population***

This study took place in Guadeloupe (French West Indies), a Caribbean archipelago, where most of the inhabitants are of African descent. It was prospectively conducted on incident prostate cancer patients who initially participated in a population-based case-control study between June 2004 and December 2007,<sup>14</sup> subsequently underwent radical prostatectomy in one single center (Urology Department of the University Hospital of Guadeloupe) (n = 392), and from whom we were able to obtain plasma samples and determine the POPs

Accepted Article

concentrations one to three months before surgery (n = 340). All patients were followed by serial PSA determinations and clinical visits every six months for the first three years after surgery and annually thereafter. Biochemical recurrence was defined as two consecutive (usually four weeks apart) PSA measurements above 0.2 ng/ml, with the date of the first increase after nadir as the recurrence date.<sup>23</sup> Fourteen patients were excluded because they were treated with neoadjuvant hormonal therapy or radiotherapy or because the PSA value did not return below 0.2 ng/ml six weeks after surgery. The study was approved by the Guadeloupean ethics committee for studies involving human subjects. Each participant received, completed, and signed written informed consent.

Before surgery, we collected information on the demographic characteristics, Caribbean origin (French West Indies, Haiti or Dominica), education (primary, secondary, high school or higher), weight and height allowing the calculation of body mass index (kilograms per meter squared), smoking (never, former or current), alcohol consumption (never, former, or current), diabetes type 2 (no, yes), and family history of prostate cancer (first degree relatives: no, yes, not known), and preoperative PSA level. After surgery, we collected data on the pathological stage, prostatectomy ISUP Gleason grade,<sup>24</sup> surgical margins, tumour density, and follow-up PSA levels.

#### ***Laboratory Assays.***

A high-resolution gas chromatograph (Thermo Quest Trace 2000, Milan, Italy) equipped with a Ni63 electron capture detection system was used to determine the blood concentration of chlordecone, *p,p'*-DDT, *p,p'*-DDD (dichlorodiphenyldichloroethane), *p,p'*-DDE, the  $\alpha$ ,  $\beta$ , and  $\gamma$  isomers of hexa-chlorocyclohexane (HCH), and 24 PCB congeners (International Union of

Pure and Applied Chemistry number): 6 dioxin-like (77, 105, 118, 126, 156, and 169) and 18 non-dioxin-like (18, 28, 52, 101, 110, 128, 138, 143, 149, 153, 170, 180, 183, 187, 194, 195, 206, and 209). The limit of detection (LOD) was 0.05 µg/L for all POPs except 0.06 µg/L for chlordécone (0.06 µg/L).<sup>12</sup>. Detailed information about sampling, analysis, and quality assurance and control has been provided elsewhere.<sup>14, 25</sup> Plasma total cholesterol and total triglyceride concentrations were determined enzymatically (DiaSys Diagnostic Systems GmbH; Holzheim, Germany) and total lipid concentration was calculated as previously described.<sup>26</sup>

### *Statistical Analysis.*

We restricted our analysis to chemicals detected at a rate of more than 80% (DDE; PCB congeners 138, 153, and 180; and chlordecone). Correlations between concentrations of the frequently detected PCBs were explored by Spearman's rank correlation analysis (Supporting information, supplemental Table 1). The concentrations of the various PCBs were highly correlated (Spearman's  $\rho \geq 0.75$ ; all  $p$ -values  $< 0.001$ ), so we restricted further analysis to PCB-153.

Plasma POPs concentrations were categorized into quartiles, based on their distribution among patients. Time to event was defined as the duration between the date of surgery and the PSA value that defined the recurrence event. Patients who did not relapse, or were lost to follow-up, were censored at the last normal post-operative PSA measurement before October 30, 2017. Five-year BCR-free survival analysis were carried out using the Kaplan–Meier method and log-rank test for trend. The Cox proportional hazards method was used to compute adjusted survival curves and test for trend were assessed with quartiles of



exposure scored as ordinal variables in ascending order. We also used multivariable Cox proportional hazards regression models and 95% confidence intervals (CI) to estimate the hazard ratio (HR) of biochemical recurrence associated with POPs exposure. Potential confounders were included in all statistical models if they predicted ( $p < 0.05$ ) biochemical recurrence status: body mass index, family history of prostate cancer, pathological Gleason grade, pathological stage, surgical margins, tumor density (Table 1) and then selected by applying a backward selection procedure at  $p < 0.1$ . Missing data for covariates varied from none to two (0.6 %) for education, two (0.6 %) for alcohol, two (0.6 %) for family history of prostate cancer, three (0.9 %) for smoking, and eight (2.5 %) for diabetes. Missing data were handled using missing value indicator categories. The proportional hazards assumption was verified by the log-negative-log survival distribution function of all variables. Tests for linear trend across exposure categories were performed, with the natural log transformed POPs concentration treated as a continuous variable. Exposure levels equal to or below the LOD were estimated by a maximum likelihood estimation method.<sup>27</sup> Sensitivity analyses were conducted by excluding patients with positive surgical margins or prostatectomy ISUP Gleason grade 3 or higher, or advanced pathological stage (pT3 or pT4 or N+). Statistical analyses were carried out using SAS software version 9.3 (SAS Institute, Cary, NC, USA). All tests were two-sided, and  $p < 0.05$  were considered statistically significant.

