

1 Letter to the Editor Regarding the Article "Rotenone Increases Isoniazid Toxicity but Does Not
2 Cause Significant Liver Injury: Implications for the Hypothesis that Inhibition of the
3 Mitochondrial Electron Transport Chain Is a Common Mechanism of Idiosyncratic Drug-
4 Induced Liver Injury" by Cho and Co-Workers, 2019
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13 To the Editor: Idiosyncratic drug-induced liver injury (iDILI) is a major issue for the treated
14 patients because of its unpredictability and its potential severity. It is also a concern for the
15 pharmaceutical companies since severe or lethal liver injury can lead to the withdrawal of drugs
16 from the market, or earlier during clinical trials, thus causing significant financial losses.¹ While
17 immune response plays a major role,² it is now accepted that mitochondrial dysfunction is also
18 an important mechanism whereby drugs can induce iDILI.^{1,3,4} The unpredictability of drug-
19 induced mitochondrial dysfunction in iDILI can be due to different causes such as drug-drug
20 pharmacokinetic and/or pharmacodynamic interactions, the presence of an underlying liver
21 disease, or different types of genetic predisposition affecting mitochondrial function.^{1,5} Genetic
22 defects affecting the mitochondrial electron transport chain (mtETC) or the mitochondrial fatty
23 acid oxidation (mtFAO) pathway have been reported to favor mitochondrial dysfunction and
24 liver injury induced by several drugs such as the anticonvulsant drug valproic acid and some
25 antiretroviral agents.^{1,5}
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36 In a recent article published in *Chemical Research in Toxicology*, Cho and co-workers
37 treated C57BL/6 mice with rotenone (a prototypical inhibitor of the MRC complex I), isoniazid
38 (an antituberculosis drug inhibiting the MRC complex II), or both compounds, over a six-week
39 period.⁶ Whereas treatment with rotenone alone (0.05 or 0.1% w/w in food) or isoniazid alone
40 (0.2% w/w in food) did not cause death among the animals, the coadministration of rotenone
41 and isoniazid led to lethality in 100% of the mice. This toxicity was not related to liver injury
42 as assessed by hepatic histology and serum glutamate dehydrogenase (GLDH) activity.
43 Notably, the latter investigations were performed after only 3 and 6 days since the cotreated
44 mice did not survive afterwards. From these results, the authors conclude that inhibition of the
45 mtETC is not a significant mechanism of iDILI.⁶ Although this *in vivo* study provides
46 interesting data, this conclusion is somewhat misleading for the following reasons.
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56 First and foremost, there is already strong evidence that mtETC impairment is responsible
57 for liver injury with different drugs, especially with drugs altering the replication or translation
58 of mitochondrial DNA (mtDNA), which encodes for 13 mtETC polypeptides. Of note, such
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3 alteration eventually leads to an impairment of electron transfer from both complexes I and II
4 and onwards of the hepatic mtETC. For instance, phase II clinical trials with the anti-HBV drug
5 fialuridine, which inhibits mtDNA replication,⁷ were prematurely interrupted due to serious
6 adverse effects including unmanageable lactic acidosis, microvesicular steatosis and liver
7 failure requiring liver transplantation, or even leading to death.^{8,9} Similar adverse effects can
8 be induced by different antiretroviral nucleoside reverse-transcriptase inhibitors (NRTIs) such
9 as zalcitabine (ddC), stavudine (d4T) and didanosine (ddI),^{10,11} which also strongly inhibit
10 mtDNA replication.^{7,10} Experimental investigations with fialuridine and NRTIs demonstrated
11 the major role of mtDNA depletion and mtETC impairment in the development of liver
12 injury.^{10,12,13} NRTI-induced hepatotoxicity occurs only in some patients, thus suggesting the
13 role of underlying factors such as genetic predisposition.^{5,14} Furthermore, other experimental
14 investigations support the role of mtETC impairment in liver injury induced by amiodarone,^{15,16}
15 perhexiline¹⁷ and buprenorphine.¹⁸

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Second, in the study of Cho and colleagues, the premature death from extra-hepatic causes
of the mice cotreated with rotenone and isoniazid most probably precluded the possibility to
observe delayed mitochondrial toxicity in liver and subsequent hepatic injury.⁶ Notably, liver
is not the primary target organ during rotenone toxicity.¹⁹ Human and animals studies showed
that rotenone-induced mitochondrial toxicity causes respiratory depression, cardiovascular
collapse and severe metabolic (i.e. lactic) acidosis.¹⁹ In addition to heart and lungs, kidney and
spleen are also damaged after rotenone exposure.²⁰ Regarding isoniazid, although
hepatotoxicity is a significant adverse event in treated patients, hypotension, renal failure and
metabolic acidosis could also occur.^{21,22} A thorough necropsy of the deceased mice might have
uncovered which vital organs were severely damaged by rotenone. Moreover, *in vitro*
investigations in hepatocytes might have permitted to evaluate the toxicity of rotenone,
isoniazid and their combination. Indeed, previous investigations reported that rotenone and
isoniazid could induce toxicity in hepatic cells.^{23,24}

Third, the authors looked for hepatic injury in liver sections by using hematoxylin and eosin
staining. However, this staining method is not appropriate in order to detect microvesicular
steatosis,²⁵ a liver lesion commonly occurring with drugs and toxins inducing mitochondrial
dysfunction.^{7,26} Regardless of the pathophysiological context, minor or moderate
microvesicular steatosis is better detected with Oil red O or Sudan III staining on frozen liver
section.^{25,27}

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3 Investigations in animals are useful in order to study DILI including iDILI, but different
4 factors may greatly modulate the severity of liver injury such as drug distribution and
5 metabolism as well as animal species and strain.^{1,28,29} Interestingly, isoniazid did not induce
6 microvesicular steatosis in C57BL/6J mice, whereas this liver lesion was observed in other
7 strains such as BALB/cJ, DBA/2J and LG/J mice.³⁰ The latter study and others^{1,14,31,32} underline
8 the importance of genetic susceptibility in the occurrence of drug-induced liver injury, in
9 particular when considering genes encoding for mitochondrial proteins. Importantly, drug-
10 induced mtETC toxicity can cause not only microvesicular steatosis and cell death but also
11 other liver lesions such as steatohepatitis and cholestasis.^{5,17,33} Hence, more investigations are
12 clearly needed in order to identify the main factors able to modulate drug-induced
13 mitochondrial dysfunction in liver. While *in vivo* investigations may better reflect the
14 complexity of iDILI, complementary *in vitro* experiments are also useful to decipher the
15 mechanisms whereby drugs can be toxic for mitochondria.
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