## Results

The results presented here were obtained from a study population of 326 patients. During 1,923 person-years of follow-up after surgery, 93 men (28.5%) experienced biochemical recurrence. The median follow-up time was 6.1 years and among men without progression 7.3

years. The baseline characteristics of the study population and HR for risk of biochemical recurrence are presented in Table 1. Detection levels and plasma concentrations of POPs are presented in Table 2.

The crude five-year BCR-free survival rates were 83.8%, 79.8%, 86.6%, and 65% for men in the 1<sup>st</sup>, 2<sup>nd</sup>, 3<sup>rd</sup>, and 4<sup>th</sup> quartile of chlordecone exposure, respectively (Log-rank test for trend,  $p = 0.01$ ). For DDE and PCB-153 exposure, the five-year biochemical recurrence-free survival rates were 78.7, 83.8, 70.7, and 81.1 and 81.5, 79.0, 79.7, and 75.0 for men in the 1<sup>st</sup> to 4<sup>th</sup> quartile, respectively (DDE: Log-rank test for trend,  $p = 0.85$ ; PCB-153: Log-rank test for trend,  $p = 0.83$ ). Crude and adjusted survival curves are presented in supporting information (supplemental figures 1 to 3).

Adjusted Cox analysis showed that men in the highest quartile of chlordecone plasma concentration had a significantly higher risk of biochemical recurrence than those in the lowest quartile (HR= 2.51, 95% CI: 1.39-4.56) (Table 3). The linear relationship between exposure and response was significant ( $p$ -trend = 0.002). We obtained comparable results if DDE and PCB-153 concentrations were additionally included in the multivariable model (HR= 2.62, 95% CI: 1.40-4.90 for the highest quartile relative to the lowest quartile;  $p$ -trend = 0.002). DDE and PCB-153 exposure were not associated with a risk of BCR, irrespective of the model (Table 3).

Sensitivity analysis revealed that chlordecone exposure was still significantly associated with a risk of BCR after excluding patients with positive surgical margins or prostatectomy ISUP Gleason grade 3 or higher, or advanced pathological stage (Table 4). Men who were in the highest quartile of chlordecone plasma concentration had a higher risk

of BCR than those in the lowest quartile (Adjusted HR = 2.98, 95% CI: 1.06-8.38) and the linear relationship between exposure and response was significant ( $p$ -trend = 0.003). Here again, DDE and PCB-153 exposure were not associated with a risk of BCR (Table 4).

## Discussion

This is the first study to investigate the association of POPs plasma concentrations, measured at the time of diagnosis, with biochemical recurrence following primary curative surgical treatment of prostate cancer.

In our study population, the highest quartile of exposure to chlordane was consistently associated with a more than two-fold increased risk of biochemical recurrence, with a significant exposure-response relationship. This was still true after taking into account well-known prognostic clinic-pathological features of biochemical recurrence as confounding factors and in sensitivity analysis restricted to patients who did not present these adverse clinic-pathological features. In contrast, exposure to DDE or PCB-153 was not associated with BCR.

Chlordane was extensively used from 1973 to 1993 in the French West Indies, to control the banana root borer. This pesticide undergoes no significant biotic or abiotic degradation in the environment, so permanently polluted soils and waters have remained the primary source of foodstuffs contamination, and human beings continue to be exposed to this chemical<sup>28</sup>. In our study population, chlordane plasma concentration was in the range of values observed for residents in the neighborhoods near the chlordane factory in Hopewell (VA, U.S.).<sup>29</sup> Such exposure resulted from the air pollution and illegal dumping of waste

laden into Hopewell's sewer system which in turn contaminated the James River and fish resources. There is currently no data on exposure to chlordecone in other populations. Concerning other POPs, DDE and PCB congeners 138, 153, and 180 were the most prevalent found in the blood in our population, as expected and found in most populations worldwide. Moreover, blood concentrations of these pollutants are in the range of background environmental levels currently found in the US population of similar age range<sup>30</sup>. This is not surprising, because the use of DDT in agricultural supplies or for disease vector control in the French West Indies was sporadic and industrial activities involving the significant use or emission of PCBs has only been very limited. Consequently, exposure to these chemicals is likely to be associated with background contamination of the food chain.

Biomonitoring of human tissues is considered to be a gold standard approach for assessing aggregated environmental chemical exposure, because it integrates all sources of exposure from different absorption pathways.<sup>31</sup> The exposure to POPs observed in this study was assessed based on plasma concentrations at the time of diagnosis, prior to surgery, and before the outcome occurrence. These chemicals have a long half-life in the body, varying from six months for chlordecone to several years for DDE and PCB-153. Although the use of POPs is mostly banned worldwide, they are still present in the environment and populations are continuously exposed through food consumption. Thus, a single measurement of POPs concentration in the blood reflects the body burden under steady-state conditions and provides a sufficiently confident estimation of exposure over an extended period. However, the levels of each POPs varies depending on the type of foodstuffs (*i.e.* chlordecone is preferentially found in root vegetables, fish, and meat of local origin, DDE in food crops, fish and meat, and

PCB in meat and milk). Thus, any significant changes in dietary habits may result in changes in the body burden over several years. However, no specific dietary recommendations have been made and we are not aware of any changes in the dietary habits of our patients after surgery.

Although radical prostatectomy is considered as a definitive therapy for organ-confined prostate cancer, an increase in PSA levels after a variable period of undetectable or low detectable-stable PSA indicates the presence of residual prostatic cells in the body, which may be benign or malignant and in the surgical bed or disseminated. We do not know whether the positive association between chlordecone exposure and biochemical recurrence reflects a direct effect on the PSA production of residual cells or the proliferation of these cells, which in turn secrete PSA. PSA secretion is generally under androgen regulation, but chlordecone shows low affinity binding to the androgen receptor (AR) and acts as an AR antagonist rather than agonist.<sup>8</sup> Moreover, there is currently no data supporting the involvement of chlordecone in AR-dependent or AR-independent mechanisms driving PSA mRNA expression or protein secretion.

We previously reported that chlordecone exposure was associated with an increased risk of prostate cancer occurrence and suggested that the estrogenic properties of the molecule may explain this association.<sup>14</sup> Chlordecone interferes with estradiol signaling through binding to the nuclear estrogen receptors  $\alpha$  (ER $\alpha$ ) and  $\beta$  (ER $\beta$ ), eliciting agonistic and antagonistic effects, respectively.<sup>7,32,33</sup> The ER $\alpha$  mediates the adverse effects of estrogen, such as aberrant proliferation, inflammation, and malignancy, whereas the ER $\beta$  exerts opposite and beneficial effects, such as anti-proliferative, pro-apoptotic, anti-inflammatory,

and, potentially, anticarcinogenic effects.<sup>34</sup> Thus, the combined interaction between the agonistic effects of chlordecone on the ER $\alpha$  and the antagonistic effects on the ER $\beta$  could lead to an imbalance, promoting cell proliferation. It is possible that such interplay could also play a role in the proliferation of residual prostatic tumor cells, if present, after radical prostatectomy.

Although biochemical recurrence is never followed by disease progression for some patients, it is almost certainly an indicator of persistent disease and an unquestionable risk factor for the development of subsequent distant metastases. Indeed, chlordecone has been found to promote angiogenesis through ER $\alpha$  activation,<sup>35</sup> and both chlordecone and estradiol increase expression of the homeostatic chemokine receptor type 4 and enhance vascular cell adhesion protein 1 levels.<sup>36</sup> Moreover, chlordecone disrupts cell-adherence junctions by modulating E-cadherin and  $\beta$ -catenin expression.<sup>37</sup> Collectively, these effects participate in the tumor cell-microenvironment signaling involved in tumor and metastasis progression for several types of cancers, including prostate cancer.<sup>38-42</sup> Whether biochemical recurrence associated with chlordecone exposure is predictive of an increased risk of metastasis remains to be explored through the long-term follow up of patients.

Several epidemiological studies in different locations, including in French West Indies, have suggested that exposure to DDE is associated with a higher risk of prostate cancer occurrence.<sup>12,15,19</sup> However, we did not find a significant association between DDE exposure and the risk of biochemical recurrence. DDE is known to inhibit AR-mediated effects through direct binding to the AR.<sup>8</sup> A recent study has also shown that DDE lowers PSA mRNA and protein levels in human prostate cancer cell lines in a dose-dependent

Accepted Article

manner.<sup>43</sup> DDE exposure of healthy men is negatively associated with serum concentration of 5 $\alpha$ -dihydrotestosterone,<sup>44</sup> the most potent ligand and activator of the AR. Given these properties, DDE would have been expected to be associated with a decreased risk of biochemical recurrence due to a possible direct effect on PSA production or prostate cell growth, itself an AR-mediated effect. DDE may also have an agonistic effect on ER $\alpha$ .<sup>45</sup> It is therefore difficult to predict the net effect of DDE on the prostate, given the potential effects on both the AR and ER $\alpha$ .<sup>46, 47</sup>

The literature concerning PCBs is less clear, as there are contradictory and even opposite observations regarding their association with the risk of PCa occurrence.<sup>11-13,16-18</sup> Here, we did not observe a significant association between PCB-153 exposure and the risk of biochemical recurrence, in either direction. Unlike dioxin-like PCBs, non-dioxin-like PCBs (including PCB-153), which are the most highly prevalent PCBs in the environment,<sup>48</sup> do not interact substantially with the aryl hydrocarbon receptor and may act through other pathways, such as steroid hormone signaling.<sup>6</sup> Non-dioxin-like PCBs may interact with genomic and non-genomic ER pathways and exhibit both estrogenic and anti-estrogenic properties, depending on the *in vivo* or *in vitro* experimental models.<sup>6,9,10</sup> The multiple potential steroidal mechanisms of non-dioxin-like PCBs are yet to be totally deciphered and it is thus difficult to have a clear picture of their actions.

In conclusions, our results shown that exposure to chlordecone, an environmental EDCs with estrogenic properties, at the time of prostate cancer diagnosis is significantly associated with a risk of biochemical recurrence after prostatectomy. Increasing evidence suggests that estrogens are critical players in human prostate cancer.<sup>49,50</sup> Our findings add

support to the hypothesis that human exposure to environmental estrogens may influence prostate cancer progression

**Acknowledgments:** This work was supported by the French National Health Directorate. E Emeville was supported by a PhD fellowship from the Ligue Nationale Contre le Cancer. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. The authors declare they have no actual or potential competing financial interest related to this work.

## References

1. Center MM, Jemal A, Lortet-Tieulent J, Ward E, Ferlay J, Brawley O, Bray F. International variation in prostate cancer incidence and mortality rates. *Eur Urol.* 2012; 61:1079-92
2. Bosland MC. The role of steroid hormones in prostate carcinogenesis. *J Natl Cancer Inst Monogr.* 2000; 27:39-66
3. Prins GS. Endocrine disruptors and prostate cancer risk. *Endocr Relat Cancer* 2008;15:649-56.
4. Soto AM, Sonnenschein C. Environmental causes of cancer: endocrine disruptors as carcinogens. *Nat Rev Endocrinol.* 2010; 6:363-70.



5. Sweeney MF, Hasan N, Soto AM, Sonnenschein C. Environmental endocrine disruptors: Effects on the human male reproductive system. *Rev Endocr Metab Disord*. 2015; 16:341-57.
6. Cooke PS, Sato T, Buchanan DL. Disruption of steroid hormone signaling by PCBs. In: recent Advances in Environmental Toxicology and Health effects (Robertson LW, Hansen LG, eds). Lexington, KY: University press of Kentucky, 2001; 257-63.
7. Hammond B, Katzenellenbogen, Krauthammer N, McConnell J. Estrogenic activity of the insecticide chlordecone (Kepone) and interaction with uterine estrogen receptors. *Proc Natl Acad Sci USA* 1979; 76:6641-59.
8. Kelce WR, Stone CR, Laws SC, Gray LE, Kemppainen JA, Wilson EM. Persistent DDT metabolite pp'-DDE is a potent androgen antagonist. *Nature* 1995; 376:581-5.
9. Oh SM, Ryu BT, Lee SK, Chung KH. Antiestrogenic potentials of ortho-PCB congeners by single or complex exposure. *Arch Pharm Res*. 2007; 30:199–209.
10. Plísková M, Vondráček J, Canton RF, Nera J, Kocan A, Petrík J, Trnovec T, Sanderson T, van den Berg M, Machala M. Impact of polychlorinated biphenyls contamination on estrogenic activity in human male serum. *Environ Health Perspect*. 2005; 113:1277–84.
11. Aronson KJ, Wilson JW, Hamel M, Diarsvitri W, Fan W, Woolcott C Heaton JP, Nickel JC, Macneily A, Morales A. Plasma organochlorine levels and prostate cancer risk. *J Expo Sci Environ Epidemiol*. 2010; 20:434-45.
12. Emeville E, Giusti A, Coumoul X, Thomé JP, Blanchet P, Multigner L. Associations of plasma concentrations of dichlorodiphenyldichloroethylene and polychlorinated biphenyls

with prostate cancer: a case-control study in Guadeloupe (French West Indies). *Environ Health Perspect.* 2015; 123:317-23.

13. Koutros S, Langseth H, Grimsrud TK, Barr DB, Vermeulen R, Portengen L, Wacholder S, Freeman LE, Blair A, Hayes RB, Rothman N, Engel LS. Prediagnostic Serum Organochlorine Concentrations and Metastatic Prostate Cancer: A Nested Case-Control Study in the Norwegian Janus Serum Bank Cohort. *Environ Health Perspect.* 2015; 123:867-72.

14. Multigner L, Ndong JR, Giusti A, Romana M, Delacroix-Maillard H, Cordier S, Jegou B, Thome JP, Blanchet P. Chlordecone exposure and risk of prostate cancer. *J Clin Oncol.* 2010; 28:3457-62.

15. Pi N, Chia SE, Ong CN, Kelly BC. Associations of serum organohalogen levels and prostate cancer risk: Results from a case-control study in Singapore. *Chemosphere* 2016; 144: 1505-12.

16. Ritchie JM, Vial SL, Fuortes LJ, Guo H, Reedy VE, Smith EM. Organochlorines and risk of prostate cancer. *J Occup Environ Med.* 2003; 45:692-02.

17. Ritchie JM, Vial SL, Fuortes LJ, Robertson LW, Guo H, Reedy V, Smith EM. Comparison of proposed frameworks for grouping polychlorinated biphenyl congener data applied to a case-control pilot study of prostate cancer. *Environ Res.* 2005; 98:104-13.

18. Sawada N, Iwasaki M, Inoue M, Itoh H, Sasazuki S, Yamaji T, Shimazu T, Tsugane S. Plasma organochlorines and subsequent risk of prostate cancer in Japanese men: a nested case-control study. *Environ Health Perspect.* 2010; 118:659-65.

19. Xu X, Dailey AB, Talbott EO, Ilacqua VA, Kearney G, Asal NR. Associations of serum concentrations of organochlorine pesticides with breast cancer and prostate cancer in U.S. adults. *Environ Health Perspect.* 2010; 118:60-6.
20. Srigley JR, Amin M, Boccon-Gibod L, Egevad L, Epstein JI, Humphrey PA, Mikuz G, Newling D, Nilsson S, Sakr W, Wheeler TM, Montironi R. Prognostic and predictive factors in prostate cancer: historical perspectives and recent international consensus initiatives. *Scand J Urol Nephrol Suppl.* 2005; 216:8-19.
21. Punnen S, Freedland SJ, Presti JC Jr, Aronson WJ, Terris MK, Kane CJ, Amling CL, Carroll PR, Cooperberg MR. Multi-institutional validation of the CAPRA-S score to predict disease recurrence and mortality after radical prostatectomy. *Eur Urol.* 2014; 65:1171-1177.
22. Brureau L, Emeville E, Multigner L, Blanchet P. Predictors of biochemical recurrence after radical prostatectomy in an Afro-Caribbean population in Guadeloupe (French West Indies). *Prog Urol.* 2018 ; 28:442-449
23. Heidenreich A, Bastian PJ, Bellmunt J, Joniau S, Mason M, Matveev V, Mottet N, Schmid HP, van der Kwast T, Wiegel T, Zatton. EAU Guidelines on Prostate Cancer. Part 1: Screening, Diagnosis, and Treatment of Clinically Localised Disease. *Eur. Urol.* 2014; 59: 61-71.
24. WHO. Classification of Tumours. Pathology and Genetics of Tumours of the Urinary System and Male Genital Organs. Moch H, Humphrey PA, Ulbright TM, Reuter VE (Eds). IARC Press: 4th Edition, 2016, Volume 8.

25. Debier C, Pomeroy PP, Dupont C, Joiris C, Comblin V, Le Boulenge E, Larondelle Y, Thome JP. Dynamics of PCB transfer from mother to pup during lactation in UK grey seals *Halichoerus grypus*: differences in PCB profile between compartments of transfer and changes during the lactation period. *Mar Ecol Prog Ser*. 2003; 247: 237-48.
26. Bernert JT, Turner WE, Patterson DG Jr, Needham LL. Calculation of serum “total lipid” concentrations for the adjustment of persistent organohalogen toxicant measurements in human samples. *Chemosphere* 2007; 68: 824–31
27. Jin Y, Hein MJ, Deddens JA, Hines CJ. Analysis of lognormally distributed exposure data with repeated measures and values below the limit of detection using SAS. *Ann Occup Hyg*. 2011; 55: 97–112.
28. Multigner L, Kadhel P, Rouget F, Blanchet P, Cordier S. Chlordecone exposure and adverse effects in French West Indies populations. *Environ Sci Pollut Res Int*. 2016; 23:3-8
29. Cannon SB, Veazey JM Jr, Jackson RS, Burse VW, Hayes C, Straub WE, Landrigan PJ, Liddle JA. Epidemic kepone poisoning in chemical workers. *Am J Epidemiol*. 1978; 107:529-537
30. Centers for Disease Control and Prevention. 2009. Fourth National Report on Human Exposure to Environmental Chemicals. Updated Tables. Atlanta, GA:U.S. Department of Health and Human Services, Centers for Disease Control and Prevention. Available: [https://www.cdc.gov/biomonitoring/pdf/fourthreport\\_updatedtables\\_volume1\\_jan2017.pdf](https://www.cdc.gov/biomonitoring/pdf/fourthreport_updatedtables_volume1_jan2017.pdf) [accessed 24 February 2019].

31. Pirkle JL, Sampson EJ, Needham LL, Patterson DG, Ashley DL. Using biological monitoring to assess human exposure to priority toxicants. *Environ Health Perspect.* 1995; 103: S3, 45-8.
32. Kuiper GG, Lemmen JG, Carlsson B, Corton JC, Safe SH, van der Saag PT, van der Burg B, Gustafsson JA. Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor beta. *Endocrinology* 1998; 139:4252-63.
33. Lemaire G, Mnif W, Mauvais P, Balaguer P, Rahmani R. Activation of alpha- and beta-estrogen receptors by persistent pesticides in reporter cell lines. *Life Sci.* 2006; 79: 1160-9.
34. Ellem SJ, Risbridger GP. The dual, opposing roles of estrogen in the prostate. *Ann N Y Acad Sci.* 2009; 1155:174-86.
35. Clere N, Lauret E, Malthiery Y, Andriantsitohaina R, Faure S. Estrogen receptor alpha as a key target of organochlorines to promote angiogenesis. *Angiogenesis* 2012; 15:745-60.
36. Wang F, Roberts SM, Butfiloski EJ, Morel L, Sobel ES. Acceleration of autoimmunity by organochlorine pesticides: a comparison of splenic B-cell effects of chlordane and estradiol in (NZBxNZW)F1 mice. *Toxicol Sci.* 2007; 99:141-52.
37. Starcevic SL, Bortolin S, Woodcroft KJ, Novak RF. Kepone (chlordane) disrupts adherens junctions in human breast epithelial cells cultured on matrigel. *In Vivo* 2001; 15: 289-94.
38. Bonkhoff H. Estrogen receptor signaling in prostate cancer: Implications for carcinogenesis and tumor progression. *Oncotarget* 2018; 78:2-10.

39. Schlesinger M, Bendas G. Vascular cell adhesion molecule-1 (VCAM-1)-an increasing insight into its role in tumorigenicity and metastasis. *Int J Cancer*. 2015; 136:2504-14.
40. Sun X, Cheng G, Hao M, Zheng J, Zhou X, Zhang J, Taichman RS, Pienta KJ, Wang J. CXCL12 / CXCR4 / CXCR7 chemokine axis and cancer progression. *Cancer Metastasis Rev*. 2010; 29:709-22.
41. Tai HC, Chang AC, Yu HJ, Huang CY, Tsai YC, Lai YW, Sun HL, Tang CH, Wang SW. Osteoblast-derived WNT-induced secreted protein 1 increases VCAM-1 expression and enhances prostate cancer metastasis by down-regulating miR-126. *Oncotarget* 2014; 5:7589-98.
42. Taichman RS, Cooper C, Keller ET, Pienta KJ, Taichman NS, McCauley LK. Use of the stromal cell-derived factor-1/CXCR4 pathway in prostate cancer metastasis to bone. *Cancer Res*. 2002; 62: 1832–7.
43. Wong LI, Labrecque MP, Ibuki N, Cox ME, Elliott JE, Beischlag TV. p,p'-Dichlorodiphenyltrichloroethane (p,p'-DDT) and p,p'-dichlorodiphenyldichloroethylene (p,p'-DDE) repress prostate specific antigen levels in human prostate cancer cell lines. *Chem Biol Interact*. 2015; 230:40-9.
44. Emeville E, Giton F, Giusti A, Oliva A, Fiet J, Thomé J, Blanchet P, Multigner L. Persistent organochlorine pollutants with endocrine activity and blood steroid hormone levels in middle-aged men. *PLoS One* 2013;8(6):e66460.
45. Li J, Li N, Ma M, Giesy JP, Wang Z. In vitro profiling of the endocrine disrupting potency of organochlorine pesticides. *Toxicol Lett*. 2008; 183:65-71.

46. Carruba G. Estrogen and prostate cancer: an eclipsed truth in an androgen-dominated scenario. *J Cell Biochem.* 2007; 102:899–11.
47. Ellem SJ, Risbridger GP. Aromatase and regulating the estrogen:androgen ratio in the prostate gland. *J Steroid Biochem Mol Biol.* 2010; 118:246–51.
48. McFarland VA, Clarke JU. Environmental occurrence, abundance, and potential toxicity of polychlorinated biphenyl congeners: considerations for a congener-specific analysis. *Environ Health Perspect.* 1989; 81:225-39.
49. Cussenot O, Azzouzi AR, Nicolaiew N, Fromont G, Mangin P, Cormier L, Fournier G, Valeri A, Larre S, Thibault F, Giordanella JP, Pouchard M, Zheng Y, Hamdy FC, Cox A, Cancel-Tassin G. Combination of polymorphisms from genes related to estrogen metabolism and risk of prostate cancers: The hidden face of estrogens. *J Clin Oncol.* 2007; 25:3596-602.
50. Dobbs RW, Malhotra NR, Greenwald DT, Wang AY, Prins GS, Abern MR. Estrogens and prostate cancer. *Prostate Cancer Prostatic Dis.* 2018; doi: 10.1038/s41391-018-0081-6.

**Table 1****Baseline characteristics of patients and HR (95% CIs) of biochemical recurrence of prostate cancer**

<b>Characteristic</b>	<b>n (%)</b>	<b>HR (95%CI)</b>
Age (years), [mean (interquartile range)]	63.7 (58.5-67.9)	1.01 (0.98-1.04)
Caribbean origin		
French West Indies	313 (96.0)	1.0 (reference)
Haiti or Dominica	13 (4.0)	1.43 (0.58-3.43)
Education		
Primary	165 (60.0)	1.0 (reference)
Secondary	97 (29.9)	0.64 (0.39-1.04)
High school and higher	62 (19.1)	0.98 (0.38-1.20)
Missing data	2	
Body mass index (kg/m <sup>2</sup> )		
< 25	141 (43.3)	1.0 (reference)
25 to 30	154 (47.2)	1.19 (0.76-1.85)
≥ 30	31 (9.5)	2.18 (1.15-4.14)
Smoking		
Never	202 (62.5)	1.0 (reference)
Former or current	121 (37.5)	0.91 (0.59-1.40)
Missing data	3	
Alcohol consumption		
Never	49 (15.1)	1.0 (reference)
Ever	275 (84.9)	1.02 (0.58-1.80)
Missing data	2	
Type 2 diabetes		
No	266 (83.7)	1.0 (reference)
Yes	52 (16.3)	1.03 (0.59-1.79)
Missing data	8	
Family history of prostate cancer		
No	180 (55.6)	1.0 (reference)
Yes	89 (27.5)	1.60 (1.01-2.54)
Do not know	55 (16.9)	1.47 (0.85-2.55)
Missing data	2	
Pre-operative PSA (ng/mL) [median (interquartile range)]	7.3 (5.5-10.5)	1.32 (0.90-1.94)
Total lipids (g/L) [median (interquartile range)]	5.2 (4.6-5.6)	0.96 (0.78-1.18)
Pathological Gleason grade		
ISUP Gleason grade 2 or lower	280 (85.9)	1.0 (reference)
ISUP Gleason grade 3 or higher	46 (14.1)	3.68 (2.36-5.74)
Pathological stage		
pT2 and N0	269 (82.5)	1.0 (reference)
pT3a or pT3b, or N <sup>+</sup>	57 (17.5)	2.90 (1.88-4.49)
Surgical margins		
Negative	233 (71.5)	1.0 (reference)
Positive	93 (28.5)	2.72 (1.80-4.10)
Tumor density (%) [median (interquartile range)]	9.0 (5.0-17.0)	2.62 (1.60-4.29)
Biochemical recurrence status		
No	233 (71.5)	
Yes	93 (28.5)	
Follow-up time (years) [median (interquartile range)]	6.1 (2.8-8.9)	



**Table 2**

**Detection and concentrations of persistent organic pollutants in plasma samples from the study population (µg/L)**

POPs <sup>a</sup>	Detection frequency (%)	Percentile				Maximum
		10th	25th	50th	75th	
Chlordecone	80.1	<LOD	0.16	0.38	0.69	19.1
DDE	94.7	0.38	0.93	2.33	4.68	40.1
DDD	22.1	<LOD	<LOD	<LOD	<LOD	0.95
DDT	28.0	<LOD	<LOD	<LOD	0.06	2.3
PCB-28	47.7	<LOD	<LOD	<LOD	0.24	2.94
PCB-52	50.0	<LD	<LD	0.05	0.39	6.72
PCB-101	45.7	<LOD	<LOD	<LOD	0.12	0.83
PCB-118	57.3	<LOD	<LOD	0.07	0.16	2.38
PCB-138	98.3	0.16	0.29	0.53	0.82	3.60
PCB-153	97.0	0.20	0.38	0.76	1.17	5.96
PCB-180	96.3	0.23	0.35	0.59	0.86	3.16
α-HCH	28.0	<LOD	<LOD	<LOD	<LOD	1.1
β-HCH	35.3	<LOD	<LOD	<LOD	0.05	2.2
γ-HCH	19.5	<LOD	<LOD	<LOD	<LOD	0.60

<sup>a</sup> Chlordecone was measured in 326 plasma samples, and other POPs in 300 plasma samples.

PCB congeners 18, 77, 101, 105, 110, 126, 128, 143, 149, 156, 169, 170, 183, 187, 194, 195, 206, and 209 were below the LOD in all plasma samples.

**Table 3****HR (95% CIs) of biochemical recurrence of prostate cancer according to quartile of chlordecone, DDE and PCB-153 exposure**

<b>Exposure</b>	<b>Without biochemical recurrence (n)</b>	<b>With biochemical recurrence (n)</b>	<b>Crude HR (95% CI)</b>	<b>Adjusted HR (95% CI)<sup>a</sup></b>	<b>Adjusted HR (95% CI)<sup>b</sup></b>
<b>Chlordecone (µg/L)</b>					
< 0.16	62	18	1.0 (reference)	1.0 (reference)	1.0 (reference)
0.16 - 0.37	57	27	1.50 (0.83-2.73)	1.63 (0.89-2.98)	1.55 (0.81-2.93)
0.38 - 0.68	64	18	0.97 (0.50-1.86)	1.03 (0.53-2.00)	0.94 (0.46-1.90)
≥ 0.69	50	30	1.96 (1.09-3.52)	2.51 (1.39-4.56)	2.62 (1.40-4.90)
<i>P</i> trend			0.01	0.002	0.002
<b>DDE (µg/L)</b>					
< 0.93	54	21	1.0 (reference)	1.0 (reference)	1.0 (reference)
0.93 - 2.31	53	21	0.84 (0.46-1.54)	0.54 (0.29-1.01)	0.54 (0.27-1.04)
2.32 - 4.66	48	27	1.18 (0.67-2.09)	0.91 (0.50-1.66)	0.87 (0.45-1.71)
≥ 4.67	59	17	0.71 (0.37-1.35)	0.55 (0.29-1.06)	0.50 (0.24-1.05)
<i>P</i> trend			0.89	0.51	0.35
<b>PCB-153 (µg/L)</b>					
< 0.38	54	20	1.0 (reference)	1.0 (reference)	1.0 (reference)
0.38 - <0.74	51	25	1.28 (0.71-2.30)	1.09 (0.59-1.99)	1.19 (0.62-2.28)
0.75 - 1.16	55	20	0.94 (0.50-1.74)	0.88 (0.47-1.64)	1.05 (0.53-2.06)
≥ 1.17	60	21	1.04 (0.56-1.92)	0.93 (0.49-1.76)	1.12 (0.55-2.28)
<i>P</i> trend			0.81	0.41	0.82

<sup>a</sup> For chlordecone: adjusted for pathological Gleason grade, pathological stage, surgical margins, body mass index, and family history of prostate cancer; For DDE or PCB-153: adjusted for pathological Gleason grade, pathological stage, surgical margins, and body mass index.

<sup>b</sup> For chlordecone: adjusted for pathological Gleason grade, pathological stage, surgical margins, family history of prostate, body mass index, DDE, and PCB-153; For DDE: adjusted for pathological Gleason grade, pathological stage, surgical margins, body mass index, chlordecone, and PCB-153; For PCB-153: adjusted for pathological Gleason grade, pathological stage, surgical margins, body mass index, chlordecone, and DDE.

**Table 4**

**Sensitivity analysis among patients with negative surgical margins, low-grade pathological Gleason score, and organ-confined disease.**  
**HRs (95% CIs) of biochemical recurrence of prostate cancer according to quartile of chlordecone, DDE and PCB-153 exposure**

<b>Exposure</b>	<b>Without biochemical recurrence (n)</b>	<b>With biochemical recurrence (n)</b>	<b>Crude HR (95% CI)</b>	<b>Adjusted HR (95% CI)<sup>a</sup></b>
<b>Chlordecone (µg/L)</b>				
< 0.16	35	6	1.0 (reference)	1.0 (reference)
0.16 - 0.37	43	6	0.87 (0.28-2.70)	0.61 (0.18-2.09)
0.38 - 0.68	42	3	0.44 (0.11-1.75)	0.42 (0.10-1.79)
≥ 0.69	34	15	2.48 (0.96-6.39)	2.98 (1.06-8.38)
<i>P</i> trend			0.008	0.003
<b>DDE (µg/L)</b>				
< 0.93	38	6	1.0 (reference)	1.0 (reference)
0.93 - 2.31	31	7	1.24 (0.42-3.70)	0.99 (0.30-3.32)
2.32 - 4.66	34	8	1.24 (0.43-3.58)	1.32 (0.39-4.48)
≥ 4.67	39	8	1.25 (0.43-3.62)	1.21 (0.36-4.05)
<i>P</i> trend			0.66	0.96
<b>PCB-153 (µg/L)</b>				
< 0.38	42	7	1.0 (reference)	1.0 (reference)
0.38 - <0.74	31	8	1.49 (0.54-4.10)	1.36 (0.48-3.85)
0.75 - 1.16	30	8	1.33 (0.48-3.68)	0.92 (0.29-2.92)
≥ 1.17	33	6	0.98 (0.33-2.93)	0.82 (0.25-2.72)
<i>P</i> trend			0.94	0.89

<sup>a</sup> For chlordecone: adjusted for family history of prostate cancer, body mass index, DDE and PCB-153; For DDE: adjusted for body mass index, chlordecone, and PCB-153; For PCB-153: adjusted for body mass index, chlordecone, and DDE.

Some environmental chemicals can mimic human hormones, and may spur cancer progression. Here, the authors studied the effect of three prevalent organic chemicals on prostate cancer recurrence: the estrogenic insecticide chlordecone; DDE, a stable metabolite of DDT which has anti-androgenic properties; and PCB-153, which has estrogenic and anti-estrogenic properties. No effect was associated with DDE or PCB-153. However, those in the highest exposure quartile for chlordecone had 2-fold increased risk of biochemical recurrence, defined as two consecutive elevated PSA measurements, after prostatectomy. These findings add to the evidence that environmental estrogens could promote cancer progression.